



# Combining ability and heterosis analysis for drought tolerant traits in rice (Oryza sativa L.)

# V. Karpagam\*1, S. Jebaraj2 and S. Rajeswari3

<sup>1</sup>Department of Plant Breeding and Genetics, Vanavarayar Institute of Agriculture, Pollachi- 642103 (Tamil Nadu), INDIA

<sup>2</sup>Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Madurai- 625104 (Tamil Nadu), INDIA

<sup>3</sup>Centre for Plant Breeding and Genetics, Tamil Nadu Agricultural University, Coimbatore – 641003 (Tamil Nadu), INDIA

\*Corresponding author. E-mail: priyatnau2007@yahoo.co.in

Received: August 5, 2015; Revised received: January 21, 2016; Accepted: April 30, 2016

**Abstract:** Rice is the most important staple food for more than half of the world's population and also for most of the countries. A Line x Tester analysis was undertaken to study the nature of gene action for yield and drought tolerant traits. The ratio of SCA and GCA was less than unity for all the characters which revealed that the preponderance of non- additive gene action governing the traits concerned. The lines *viz.*, ADT 43, ADT (R) 49, CO (R) 50 and the testers *viz.*, PMK (R) 3, Chandikar and Anna (R) 4 were adjudged as the best general combiners for drought tolerant traits. The cross combinations *viz.*, ADT 39 x Vellaichitraikar had exhibited significant values for dry root weight (9.66), root/shoot ratio (0.31), root length (3.82), number of roots per plant (37.08), root thickness (0.11), root volume (4.27) and root length density (0.03) ADT (R) 49 x Chandikar for 70 percent relative water content (8.85), dry root weight (18.03), dry shoot weight (40.55), root length (3.10), number of roots per plant (140.16) root thickness (0.38) and root volume (23.14) were found to be specific combiners for most of the drought tolerant traits. The cross combinations, *viz.*, ADT 43 x Anna (R) 4, ADT (R) 49 x Chandikar and ADT 43 x PMK (R) 3 had highly significant standard heterosis. Breeding for drought tolerance in rice would be of immense value to the farmers economic health, family well-being and harmony in the society.

Keywords: Combining, Drought, GCA, Heterosis and SCA

#### INTRODUCTION

Rice is the most important food source for more than half of the world's population and also main staple food for most of the countries which comprises of 86 million people (Luu et al., 2012). Globally rice is cultivated under 158.4 million hectares with an annual production of around 697.2 million tonnes and an average productivity of 2.85 t/ha. Climate change threatens the sustainability of modern-day agriculture. Constantly changing climatic conditions around the world demand constant efforts to understand and adapt to environmental challenges for sustainable crop production (Dixit and Kumar, 2014). Land races are one of the important components of the germplasm and serve as the donors for the drought tolerance. Local land races are naturally adapted to utilize the natural resource-base better than the introduced modern cultivars (Bhattacharya and Ghosh 2004). Moreover the land races have broad genetic base, which provides wider adaptability and protection from various biotic and abiotic stresses. Hence, we could develop the high yielding hybrids with an added advantage of drought tolerance by crossing the drought tolerant land races with high yielding varieties, which are susceptible to drought (Muthuramu *et. al.*, 2010).

The concept of combining ability helps the breeder to determine the nature of gene action involved in the expression of quantitative traits of economic importance. The choice of suitable breeding method for the improvement of drought tolerant traits primarily depends on the relative importance of GCA and SCA variances. Proper choice of parents on the basis of their combining ability status for putative drought tolerant attributes as well as productive traits and selection in typical target environment will help in combining complex traits such as productivity and drought tolerance (Hanamaratti et al., 2004). Heterosis for drought tolerant traits will be a boon to drought tolerance breeding since most of the hybrids developed so far lack tolerance to abiotic and biotic stresses (Muthuramu et. al., 2010). The drought resistant variety is expected to possess the ability to withstand drought stress. Breeding for drought tolerance in rice has immense value to the farmers economic health, family well-being and harmony in the society. Besides

providing information on nature and magnitude of gene action governing yield and yield attributes, combining ability analysis also helps in the identification of the potential parents, cross combinations and formulation of systematic breeding plan for augmenting grain yield. The present study was undertaken to estimate the nature and magnitude of gene action governing important quantitative and drought traits of rice, Oryza sativa under drought situation and to estimate the general combining ability of parents, specific combining ability of hybrids and heterosis.

#### MATERIALS AND METHODS

The genetic materials consisted of ten lines, viz., ADT 36, ADT 39, ADT 43, ADT (R) 49, ASD 16, BPT 5204, CO 47, CO(R) 49, CO (R) 50 and IR 50 (high yielding cosmopolitan rice varieties) and six testers viz., Anna (R) 4, Chandikar, Chinnar 20, Nootripathu, PMK(R) 3 and Vellaichitraikar (drought tolerant genotypes), which were crossed in a Line x Tester design and CO (R) 50 was used as check variety. Seeds from the 60 cross combinations along with their parents were sown in raised nursery beds during Kharif, 2013. Twenty two days old seedlings of 60 crosses along with parents were transplanted under moisture stress condition in a Randomized Block Design replicated twice adopting a spacing of 25cm between rows and 20cm between plants within a row for studying combining ability and heterosis. Single seedling was transplanted per hill in two rows of three meter row length for each cross combination and in each replication. For estimating heterosis, the seedlings of parents were also transplanted on 22<sup>nd</sup> day of sowing in an adjacent plot with two replications adopting the same spacing as that of hybrids in rows three metre length. The recommended package of practices and plant protection measures were followed to obtain good standing of the crop. The stress was imposed at active tillering stage. Irrigation was stopped on 70<sup>th</sup> day after sowing and the stress was imposed for 15 days. Observations were recorded individually on five random plants in each replication for each hybrid and parents for estimating combining ability and heterosis. The mean values recorded for biometrical characters in the parents and F<sub>1</sub> generations were used for statistical analysis. The analysis was done using the TNAUSTAT statistical package.

