


*Research Article***Isolation of cis-2-methylpalmitolein-9 acid from the fungus *Trichoderma asperellum* Uz-A4 and its Spectroscopic analysis****Mohigul Nomozova\*** 

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Today, the use of natural and non-harmful substances is one of the main and a pressing issue in all industries. In this regard, most researchers are focused on extracting the necessary components from natural sources and raw materials. At the same time, the purpose of the present practical work is based on the extraction of biologically active substances from microbodies (fungi). In this exploratory study, the extraction of secondary metabolites from the strain *Trichoderma asperellum* Uz-A4 was performed. Fungal biomass was prepared and extracted using alcoholic extraction, and the resulting extract was fractionated by column chromatography using various solvents. In the chloroform: methanol 9:1 system step of the fractionation process, a high content of cis-2-methylpalmitolein-9 acid was formed, along with a number of secondary metabolites. For the first time, this unsaturated fatty acid was isolated from the strain of *Trichoderma asperellum* Uz-A4. When the structure of this substance was isolated as a pure substance and its structure was studied by other spectroscopic assays, it turned out to be cis-2-methylpalmitoleic acid.

**Keywords:** Antifungal, *cis-2-methylpalmitolein-9 acid*, Extract, Fungus, Metabolism, Spectroscopic, *Trichoderma asperellum***INTRODUCTION**

Palmitoleic acid is a fatty acid with good bioactivity, possessing several important properties that can be utilised in both the nutritional and pharmaceutical fields. The production of volatile organic metabolites (VOCs) is a manifestation of the action properties of *Trichoderma* spp. One of them is the resistance to the growth and development of pathogenic fungi or bacteria. In addition to showing an average inhibition value of 64.49% in *T. asperellum* (T4) and 61.59% in *T. harzianum* (T12), 98 volatile metabolites were identified by GC-MS analysis. For example, phenylethyl alcohol, several phenolic isomers, benzene derivatives, D-limonene, octadecanoic acid methyl ester, hexadecanoic acid, toluene, eicosamine and eicosamine-like vola-

tiles with antifungal activity were identified (Reghmit *et al.*, 2024). Additionally, in an experiment conducted on another *Trichoderma* strain, two new antifungal metabolites, designated as cytosporones Y (1) and Z (2), were identified in the broth prepared from *Trichoderma* sp. FKI-6626. MS and NMR studies of the identified substances were carried out and both metabolites were identified. Both compounds were found to have antifungal activity against 4 species of *Aspergillus*, which are causative agents of aspergillosis (Azami *et al.*, 2024). Fatty acids in general are important molecules that perform the main function in most organisms and play a role as signaling agents in plant-plant, plant-microbial and plant-environment interactions (Raffaele *et al.*, 2009). Several studies have shown that fatty acids and

their derivatives directly inhibit the growth of plant pathogens in the rhizosphere and improve the environment of the plant rhizosphere to reduce the occurrence of diseases found in crops and improve crop growth (Davis *et al.*, 1997; Liu *et al.*, 2012 and Liu *et al.*, 2008). Palmitic acid (PK) is a saturated long-chain fatty acid compound that is common in beans, sunflowers, and cotton seeds (Lukonge *et al.*, 2007; Perez-Vich *et al.*, 2016; Zhao *et al.*, 2019). In addition, it is found in high levels in exudates and residues of many plant roots (Guo *et al.*, 2010; Pan *et al.*, 2013). Previous studies have demonstrated that PK can inhibit the growth of plant-based pathogens and promote the growth of seedlings (Abdel-Naime *et al.*, 2019; Ding *et al.*, 2019). This fatty acid (9-hexadecenoic acid or palmitic acid) has also been detected in *T. hamatum* filtrate (peak area-3.96%), initially identified as an indicator of antimicrobial activity (Khairillah *et al.*, 2021 and Ghavam *et al.*, 2021). Additionally, palmitic acid has been detected in the filtrate of *T. asperellum* and is utilised as a herbal feed supplement as well as in the treatment of certain diseases (Singh *et al.*, 2021). *T. asperellum* culture fluids have significant cytotoxic effects against plant pathogens (Stracquandano *et al.*, 2020) and palmitic acid has also been identified as a biomarker of *T. virens* against cotton fungus disease caused by *Rhizoctonia solani* (Gajera *et al.*, 2020), both of which are *T. sp.* detected in the extract of. To date, 75 species of the genus *Trichoderma* have been studied genetically, and work in this regard continues with consistency (Druzhinina *et al.*, 2011). Having familiarised with the above sources, the present study aimed to determine that cis-2-methylpalmitolein-9 acid, one of the secondary volatile metabolites produced by the fungus of the *Trichoderma* family, has an important antifungal function.

## MATERIALS AND METHODS

### Technology for preparing fungal extract

A strain of *Trichoderma asperellum* Uz-A4, isolated from the rhizosphere of corn roots growing in the Tashkent region of the Republic of Uzbekistan, was grown in Mandeléa agar medium (in a test tube) for 6 days and used as an ectopic material from a suspension at a concentration of  $10^6$ - $10^7$  spores/mL.