Analysis of variance of ten lines, six testers and 60 hybrids were carried out for the quantitative characters following the procedure outlined by Panse and Sukhatme (1964). Combining ability analysis was carried out according to the methodology of Kempthorne (1957). The general combining ability of the parents and specific combining ability of the hybrids were assessed. The expected mean squares due to the different sources of variation and their genetic expectations were calculated as indicated in the following ANOVA table.

Where, r = number of replications, l = number of lines, t = number of testers

Estimates of covariance of full sibs and half sibs were calculated from the genetic expectations of mean squares as:

 $Cov (F.S.) = (M_1 - M_4) + (M_2 - M_4) + (M_3 - M_4) + 6r Cov$ (H.S.) - r(1 + t) Cov (H.S.) / 3r

Cov (H.S.) =  $(M_1-M_3) + (M_2-M_4) / r (1+t)$ 

Where, Cov (F.S.) = Covariance between full sibs, Cov (H.S.) = Covariance between half sibs.

From the covariance of full sibs and covariance of half sibs, variance due to general combining ability (GCA) and specific combining ability (SCA) were estimated as follows:

 $6^2$  GCA = Cov. H.S.

 $6^2$  SCA = Cov. F.S. - 2 Cov. H.S.

Estimation of GCA and SCA effects: The GCA and SCA effects of parents and hybrids respectively were estimated based on the following model.

$$\begin{split} X_{ijk} &= \mu + g_{i.} + g_{.j} + s_{ij} + e_{ijk} \\ \text{Where, } X_{ijk} &= \text{value of the } ijk^{th} \text{ observation, } \mu = \text{population mean, } g_{.i} = \textit{gca} \text{ effect of } i^{th} \text{ line, } g_{.j} = \textit{gca} \text{ effect of the } j^{th} \text{ tester, } S_{ij} = \textit{sca} \text{ effect of the } ij^{th} \text{ hybrid, } e_{ijk} = \text{constant} \text{ the } ij^{th} \text{ line, } ij^{th} \text{ line,$$
error effect associated with ijkth observation, i = number of lines, j = number of testers, k = number of replications. The individual effects of GCA and SCA were obtained from the two way table of lines versus testers, in which each figure was a total over replications.

And was calculated as follows:

$$\mu = X.../ rlt$$

GCA effects of lines,  $g_i = \frac{x_i}{rt} - \frac{x_{...}}{rlt}$ 

GCA effects of testers,  $g_j = \frac{x.j.}{rl} - \frac{x...}{rlt}$ SCA effects of hybrids,  $S_{ij} = \frac{xij.}{r} - \frac{xi...}{rt} - \frac{x.j.}{rl} + \frac{x...}{rlt}$ Where, Where,

X... = total of all hybrid combinations;  $X_{i..}$  = total of  $i^{th}$  line over '1' testers and 'r' replications;  $X_{ij}$ . = total hybrids between  $i^{th}$  line and  $j^{th}$  tester over replications;  $X_{\cdot j}$ . = total of  $j^{th}$  tester over '1' lines and 'r' replications

The standard errors pertaining to GCA and SCA effects were calculated as given below:

Standard error for testing the GCA effects of lines  $SE(g_i) = [EMS/rt]^{1/2}$ 

Standard error for testing the GCA effects of testers  $SE(g_j) = [EMS/rl]^{1/2}$ 

Standard error for testing the SCA effects of hybrids  $SE(s_{ii}) = [EMS/r]^{1/2}$ 

Test of significance:

For lines,  $\mathbf{t}_{gi} = gi / \text{SE}(gi)$ For testers,  $\mathbf{t}_{gj} = gj / \text{SE}(gj)$ For hybrids,  $\mathbf{t}_{sij} = sij / \text{SE}(sij)$ 

Estimation of heterosis: Heterosis was estimated from the overall mean of each hybrid for each trait. Standard heterosis (diii) for each character was expressed as per cent increase or decrease of F<sub>1</sub> value

**ANOVA Table** 

Source	Degrees of Freedom	Mean squares	Expected mean squares
Replication	(r - 1)	-	
Hybrids	(lt-1)	-	
Lines	(1-1)	$M_1$	EMS + r [Cov (F.S.) - 2 Cov (H.S.)] + rt [Cov (H.S.)]
Testers	(t-1)	$M_2$	EMS + r [(Cov (F.S.) - 2 Cov (H.S.)] + rl [Cov (H.S.)]
Lines x Testers	(1-1)(t-1)	$M_3$	EMS +r $[Cov(F.S.) - 2 Cov(H.S.)]$
Error	(r-t)(lt-1)	$M_4$	EMS
Total	rtl - 1		

over the standard variety (SV).

Standard heterosis (diii)

diii 
$$= \overline{F1} - \overline{SV} \times 100$$
Where,

 $\overline{F1}$  = Average performance of the hybrid

 $\overline{SV}$  = Average performance of standard variety

Significance of heterosis was tested using the formula given by Snedecor and Cochran (1967).

 $SE_{(diii)} = \sqrt{2EMS/r}$ 

Where, EMS = error mean square, obtained from analysis of variance.

r = number of replications

 $t (cal) = d_{iii}/SEd_{iii}$ 

The calculated 't' value was tested against table't' value at error degrees of freedom for five and one per cent probability levels.