The fungus of *T. asperellum* Uz-A4 was grown in 250 ml nutrient medium in 500 ml volume Erlenmeyer tubes under feed conditions, in a shaker at a speed of 180 cycles/min (IKA® KS 130 shakers), for 14 days at a temperature of 24-26°C, and separated from its cultural fluid biomass by filtering. The separated biomass and cultured liquid were extracted. The total weight of the extract sum was 9.25g. The resulting extraction sum was adsorbed to 400 g Silpearl silicagel. The adsorbent was placed in a Columnar chromatographic column. When columnar chromatography using a chloroform:methanol system (9:1) was performed, 7.3 g of cis-2-methylpalmitolein-9 acid (I) was isolated, and its structure was studied (Jurakulova *et al.*, 2021; Kamolov *et al.*, 2021). Chloroform:methanol 9:1 fraction with thin layer and gaseous liquid chromatography (Fig. 1)

### Metabolite analysis methods

Volatile metabolites were analyzed by gas chromatography and mass spectrometry using a YL 6900 GX/MS (Young In Chromass, Korea) equipped with a DB-5MS column (30 m × 0.25 mm inner width and 0.25 µm film thickness). The initial temperature of the apparatus was 80 °C, and the heating rate was 15 °C/min to 250 °C. The holding time was 3.0 min, and helium was used as the carrier gas at a flow rate of 1.0 mL/min. The volatilization temperature was 280°C, the flow rate was 1/20,

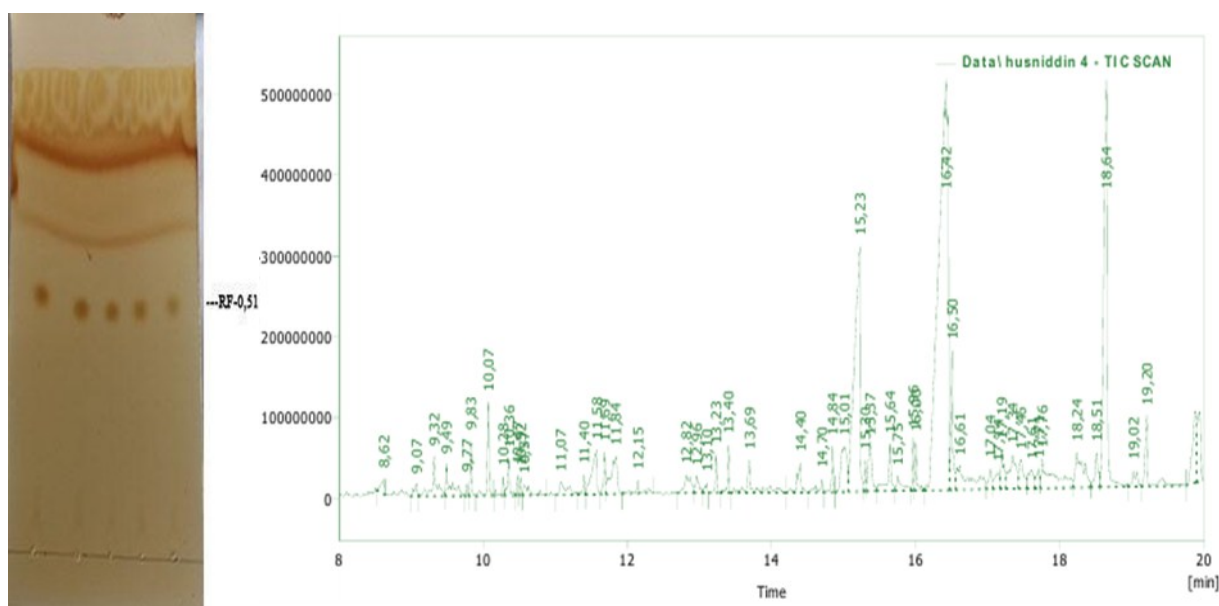


Fig. 1. Thin layer and gaseous liquid chromatography

and the analysis time was 17 min. Liquid samples were injected into the syringe using a 1  $\mu$ L microsyringe. The transfer line temperature was 300°C, the ionization voltage was -70 eV, and the temperature of the ion generating section was 230°C. The scanning range was 30-350 a.m.u. The substances were identified by comparing their spectra with those of the available spectral data in the NIST 2017 MS database. YaMP spectra were obtained in deuteropyridine and deuteriochloroform ( $\delta$ , m.d., 0-TMS) in the UNITY plus 400 instrument (Kamolov *et al.*, 2021).

## RESULTS AND DISCUSSION

### IQ spectrum of cis-2-methylpalmitolein-9 acid

The extract from biomass was obtained by fractionating a chloroform:methanol system in a 9:1 ratio, using 500  $\mu$ g of the extracted metabolite I with a concentration of 500  $\mu$ g/mL. An IQ spectrum in tablet form was obtained. The present study investigated metabolite I using IQ spectra (Fig. 2). It is well established that physical-chemical research methods provide valuable data in the study of organic substance structure and the composition of isolated metabolites. The absorption lines of the C-H bond bonded to the  $sp^2$  hybrid C atom in the 3008  $cm^{-1}$  aliphatic chain were observed in the IQ spectrum of palmitoleic acid. In the range 2919-2853  $cm^{-1}$  absorption lines of the C-H bond bound to the  $sp^3$  hybrid C atom were observed. In the field range, we can see absorption lines formed by the deformation oscillations of the O-H bond in the carboxyl group.