### RESULTS AND DISCUSSION

In the estimate of variances, the mean squares due to lines, testers, hybrids and line x tester interactions for different drought tolerant traits are presented in Table 1. In the present investigation, results showed that these ratios were less than unity for all the characters and greater proportion of SCA variance was observed for all the characters studied which revealed that the preponderance of non- additive gene action governing the traits concerned. This showed the presence of greater variance of non-additive gene action for all the characters, which offer scope for exploitation of hybrid vigour through heterosis breeding and selection procedure in late or advanced generation will be very important to improve these traits. These results were in agreement with those reported by Hosseini et al. (2005) and Malarvizhi et al. (2010) for spad chlorophyll meter reading and dry shoot weight in rice, Gnanasekaran et al. (2006) for biomass yield in rice, Das et al. (2005) for relative water content in rice; Ganesh et al. (2004), Sathya and Jebaraj (2013) and Utharasu and Anandakumar (2013) for dry root weight and root/shoot ratio in rice.

The GCA effects represent the nature of gene action. A best general combiner is characterized by its better breeding value when crossed with number of other parents. Dhillon (1975) pointed out that combining ability of parents gives useful information on the choice of parents in terms of expected performance of their progenies. The general combining ability effects

of parents are furnished in Table 2 for different drought tolerant traits. The lines *viz.*, ADT 43, ADT (R) 49, BPT 5204 and CO 47 were adjudged as the best general combiners for yield and drought tolerant traits. Based on GCA performance, the testers *viz.*, PMK (R) 3, Chandikar and Anna (R) 4 were considered as good general combiners for drought component traits and crosses involving them would result in the identification of superior segregants with favourable genes for yield and drought tolerant traits in rice This is in accordance with the earlier findings of Manonmani and Fazhullah Khan (2005) for relative water content and root length in rice.

SCA is defined as the deviation from the mean performance, predicted on the basis of GCA (Allard, 1960) and specific combining ability is due to non-additive genetic interaction (Sprague and Tatum, 1942). The SCA effects of 60 hybrids are presented in Table 4 for drought tolerant related traits. The cross combinations viz., ADT 39 x Vellaichitraikar had exhibited significant values for dry root weight (9.66), root/shoot ratio (0.31), root length (3.82), number of roots per plant (37.08), root thickness (0.11), root volume (4.27) and root length density (0.03) ADT (R) 49 x Chandikar for 70 percent relative water content (8.85), dry root weight (18.03), dry shoot weight (40.55), root length (3.10), number of roots per plant (140.16) root thickness (0.38) and root volume (23.14) were found to be specific combiners for most of the drought tolerant traits. The hybrids viz., ADT (R) 49 x Chinnar 20, ADT 43 x Anna (R) 4, ADT 36 x Anna (R) 4, BPT 5204 x Chinnar 20, CO (R) 50 x Nootripathu and CO (R) 50 x PMK (R) 3 were also found to be specific combiners. Similar results were obtained by Sathya and Jebaraj (2013) for the traits 70 percent relative water content, dry root weight, dry shoot weight and root length in rice. For these combinations, since they involved non-additive gene action, cyclic method of breeding involving selection of desired recombinants and their inter crossing would be more desirable (Muthuramu et al., 2010).

In the estimation of heterosis, The cross combination, ADT 43 x Anna (R) 4 ( $L_3$  x  $T_1$ ) had significantly high values for spad chlorophyll meter reading (16.63), dry root weight, (44.64), dry shoot weight (42.68), root length (24.30), number of roots per plant (78.14) root thickness (42.38) and root volume (44.37) (Table 5). The hybrid, ADT (R) 49 x Chandikar ( $L_4$  x  $T_2$ ) observed significant and high standard heterosis value for dry root weight (81.96), dry shoot weight (50.77),

Table 1. Analysis of variance for combining ability for different drought tolerant traits.

		Mean squares									
Source of variation	df	SPAD chlorophyll meter reading	70% relative water content	Dry root weight	Dry shoot weight	Root/shoot ratio	Root length	No. of roots per plant	Root thickness	Root	Root length density
Replication	-	0.5562	39.68	0.04	0.05	0.032	0.71	91.87	0.008	1.77	0.0009
Hybrids	59	20.95**	65.71**	257.25**	1506.81**	0.07**	20.54**	11979.94**	0.12*	276.79**	0.25*
Lines	6	22.24**	63.46**	324.90**	1388.36**	**90.0	38.58**	12425.32**	0.24**	291.11**	0.32**
Testers	S	36.30**	61.33**	386.99**	1755.05**	0.046**	28.68**	10843.87**	0.18*	320.47**	0.22*
LXT	45	18.99**	- <del>X-</del>	229.30**	1502.92**	0.07**	16.02**	12017.09**	**60.0	269.08**	0.25*
Error	59	0.90	20.39	1.66	2.15	0.01	0.19	58.18	0.02	09.0	0.02
$\sigma^2$ GCA		0.03		0.56	0.07	0.01	60.0	0.75	0.0006	0.15	0.02
$\sigma^2$ SCA		9.04		113.82	750.38	0.03	7.91	5979.45	0.04	134.23	0.12
$\sigma^2$ GCA / SCA		0.004	0.0008	0.004	0.0001	0.29	0.01	0.0001	0.01	0.001	0.17

\*Significant at 5% level, \*\*Significant at 1% level.

Table 2. General combining ability effects of lines and testers for different drought tolerant traits.