### PMR spectrum of cis-2-methylpalmitolein-9 acid

The PMR spectrum of the substance is given (Fig. 3). In the PMR analysis of the substance, R-CH<sub>3</sub> in the  $sp^3$  hybrid methyl group C-16 knocks out 3H triplet signals

at 0.880 m.h. of carbon-bound hydrogen. Multiple signals of 10 methylene-CH<sub>2</sub>-group hydrogens can be observed at 1,255 mHz and 1,309 mHz. C-11 and C-8 emit quartet signals at 1,629 m.h. and 2,018 m.h. because they bind to the carbon-containing methylene-CH<sub>2</sub>- group. Jump to search Hydrogen signals located in the -OH group in the carboxyl group produce a singlet signal at 5,344 m.h.

500  $\mu$ g of cis-2-methylpalmitolein-9 acid was dissolved in methanol and placed in a nuclear magnetic tube. <sup>1</sup>H-NMR and <sup>13</sup>C-NMR analysis of the sample Bruker spectrometer (400 MHz) (Bruker, Germany) (Van Leeuwen *et al.*, 2008) was performed for analysis.

### GDP C<sup>13</sup> spectrum of cis-2-methylpalmitolein-9 acid

YaMR C<sup>13</sup> spectroscopic (nuclear magnetic resonance) methods were used to determine the cis-2-methylpalmitolein-9 acid obtained from this fraction (Fig. 4).

In the analysis of the substance on GDP C<sup>13</sup>, the  $sp^3$  hybridized R-CH<sub>3</sub> carbon signal was observed at 14,256 m.h. (million contributions), and the R-CH<sub>2</sub>-R carbon signal in the methylene group was observed at 27.279 mHz to 29.896 mHz. (14 carbon signals for the presence of 14 methylene). Specific signals were observed at 129 m.h. and 130 m.h. of the two carbons C=C. The signal was observed at 180 m.h. of R-COOH carbon, which belongs to the carboxyl group.

### HMBC spectrum of cis-2-methylpalmitolein-9 acid

Our next result is the HMBC spectrum of 2-methylpalmitolein-9-acid, which shows the correlation effects of geminal and vicinal carbons and hydrogens (Fig. 5).

The spin-spin interaction of the proton of the -CH<sub>3</sub> radi-

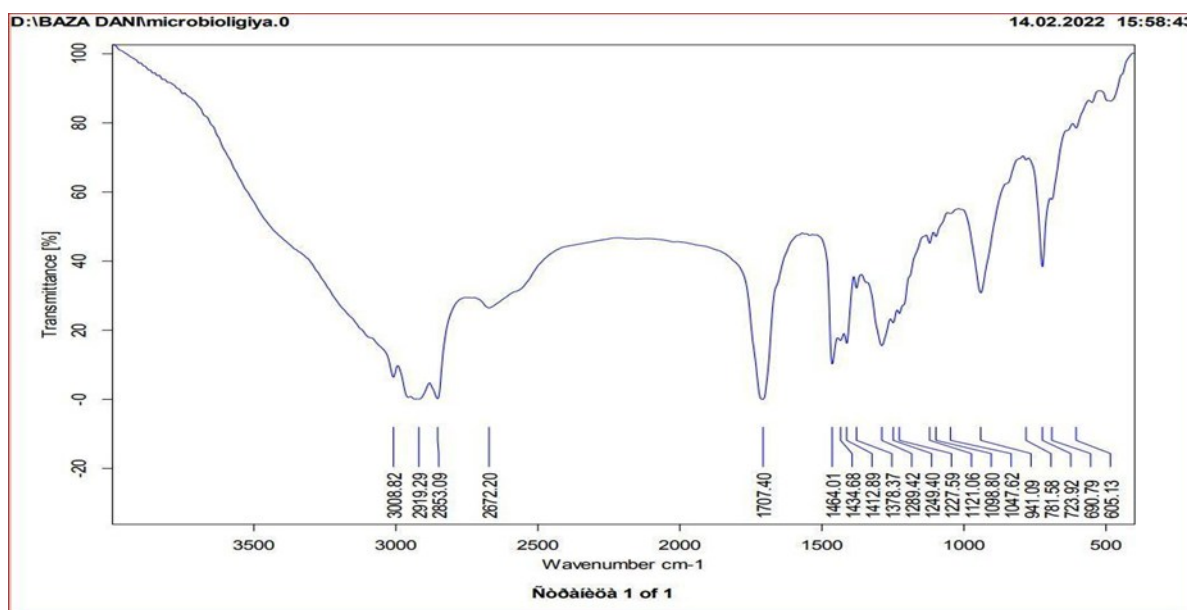


Fig. 2. IQ spectrum of cis-2-methylpalmitolein-9 acid

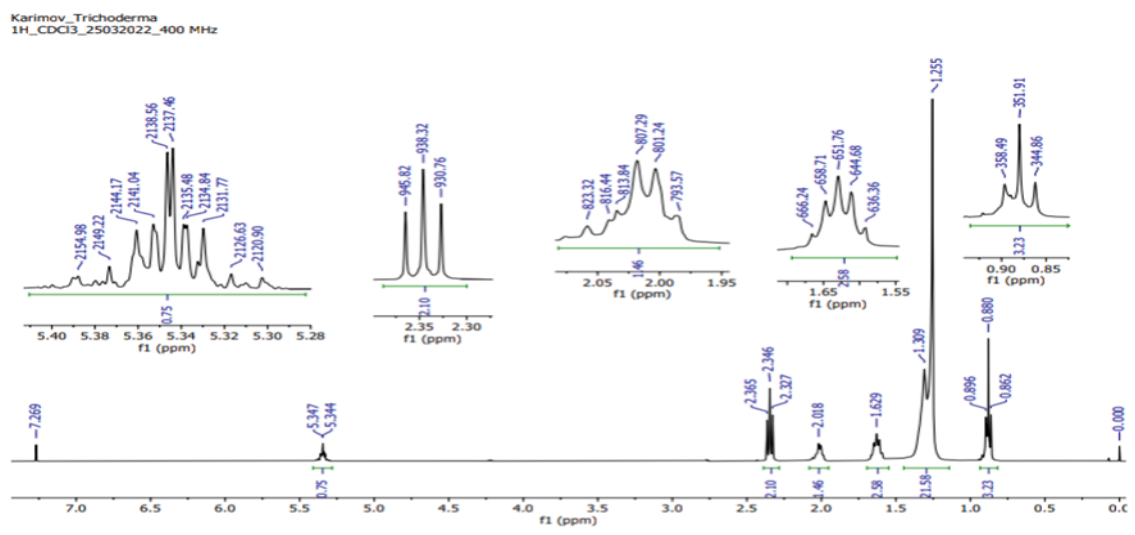


Fig. 3. PMR spectrum of *cis*-2-methylpalmitolein-9 acid

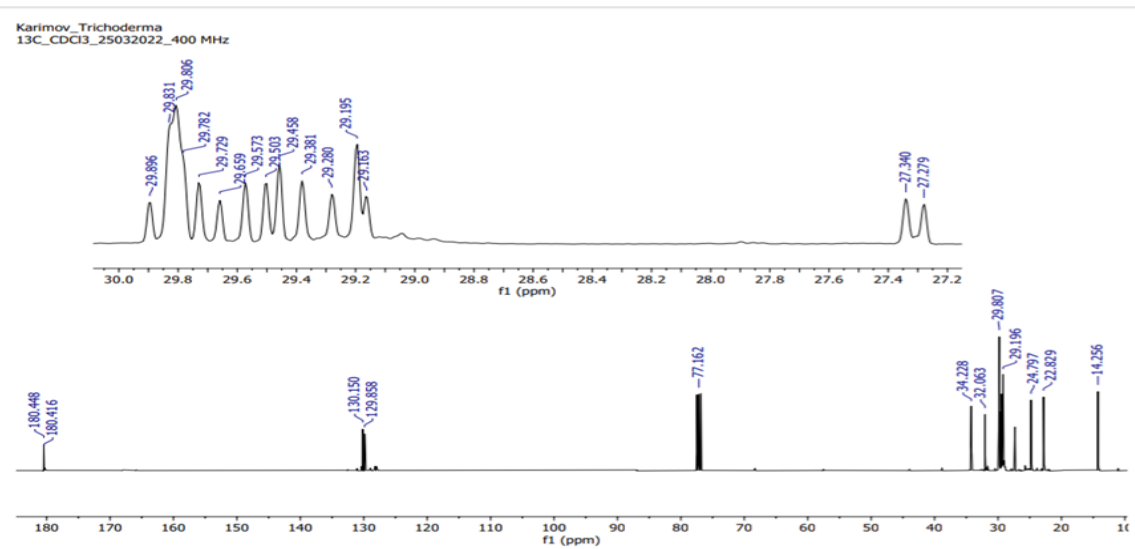


Fig.4. GDP  $^{13}\text{C}$  spectrum of *cis*-2-methylpalmitolein-9 acid

cal at 1.629 m.p. in the HMBC spectrum of 2-methylpalmitolein-9 acid with the carbonyl group at carbon C-1 indicates that it is vicinal to the carbonyl group. The interaction of the proton at carbon C-2 at 2.346 m.p. in the HMBC spectrum with the C=C carbon at 130.162 m.p. indicates that the protons are located in the double bond. The interaction of the signal at 5.347 m.p. specific to the proton in the -OH group of 2-methylpalmitolein-9 acid with the signal at 27.216 m.p. specific to the C-2 carbon indicates that the -OH group is vicinal.