Parents		SPAD chlorophyll meter reading	70% relative water content	Dry root weight	Dry shoot weight	Root/ shoot ratio	Root length	No. of roots per plant	Root thick- ness	Root volume	Root length density
Lines											
ADT 36	L	1.16**	-1.02	-0.29	5.13**	-0.02*	-1.63**	-28.34**	-0.19**	-3.65**	-0.04**
ADT 39	$\Gamma_2$	-2.11**	-4.18**	-1.56**	-6.53**	**80.0	-1.12**	9.32**	-0.17**	-0.12	-0.13**
ADT 43	Ļ	1.76**	-1.81	7.52**	18.07**	0.02	-0.45**	46.58**	0.07**	2.93**	-0.12**
ADT (R) 49	ľ	-1.72**	0.21	5.81**	8.43**	0.03**	0.82**	57.24**	0.19**	7.30**	-0.15**
ASD 16	Ļ	1.65**	-1.17	**00.6-	-15.46**	-0.10**	0.25	-34.51**	0.02	-5.21**	0.22**
BPT 5204	Ľ	-0.04	0.64	-2.62**	7.97**	***/0-0-	0.85**	**89'8-	0.10**	3.82**	-0.10**
CO 47	$\Gamma_7$	-1.05**	0.35	2.47**	**99.6-	0.14**	-1.66**	10.16**	**80.0	0.23	-0.16**
CO (R) 49	٦̈́	-0.35	0.25	5.03**	4.30**	0.04**	-0.31*	-2.01	-0.1**	-2.73**	0.03*
CO (R) 50	J	-0.21	4.12**	-2.14**	0.42	**80.0-	4.39**	-11.26**	0.17**	5.44**	0.24**
IR 50	$L_{10}$	0.91**	2.61*	-5.21**	-12.67**	-0.04**	-1.13**	-38.51**	-0.15**	-8.01**	0.21**
Testers											
Anna (R)4	$T_1$	2.20**	-1.23	-2.18**	2.73**	-0.05**	0.55**	26.16**	-0.01	-0.35	0.13**
Chandikar	$T_2$	-0.02	0.27	1.61**	2.87**	0.01	0.78**	12.51**	0.04**	4.76**	**60.0-
Chinnar 20	$T_3$	-2.01**	1.24	1.06**	-6.70**	**90.0	-0.23*	-16.79**	0.08**	-0.37*	0.04**
Nootripathu	$T_4$	0.19	-1.83	-5.43**	-8.23**	**90.0-	-2.22**	-21.94**	-0.10**	-4.23**	-0.02
PMK(R) 3	$T_5$	-0.36	-1.18	7.30**	16.22**	0.04**	1.13**	23.56**	0.11**	4.53**	-0.15**
Vellaichitraikar	$T_6$	0.01	2.73**	-2.35**	**68'9-	0.01	0.02	-23.49**	-0.13**	-4.34**	0.10

\*Significant at 5% level, \*\*Significant at 1% level.

Table 3. Specific combining ability of hybrids for different drought tolerant traits.

'n	Hybrids	Spad	70% rela-	Dry root	Dry snoot	Koot/shoot	K00t	No. 01 roots	Koot thick-	K001 V0I-	1004
ż		chlorophyll	tive water	weight	weight	ratio	length	per plant	ness	nme	length
		meter reading	content								density
  -	$L_1 \times T_1$	2.43**	-2.29	6.23**	16.27**	-0.01	4.91**	18.09**	-0.05	7.55**	-0.16**
2	$L_1 \times T_2$	-1.86**	-5.87	4.40**	-24.71**	0.27**	0.20	-76.76**	0.05	-10.30**	0.42**
3	$L_1 \times T_3$	-0.43	-9.64**	89.0-	-17.01**	0.11**	-0.27	-45.96**	90.0	-6.17**	0.26**
4	$L_1 \times T_4$	-4.06**	1.42	-4.46**	4.99**	-0.12**	-0.60	32.69**	0.03	9.34**	-0.31**
5	$L_1 \times T_5$	4.05**	6.85*	-3.63**	22.22**	-0.17**	-0.81*	11.19*	0.05	-0.92	0.01**
9	$L_1 \times T_6$	-0.12	9.53*	-1.85*	-1.76	***/0'0-	-3.43**	60.74**	-0.15**	0.50	-0.20**
7	$L_2 \times T_1$	-0.40	8.16*	×*69°L	41.17**	-0.15**	0.82**	-42.08**	0.17	1.62**	-0.16**
8	$L_2 \times T_2$	1.65*	-5.52	3.99**	19.04**	-0.13**	-0.26	71.07**	0.07	6.62**	-0.15**
6	$L_2 \times T_3$	68.0	2.99	-11.29**	-5.17**	-0.28**	1.91**	17.88**	*60.0-	-9.11**	0.51**
10	$L_2 \times T_4$	-0.65	-1.40	-2.83**	2.88**	-0.16	-1.80**	-32.47**	0.02	1.50**	-0.17**
11	$L_2 \times T_5$	69.0	-2.72	-7.22**	-46.90**	0.40**	-4.47**	-51.47**	-0.28**	-4.90**	-0.05**
12	$L_2 \times T_6$	-2.18**	-1.51	**99.6	-11.01**	0.31**	3.82**	37.08**	0.11**	4.27**	0.03**
13	$L_3x T_1$	2.73**	-0.36	10.38**	25.08**	0.01	5.18**	163.68**	0.42**	21.02**	-0.37**
4	$L_3 \times T_2$	3.82**	-1.73	-18.44**	-47.13**	-0.04	-4.95**	-137.18**	-0.07	-16.74**	0.17**
15	$L_3 \times T_3$	-0.40	-2.06	-3.93**	23.26**	-0.18**	0.33	-45.88**	-0.05	-3.75**	-0.04**
16	$L_3 \times T_4$	1.16	1.20	16.28**	28.32**	**90.0	0.52	23.77**	-0.05	8.40**	-0.24**
17	$L_3 \times T_5$	0.61	4.07	-4.29**	-50.02**	0.29**	-0.44	28.77**	*80.0	1.35*	-0.05**
18	$L_3 \times T_6$	-7.93**	-1.12	-0.01	20.49**	-0.12**	-0.64*	-33.17**	-0.33**	-10.28**	0.52**
19	$L_4 \times T_1$	-1.47*	-1.24	-0.91	-27.96**	0.21**	-4.80**	20.51**	-0.36**	-16.85**	0.13**
20	$L_4 \times T_2$	0.20	8.85**	18.03**	40.55**	0.02	3.10**	140.16**	0.38**	23.74**	-0.22**
21	$L_4 \times T_3$	-3.04**	4.42	15.74**	3.34**	0.21**	1.09**	69.46**	*80.0	15.12**	-0.31**
22	$L_4 \times T_4$	-1.76*	3.93	-10.73**	-17.19**	-0.10**	-0.82**	-34.39**	0.17**	-1.52**	-0.11**
23	$L_4 \times T_5$	2.21**	-2.43	-22.21**	-47.86**	-0.12**	0.59	-151.89**	0.07	-16.08**	0.46**
24	$L_4 \times T_6$	3.86**	-13.53**	60.0	49.13**	-0.20**	0.83**	-43.84**	-0.33**	-4.41**	0.04**
25	$L_5x T_1$	-4.35**	2.28	0.85	-2.87**	0.02	3.88**	61.76**	0.31**	16.01**	-0.55**
26	$L_5 \times T_2$	2.35**	0.05	-0.92	-10.78**	**/0.0	-4.58**	**60.89-	-0.10*	-9.04**	-0.05**
27	$L_5 \times T_3$	-1.47*	-6.27	-5.07**	3.22**	-0.13**	-2.39**	18.71**	-0.29**	-5.06**	0.01**
28	$L_5 \times T_4$	0.54	-0.93	5.72**	-2.21*	0.16**	1.55**	-44.64**	0.11**	-1.45**	0.27**
29	$L_5x T_5$	-0.79	-2.80	3.94**	25.50**	**90.0-	1.28**	18.86**	*80.0-	2.44**	-0.14**
30	LxT	3 72**	*191	**1	10 00 **	**700	30.0	10 41**	***	***	****