The HMBC spectrum of *cis*-2-methylpalmitolein-9-acid is presented, which describes the correlation effects between the carbon and hydrogen atoms of the CH<sub>2</sub> Groups in the geminal and vicinal states (Fig. 6).

#### HSQC spectrum of *cis*-2-methylpalmitolein-9 acid

Below is the HSQC spectrum of *cis*-2-methylpalmitolein-9-acid, which shows the correlation effects of geminal

carbon and hydrogen (Fig. 7).

The spin-spin interaction of the -CH<sub>3</sub> radical proton, which showed a signal at 0.878 m.p. in the HSQC spectrum of 2-methylpalmitolein-9 acid, with the signal at 14.104 m.p. of the C-16 carbon indicates that it is located at the C-16 carbon. The spin-spin interaction of the -CH<sub>3</sub> radical proton, which showed a signal at 1.626 m.p. in the HSQC spectrum of 2-methylpalmitolein-9 acid, with the signal at 24.731 m.p. of the C-17 carbon indicates that it is located at the C-17 carbon. The interaction of the signal at 2.010 m.p. of the proton located at the C-2 carbon with the signal at 27.222 m.p. indicates that the protons are located at the C-2 carbon. The signal at 2.342 m.p., which is characteristic of the proton located in the C=C group of 2-methylpalmitolein-9 acid, with the signal at 34.196 m.p., which is characteristic of the C-8 carbon. The interaction with the signal indicates that it is located in the geminal state at the C-8 carbon.