ontd.

Table 3. Contd......

S**	**6	**9	* *	*	*	**0	*	*	*	*	<b>5</b> **	*	<b>5</b> **	* *	2**	* *	3**	**	***	3**	*	**	2**	*	*	*	7**	**	4**
	•	•	Ū	•	•	•	_	•	•	_	•	Ţ	•	_	-0.15**	_	•		·	•	_	•	•	_	_	_	•	•	
-1.17**	11.07**	16.90**	-11.49**	**66.6-	-5.33**	6.87	-9.84**	7.19**	-5.95**	-5.80	7.51**	-10.67**	6.82**	**06.9-	7.16**	-1.54**	5.12**	-20.54**	1.26**	-5.76**	-9.50**	32.29**	2.26**	-3.84**	-3.59**	-2.46**	3.50**	3.14**	3.26**
-0.05	-0.20**	0.11**	-0.01	-0.07	0.22**	-0.03	*80.0-	0.14**	-0.03	0.10**	-0.11**	-0.45**	0.42**	*80.0-	-0.07	0.03	0.15**	*60.0	-0.10*	-0.06	0.18**	-0.19**	*80.0	-0.07	-0.36**	0.17**	-0.33**	0.27**	0.32**
-29.57**	43.08**	76.88**	-31.47**	-11.97**	-46.92**	61.59**	-37.26**	17.04**	-19.81**	-65.31**	43.74**	-65.74**	33.91**	-62.79**	72.36**	-8.14	30.41**	-126.49**	-41.34**	19.46**	-27.89**	231.61**	-55.34**	-61.74**	72.41**	-64.79**	61.86**	-1.64	-600
0.35	0.85	0.59	-3.95**	4.16**	-1.99**	-0.32	3.41**	-2.26**	0.19	-2.81**	1.79**	-4.56**	0.11	-0.83**	2.67**	**06.0	1.71**	-4.50**	*6.0-	1.52**	2.43**	2.72**	-1.51**	-0.95**	2.77**	0.33	-0.19	-1.12**	-0.83**
-0.03	-0.10**	-0.02	0.31**	-0.07**	**80.0-	0.29**	-0.20**	0.30**	-0.14**	-0.25**	0.01	-0.21**	-0.10**	**/0.0	**80.0-	-0.02	0.33**	-0.16**	**90.0-	0.01	0.11**	**80.0	0.04	0.05*	0.30**	-0.07**	-0.04	-0.07**	-0.16**
-8.24**	41.23**	23.24**	-45.60**	-7.62**	-3.00**	-10.76**	21.46**	-4.49**	-6.31**	7.04**	-6.94	-3.50**	-25.76**	-13.77**	25.43**	20.67**	-3.08**	-4.78**	-11.02**	-2.61*	6.63**	45.04**	-33.27**	-24.42**	-2.89**	-10.02**	3.06**	31.95**	2.32*
-2.08*	4.45**	9.71**	-6.16**	-3.97**	-1.95*	**99.6	-1.32	8.61**	-7.24**	-7.44**	-2.28*	-14.11**	-14.53**	-3.53**	2.96**	12.07**	17.14**	-11.86**	-8.14**	-2.17**	7.00	25.57**	-10.40**	-5.86**	12.49**	-7.40**	-0.54	7.18**	-5.88**
0.22	-1.60	0.57	-0.42	-1.21	2.45	0.10	-2.93	-4.36	-0.36	1.07	6.49*	1.21	**49.6	3.26	-5.27	-4.15	-4.72	-3.37	2.00	-5.45	2.39	4.51	-0.07	-4.71	-2.91	16.54**	-0.55	-3.20	-5.17
0.04	-0.50	1.84**	1.43*	-2.24**	-0.57	-1.59*	0.41	-2.57**	-0.81	-2.41**	**86.9	4.05**	-2.97**	-2.44**	09.0	0.78	-0.02	-0.61	-0.14	3.11**	2.67**	66.0	-6.01**	-0.83	-2.97**	4.52**	06.0	-3.90**	2.27**
$L_6 \times T_1$	$L_6 \times T_2$	$L_6 \times T_3$	$L_6 \times T_4$	$L_6x T_5$	$L_6 x T_6$	$L_7 \times T_1$	$L_7 \times T_2$	$L_7 \times T_3$	$L_7 \times T_4$	$L_7 \times T_5$	$L_7 \times T_6$	$L_8 \times T_1$	$L_8 \times T_2$	$L_8 \times T_3$	$L_8 \times T_4$	$L_8 \times T_5$	$L_8 \times T_6$	$L_9x T_1$	$L_9 \times T_2$	$L_9 \times T_3$	$L_9 \times T_4$	$L_9 \times T_5$	$L_9 \times T_6$	$L_{10} \times T_1$	$L_{10}x T_2$	$L_{10}x T_3$	$L_{10}x$ $T_4$	$L_{10}x T_5$	$L_{10}x T_6$
31	32	33	34	35	36	37	38	39	40	41	42	43	4	45	46	47	48	49	20	51	52	53	54	55	99	27	28	59	09

\*Significant at 5% level, \*\*Significant at 1% level.

Table 4. Standard heterosis for different drought tolerant traits.

ngth		*	*						*									*		*							*	Contd.
Root length density	42.00**	114.00*	108.00*	-16.00	19.00	28.00*	26.00*	-17.00	141.00*	-4.00	-8.00	57.00**	-14.00	**00.05	34.00**	-16.00	-6.00	158.00*	**00.08	-34.00*	-27.00*	3.00	91.00**	**00'95	17.00	73.00**	111.00*	$C_{\mathcal{C}}$
Root volume	-19.18**	-59.59**	-62.76**	-25.83**	-30.59**	-54.20**	-26.78**	5.23*	**98.09-	-39.46**	-32.01**	-31.06**	44.37**	-59.11**	-34.23**	-7.92**	-2.54	-67.51**	-61.81**	83.04**	39.46**	-25.52**	-43.90**	-35.02**	2.69	-60.54**	-64.18**	
Root thick- ness	-27.14**	-13.33*	-7.62	-27.62**	-5.71	-48.10**	-4.76	-10.48**	-20.95**	-28.10**	-36.19**	-21.90**	42.38**	0.10	6.19	-10.95**	20.95**	-40.48**	-21.43**	53.33**	29.52**	20.48**	31.43**	-30.00**	25.24**	-9.05	-22.86**	
No. of roots per plant	-7.80**	-50.10**	-49.51**	-20.86**	-11.50**	-10.53**	-16.57**	22.22**	-9.94**	-31.58**	-21.25**	-5.07	78.17**	-44.44**	-20.27**	4.87	24.56**	-17.93**	26.51**	67.84**	28.85**	-13.65**	-41.72**	-17.93**	6.82*	-49.12**	-26.71**	
Root length	15.00**	-13.42**	-22.8**	-37.52**	-17.54**	-41.42**	-7.71*	-13.10**	-5.71	-41.86**	-37.52**	7.83*	24.20**	-38.57**	-11.51**	-22.93**	-7.71*	-16.18**	-31.08**	20.52**	1.36	-23.34**	*88.9	1.17	20.39**	-31.81**	-24.33**	
Root/shoot ratio	-12.86	84.29**	54.29**	-45.71**	-28.57**	-11.43	-22.86**	-1.43	-30.00**	-27.14**	162.86**	127.14**	1.43	7.14	-18.57*	18.57*	112.86**	-14.29	65.71**	21.43*	98.57**	-22.86**	0.14	-32.86**	-24.29**	7.14	-37.14**	
Dry shoot weight	13.21**	-42.12**	-44.67**	-16.92**	39.55**	-24.25**	31.14**	1.36	-44.42**	-35.59**	-69.92**	-52.59**	42.68**	-54.97**	27.44**	32.22**	-40.81**	23.42**	-42.26**	50.77**	-12.63**	-42.51**	-50.95**	49.17**	-40.61**	-51.13**	-45.14**	
Dry root weight	-1.36	6.17	-15.42**	-54.92**	-2.80	-33.02**	-0.60	-0.25	-61.06**	-53.52**	-21.48**	6.32	44.64**	-51.54**	2.13	54.82**	24.70**	4.07	-5.36	81.96**	71.11**	-55.57**	-50.77**	-2.13	-55.47**	-47.69**	-65.71**	
70% relative water content	-42.18**	-47.73**	-55.15**	-33.92**	-17.73	-0.21	-22.81	-55.22**	-30.00*	-49.86**	-51.62**	-37.99**	-39.16**	-38.81**	-37.12**	-36.61**	-27.23*	-30.65*	-36.13**	-5.28	-14.47	-23.97*	-39.15**	-58.29**	-30.44*	-32.39**	-46.61**	
Spad chlorophyll me- ter reading	14.48**	-1.04	-2.38	-5.76*	12.23**	3.17	-0.07	-0.48	-7.02**	-5.44*	-3.57	-9.52**	16.63**	13.93**	-0.87	8.10**	5.48*	-13.98**	-1.67	-2.98	-15.44**	-7.14**	1.00	5.80*	-0.51	10.15**	-3.69	
Hybrids	$L_1 \times T_1$	$L_1 \times T_2$	$L_1 \times T_3$	$L_1 \times T_4$	$L_1 \times T_5$	$L_1 \times T_6$	$L_2 \times T_1$	$L_2 \times T_2$	$L_2 \times T_3$	$L_2 \times T_4$	$L_2 \times T_5$	$L_2 \times T_6$	$L_3x T_1$	$L_3 \times T_2$	$L_3 \times T_3$	$L_3 \times T_4$	$L_3 \times T_5$	$L_3 \times T_6$	$L_4 \times T_1$	$L_4 \times T_2$	$L_4 \times T_3$	$L_4 \times T_4$	$L_4 \times T_5$	$L_4 \times T_6$	$L_5x T_1$	$L_5 \times T_2$	$L_5 \times T_3$	
s z	_	7	$\epsilon$	4	5	9	7	~	6	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	56	27	