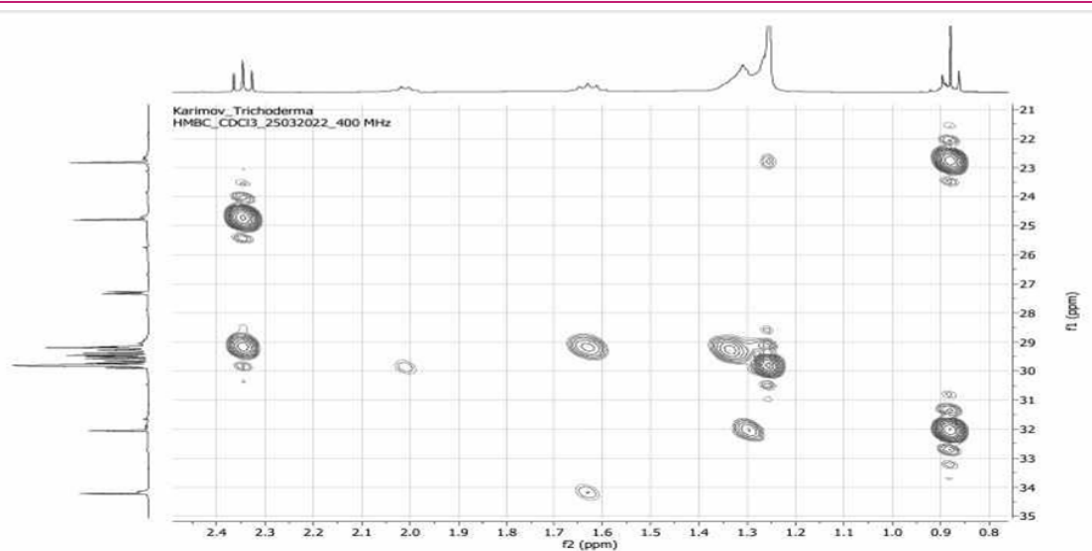


Fig. 5. HMBC spectrum of *cis*-2-methylpalmitolein-9 acid

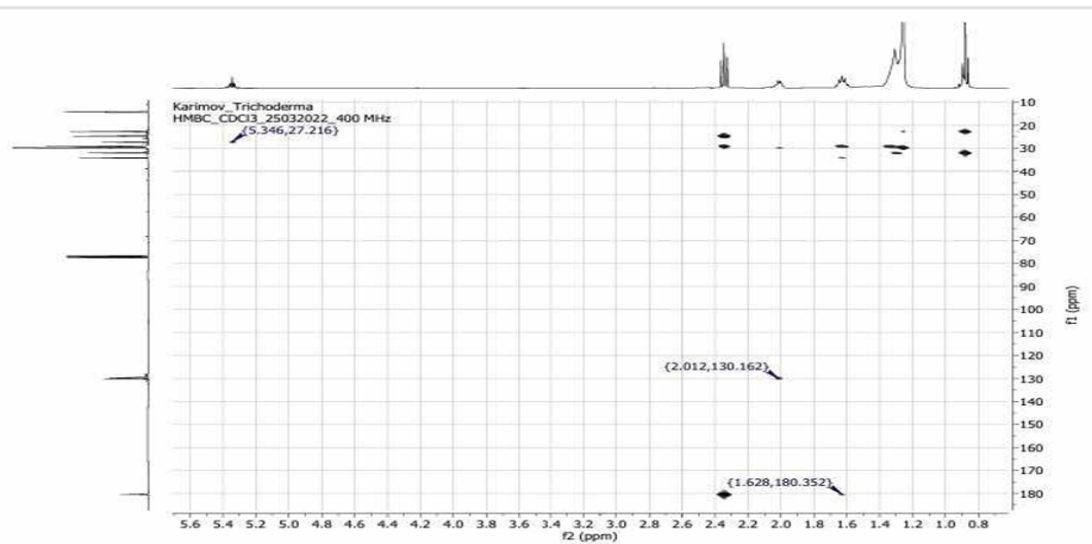


Fig. 6. HMBC spectrum of *cis*-2-methylpalmitolein-9 acid

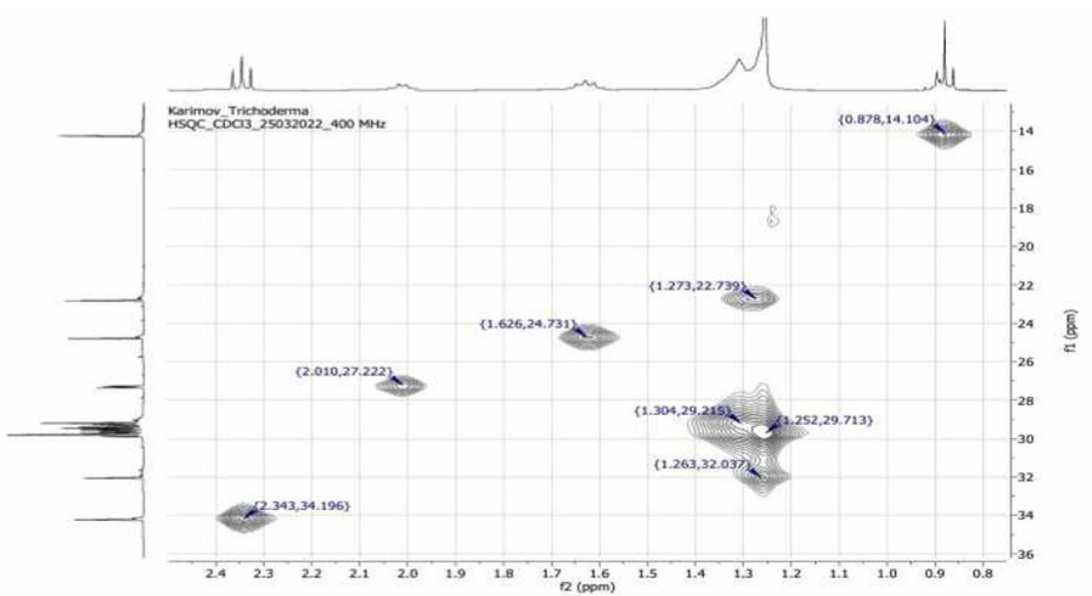
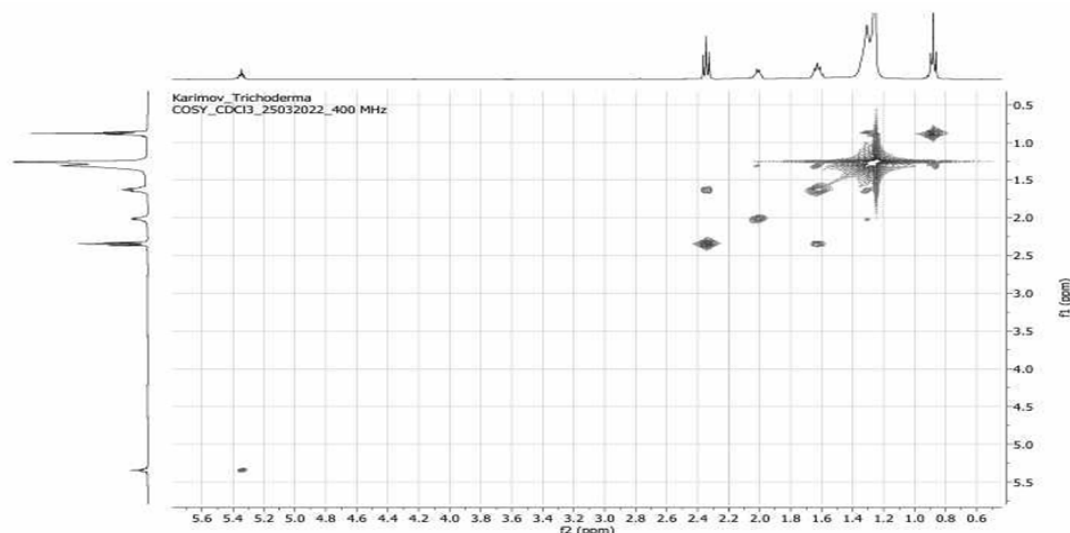
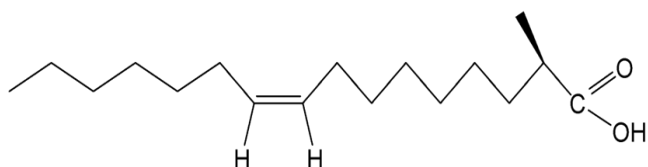


Fig. 7. HSQC spectrum of *cis*-2-methylpalmitolein-9 acid



**Fig. 8.** COSY spectrum of *cis*-2-methylpalmitolein-9 acid



**Fig. 9.** Structure of *Cis*-2-methylpalmitolein-9 acid

#### COSY spectrum of *cis*-2-methylpalmitolein-9 acid

The COSY spectrum of *cis*-2-methylpalmitolein-9-acid is given, which describes the correlation effects of protons in the geminal and vicinal states (Fig. 8).