 $L_5 \times T_4$	6.35**	-40.57**	-49.25**	-54.57**	12.86	-11.96**	-53.41**	-2.38	-64.98**	152.00**
$L_5x T_5$	1.86	-43.82**	-7.18	16.10**	-20.00*	7.61*	-10.92**	0.23	-24.88**	43.00**
$L_5 \times T_6$	13.49**	-5.57	-16.66**	-67.21**	-28.57**	-6.12*	-31.38**	-12.38*	**68.69-	211.00**
$L_6 \times T_1$	5.92*	-31.10**	-42.18**	-16.14**	-31.43**	1.81	-18.71**	0.36	-23.14**	33.00**
$L_6 \times T_2$	-0.67	-31.97**	-2.55	51.08**	-34.29**	6.47*	4.29	-9.52	31.85**	-19.00
$L_6 \times T_3$	0.15	-23.60*	15.58**	13.73**	2.86	-1.59	6.04*	23.81**	34.07**	-27.00*
$L_6 \times T_4$	4.45	-34.40**	-70.34**	-81.62**	64.29**	-43.01**	-38.21**	-4.76	-68.15**	**00.67
$L_6x T_5$	-5.63*	-34.79**	-13.04**	2.96	-14.29	29.72**	-12.87**	10.00	-35.66**	102.00**
$L_6x T_6$	-0.76	-14.63	-42.32**	-22.09**	-25.71**	-16.49**	-44.83**	14.29**	-48.97**	63.00**
$L_7 \times T_1$	-0.36	-32.19**	22.44**	-43.45**	120.00**	-18.36**	24.17**	0.27	-9.03**	-10.00
$L_7 \times T_2$	-0.90	-36.26**	-5.19	0.41	-4.29	*9′.9	-19.69**	0.35	-45.80**	**00.86
$L_7 \times T_3$	-12.74**	-37.46**	30.89**	-47.74**	152.86**	-35.62**	-9.94**	24.76**	**80.8-	-30.00**
$L_7 \times T_4$	-3.30	-35.01**	-54.99**	-52.28**	-4.29	-32.67**	-26.32**	-8.57	-61.97**	**00.77
$L_7 \times T_5$	-8.42**	-29.48*	-6.84	-1.06	-5.71	-30.45**	-26.32**	23.81**	-33.76**	5.00
$L_7 \times T_6$	14.81**	-4.66	-24.05**	-51.31**	57.14**	-8.47**	-2.14	-19.05**	-19.65**	14.00
$L_8 \times T_1$	14.73**	-29.49*	-59.03**	-14.70**	-50.00**	-36.66**	-30.21**	-57.14**	-74.01**	143.00**
$L_8 \times T_2$	-7.30**	-3.01	-46.10**	-44.67**	-2.86	-5.58	3.31	29.52**	-2.38	-3.00
$L_8 \times T_3$	-10.76**	-17.47	-5.94	-41.40**	61.43**	-17.95**	-45.81**	-13.33*	-62.12**	116.00**
$L_8 \times T_4$	1.71	-48.36**	-5.95	9.65**	-14.29	-8.34**	4.87	-29.52**	-29.79**	31.00**
$L_8 \times T_5$	0.83	-43.62**	77.95**	36.31**	31.43**	1.71	-8.77**	0.18	-29.64**	44.00**
$L_8 \times T_6$	-0.20	-34.75**	60.39**	-27.17**	122.86**	-0.41	-12.09**	-11.90**	-36.61**	57.00**
$L_9x T_1$	3.98	-31.40**	-77.97**	-21.68**	-71.43**	-6.53*	-57.50**	19.05**	-79.40**	355.00**
$L_9 \times T_2$	-0.20	-13.12	-49.10**	-29.95**	-25.71**	19.25**	-29.63**	5.71	5.86*	13.00
$L_9 \times T_3$	2.79	-30.36*	-28.24**	-31.52**	5.71	26.74**	-17.35**	14.29**	-32.65**	**00.88
$L_9 \times T_4$	**66.9	-17.68	-17.98**	-21.07**	4.29	19.89**	-37.82**	19.52**	-56.74**	179.00**
$L_9 \times T_5$	1.67	-10.31	102.27**	64.09**	24.29**	43.04**	81.09**	4.76	103.49**	-30.00**
$L_9 \times T_6$	-14.12**	-12.09	-72.97**	-73.33**	2.86	8.98**	-49.12**	7.62**	-19.81**	36.00**
$L_{10} \times T_1$	6.11*	-38.98**	-66.65**	-66.04**	0.12	-19.03**	-42.88**	-25.71**	-69.10**	162.00**
$L_{10}x$ $T_2$	-4.29	-30.20*	18.40**	-36.67**	88.57**	*90.9	4.09	-48.57**	-52.14**	121.00**
$L_{10}x T_3$	8.81**	24.15*	-60.12**	-59.30**	-1.43	-15.83**	-60.82**	5.71	-64.82**	139.00**
$L_{10}x$ T <sub>4</sub>	5.44*	-29.52*	-58.70**	-43.66**	-25.71**	-31.72**	-13.45**	-59.05**	-58.16**	63.00**
$L_{10}x T_5$	-7.31**	-34.83**	19.86**	28.61**	-5.71	-16.37**	-20.47**	18.57**	-31.54**	23.00*
$L_{10X}$ T <sub>6</sub>	8.26**	-59.68*	-67.35**	-42.84**	-41.43**	-21.76**	-40.55**	0.43	-59.27**	92.00**

root/shoot ratio (21.43), root length (20.53), number of roots per plant (67.84), root thickness (53.33) and root volume (83.04). The cross, ADT 43 x PMK (R) 3 ( $L_3$  x T<sub>5</sub>) had significant standard heterosis for the traits *viz.*, spad chlorophyll meter reading (5.48), dry root weight (24.70) Root/Shoot ratio (112.86), number of roots per plant (24.56) and root thickness (20. 59) over check variety. CO (R) 50 x PMK (R) 3 (L9 x T5) had observed positively significant heterosis for dry root weight (102.27) dry shoot weight (64.09), Root/Shoot ratio (24.29), root length (43.04), number of roots per plant (81.09) and root volume (103.49). Presence of non-additive gene action for drought tolerant traits in the hybrids resulted in high amount of vigour in F<sub>1</sub> indicating the possibility of augmenting yield and drought tolerance by exploiting heterosis (Manonmani and Fazullah Khan, 2003). These results are in agreement with those reported by Abd Allah (2004) and El-Mouhamady et al. (2013) in rice. These hybrids can be used as a potential genetic source for better root system with higher efficiency to absorb moisture effectively for tolerating drought condition and increasing yield.

#### Conclusion

In this present study, it was inferred that all the traits were governed by non additive gene action. The most appropriate breeding technique to exploit non additive gene action will be through heterosis breeding. The hybrids showing additive action can be improved by pedigree breeding and selection can be postponed to later generations. The hydrids which shows significant heterosis are highly suitable for commercial exploitation of heterosis under moisture stress conditions.

## REFERENCES

- Abd Allah, A.A. (2004). A breeding study on drought tolerance in rice (*Oryza sativa* L.). Egyptian Journal of Agricultural Research, 82 (1): 149-165.
- Allard, R.W. (1960). *Principles of Plant Breeding* John Wiley and Sons, Inc. New York.
- Bhattacharya, S. and Ghosh, S K. (2004). Association among yield related traits of 24 diverse land races of rice. *Crop Research* 27(1): 90-93.
- Das, K., Pradhan, T. Ghosh, S. and Mishra, B.K. (2005). Evaluation of drought resistance characters of upland rice cultivars. *Oryza*, 42(2): 138-144.
- Dhillon, B.S. (1975). The application of partial diallel crosses in Plant breeding A review. *Crop Improvment* 2: 1-7.
- Dixit, A.S. and Kumar, A. (2014). Rice Breeding for high grain yield under drought: Strategic solution to a complex problem pp 1-15.
- El-Mouhamady, A.A., Abdel-Sattar, A.A. and El-Seidy, E.H. (2013). Assessment of the degree of drought tolerance

- in rice through the environmental tests and molecular markers technique. *Research Journal of Agriculture and Biological Sciences*, 9(1): 40-57.
- Ganesh, S.K., Vivekanandan, P. Nadarajan, N. Chandra Babu, R. Shanmugasundaram, P. Priya, P.A. and Manickavelu, A. (2004). Proceedings of Workshop on resilient crops for water limited environments. Cuernavaca, Mexico, pp.140.
- Gnanasekaran, M., Vivekanandan, P. and Muthuramu, S. (2006). Combining ability and heterosis for yield and grain quality in two line rice (*Oryza sativa* L.) hybrids. *Indian J.ournal of Genetics* 66(1): 6-9.
- Hanamaratti, N.G., Salimath, P.M. Mohankumar, H.D. and Shailaja, H. (2004). Genetic and physiological basis of breeding productive and drought tolerant genotypes in upland rice. Proceedings of workshop on resilient crops for water limited environments. Cuernavaca, Mexico, pp 100-101.
- Hosseini, M., Nejad, R.H. and Tarang, A.R. (2005). Gene effects, combining ability of quantitative characteristics and grain quality in rice. *Iranian Journal of Agricultural Sciences 36* (1): 21-32.
- Kempthorne, O. 1957. *An Introduction to Genetic Statistics*. John Wiley and Sons, Inc., New York, pp. 399-472.
- Luu, T.N., Abdelbagi, L.M. Ismail, M. and Le, H. (2012). Introgression of salinity tolerance QTLs Saltol into AS996, the elite rice variety of Vietnam. American Journal of Plant Sciences 3: 981-987.
- Malarvizhi, D., Thiyagarajan, K. Vijayalakshmi, C. and Manonmani, S. (2010). Genetic analysis to assess the physiological efficiency of parental lines in rice. *Electronic Journal of Plant Breeding* 1(2): 100-113.
- Manonmani, S. and Fazlullah Khan, A.K. (2003). Studies on combining ability and heterosis in rice. *Madras Agricultural Journal*, 90 (6): 228-231.
- Manonmani, S. and Fazlullah Khan, A.K. (2005). Multiseason evaluation of rice hybrids. *Plant Archives* 5(2): 435-440.
- Muthuramu, S., Jebaraj, S. and Gnanasekaran, M. (2010). Combining ability and heterosis for drought tolerance in different locations in rice (*Oryza sativa L.*). Research Journal of Agricultural Sciences 1(3): 266-270.
- Panse, V.G. and P.V. Sukhatme. (1964). Statistical Methods for Agricultural Research Workers, 2<sup>nd</sup> Ed. ICAR, New Delhi.
- Sathya, R. and Jebaraj, S. (2013). Studies on choice of parents and gene action in rice hybrids involving yield and physiological traits under aerobic condition. *Plant Gene Trait*, 4 (19): 104-108.
- Snedecor, G.W. and W.G. Cochran. (1967). Statistical Methods. VI edition, Iowa state University press, Iowa, USA.
- Sprague, G.F. and Tatum, L. A. (1942). General versus specific combining ability in single cross of corn. *Journal of American Society of Agronomy 34*: 983-992.
- Utharasu, S. and C. R. Anandakumar. (2013). Heterosis and combining ability analysis for grain yield and its component traits in aerobic rice (*Oryza sativa* L.) cultivars. *Electronic Journal of Plant Breeding* 4(4): 1271-1279.