The interaction of the signal at 1.629 m.p. of the proton located at carbon C-17 with the signal at 2.010 m.p. of the proton located at carbon C-2 indicates that they are geminal to each other.

According to the obtained spectroscopic studies, the structural formula of *cis*-2-methylpalmitolein-9 acid was determined (Fig. 9).

Among the secondary metabolites isolated from the strain of *Trichoderma asperellum* Uz-A4, the unsaturated fatty acid *cis*-2-methylpalmitolein-9 was considered to be isolated for the first time. Scientific research in these areas has shown that various volatile compounds, including fatty acids (Zeiad *et al.*, 2023; Ivan *et al.*, 2023), as well as 43 other volatile compounds (Nitish *et al.*, 2017). Although representatives of different classes of organic compounds have also been identified in research studies, the fact that the same unsaturated acid has not been identified indicates that this case differs from similar studies.

#### Conclusion

In the present study, the compound *cis*-2-methylpalmitolein-9 acid was isolated from the biomass of the fungal strain *T. asperellum* Uz-A4 and separated in the chloroform: methanol (9:1) fraction. The structure

of this substance was determined based on GLX, IR, NMR <sup>13</sup>C and PMR, HMBC, HSQC, COSY spectroscopic analyses. The substance was the most abundant (7.3 g) in terms of quantity.

#### Conflict of interest

The authors declare that they have no conflict of interest.

#### REFERENCES

1. Abdel-Naime, W.A., J.R. Fahim, M.A. Fouad & M.S. Kamel. (2019). Antibacterial, antifungal, and GC-MS studies of *Melissa officinalis*. *S. Afr. J. Bot.*, 124, pp. 228-234. <https://doi.org/10.1016/j.sajb.2019.05.011>
2. Azami, H., Watanabe, Y., Kojima, H. *et al.* (2024). Cyto-sporones Y and Z, new antifungal polyketides produced by the fungal strain *Trichoderma* sp. FKI-6626. *J. Antibiot.* 77, 721–726. <https://doi.org/10.1038/s41429-024-00765-9>.
3. Davis, E.L., D.M. Meyers, C.J. Dullum & J.S. Feitelson. (1997). Nematicidal activity of fatty acid esters on soybean cyst and root-knot nematodes. *J. Nematol.*, 29 (4), pp. 677 -684. PMID: 19274268; PMCID: PMC2619835.
4. Ding, L.S., W.B. Guo & X.H. Chen (2019). Exogenous addition of alkanolic acids enhanced production of antifungal lipopeptides in *Bacillus amyloliquefaciens* Pc3Appl. *Microbiol. Biotechnol.*, 103 (13), 5367-5377.
5. Druzhinina IS, Seidl-Seiboth V, Herrera-Estrella A *et al.* (2011). *Trichoderma*: the genomics of opportunistic success. *Nat Rev Microbiol.*, 9 (10):749–759. doi: 10.1038/nrmicro2637
6. Gajera H.P., Hirpara D.G., Savaliya D.D. & Golakiya B.A. (2020). Extracellular metabolomics of *Trichoderma* bio-controller for antifungal action to restrain *Rhizoctonia solani* Kuhn in cotton. *Physiol. Mol. Plant Pathol.*, 112:101547. doi: 10.1016/j.pmpp.2020.101547.
7. Ghavam, M.; Afzali, A. & Manca, M.L. (2021). Chemotype of Damask rose with oleic acid (9 octadecenoic acid) and its antimicrobial effectiveness. *Sci. Rep.*, 11, 8027. <https://doi.org/10.1038/s41598-021-87604-1>

8. Guo, X.W., Kun, L.I., Sun, Y.N., Zhang, L.H., Xi-Xi, H.U., Hong-Gang, *et al.*, 2010. Allelopathic effects and identification of allelochemicals in grape root exudates. *Acta Horticulturae Sinica*, 37(6), 861-868. <https://www.cabdirect.org/cabdirect/abstract/20103220851>
9. Ivan Chóez-Guaranda, Fernando Espinoza-Lozano, Denys Reyes-Araujo, Christian Romero, Patricia Manzano, Luis Galarza & Daynet Sosa (2023). Chemical Characterization of *Trichoderma* spp. extracts with antifungal activity against Cocoa pathogens. *Molecules*: 4;28(7):3208. doi: 10.3390/molecules28073208
10. Jurakulova N.X., & Kamolov L.S. (2021). *Stachybotrys chartarum* secondary metabolites of poisonous Microzoquary. *International Journal of Pharmaceutical and Bio Medical Science*, 1(8), 151–158. <https://doi.org/10.47191/ijpbms/v1-i8-05>
11. Kamolov Luqmon, Sevara Tojiyeva, SHuxrat Xasanov, Elyor Berdimurodov & Orif Axmedov. 2021. Stachybotrus, a toxic microscopic fungus, produces low-molecular-weight metabolites. *Plant cell Biotechnology and Molecular Biology*, 22 (35-36):50-61. <https://ikpress.org/index.php/PCBMB/article/view/6301>
12. Khairillah, Y.N.; Sukarno, N. & Batubara, I. (2021). *Trichoderma hamatum* derived from coffee plant (*Coffea canephora*) rhizosphere inhibits *Candida albicans* growth. *Biosaintifika J. Biol. Biol. Educ.* 13, 369–378. <http://dx.doi.org/10.15294/biosaintifika.v13i3.31132>
13. Liu, S.Y., W.B. Ruan, J. Li, H. Xu, J.G. Wang, Y.B. Gao & J. Wang. (2008). Biological control of phytopathogenic fungi by fatty acids. *Mycopathologia*, 166 (2), 93-102. <https://doi.org/10.1007/s11046-008-9124-1>
14. Liu, P., Z.H. Liu, C.B. Wang, F. Guo, M. Wang, Y.F. Zhang, *et al.* (2012). Effects of three long-chain fatty acids present in peanut (*Arachis hypogaea* L.) root exudates on its own growth and the soil enzymes activities. *Allelopathy J.*, 29 (1) Lukonge, E., M.T. Labuschagne, A. Hugo. (2007). The evaluation of oil and fatty acid composition in seed of cotton accessions from various countries, *J. Sci. Food Agric.*, 87 (2), 340-347. <https://doi.org/10.1002/jsfa.2731>
15. Nitish Rattan Bhardwaj & J. Kumar. (2017). Characterization of volatile secondary metabolites from *Trichoderma asperellum*. *Journal of Applied and Natural Science*, 9 (2):954-959. DOI:10.31018/jans.v9i2.1303
16. Pan, K., L.H. Xu, F.Z. Wu, Z. Han, L.R. Chu & C.F. Qiu. (2013). Fungicidal effects of wheat root exudates on *Fusarium oxysporum* f. sp. *niveum*. *Allelopathy Journal*, 32 (2), 257-265.
17. Perez-Vich, B., L. del Moral, L. Velasco, B.S. Bushman, S.J. Knapp, A. Leon, *et al.* (2016). Molecular basis of the high-palmitic acid trait in sunflower seed oil. *Mol. Breed.* 36 (4), p. 12. <https://doi.org/10.1007/s11032-016-0462-2>
18. Raffaele, S., A. Leger & D. Roby. (2009). Very long chain fatty acid and lipid signaling in the response of plants to pathogens. *Plant Signaling Behav.*, 4 (2), 94-99. <https://doi.org/10.4161/psb.4.2.7580>
19. Reghmit Abdenaceur., Benzina-tihar Farida. & Sahir-Halouane Fatma. (2024). Volatile organic compounds activities of *Trichoderma* species isolated from olive grove soil against the wilt pathogen, *Verticillium dahlia*. *European Journal of Plant Pathology*, 01 March. Volume 170, pages 789–803. <https://doi.org/10.1007/s10658-024-02839-8>
20. Singh, G.; Tiwari, A.; Gupta, A.; Kumar, A.; Hariprasad, P. & Sharma, S. (2021). Bioformulation Development via Valorizing Silica-Rich Spent Mushroom Substrate with *Trichoderma Asperellum* for Plant Nutrient and Disease Management. *J. Environ. Manag.* 297, 113278. <https://doi.org/10.1016/j.jenvman.2021.113278>
21. Stracquadanio C., Quiles J.M., Meca G. & Cacciola S.O. (2020). antifungal activity of bioactive metabolites produced by *Trichoderma* and *Trichoderma* in liquid medium. *J. Fungi.*; 6:263. doi: 10.3390/jof6040263.
22. Van Leeuwen, S.-S., Leeflang, B. R., Gerwig, G.-J., & Kamerling, J.-P. (2008). Development of a <sup>1</sup>H-NMR structural-reporter-group-concept for the primary structural characterization of α-d-glucans. *Carbohydrate Research*, 343(6), 1114-1119. doi: 10.1016/j.carres.2008.01.043.
23. Zhao, X, H. Chang, L. Feng, Y. Jing, W.L. Teng, L.J. Qiu, *et al.* (2019). Genome-wide association mapping and candidate gene analysis for saturated fatty acid content in soybean seed. *Plant Breeding*, 138 (5), pp. 588-598.
24. Zeiad Moussa, Yasmene F Alanazi, Aiah Mustafa Khateb, Noha M Eldadamony, Marwa M Ismail, WesamEldin I A Saber & Doaa Bahaa Eldin Darwish. 2023. Domiciliation of *Trichoderma asperellum* suppresses *Globiosporangium ultimum* and promotes pea growth, ultrastructure, and metabolic features. *Microorganisms*, 12;11(1),198. doi: 10.3390/microorganisms11010198