

Research Article

Soil nematode communities as indicators of forest ecosystem health: A study in Patnitop region of District Udhampur, UT, J&K, India

Anil Bhardwaj

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

Fozia Choudhary

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

Iqra Sayeed

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

Yasmeen Kouser

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

Nazia Kouser

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

Ali Asghar Shah

Nematode Biodiversity and Genomics Research lab, BGSBU, Rajouri, UT, J&K, India

*Corresponding author. E-mail: headzoology@bgsbu.ac.in

Article Info<https://doi.org/10.31018/jans.v17i3.6732>

Received: April 23, 2025

Revised: August 21, 2025

Accepted: September 01, 2025

How to Cite

Bhardwaj, A. *et al.* (2025). Soil nematode communities as indicators of forest ecosystem health: A study in Patnitop region of District Udhampur, UT, J&K, India. *Journal of Applied and Natural Science*, 17(3), 1290 - 1298. <https://doi.org/10.31018/jans.v17i3.6732>

Abstract

Nematodes are vital bioindicators of soil health, reflecting ecological conditions and anthropogenic disturbances, such as tourism. This study provides the first comprehensive assessment of nematode diversity in the Patnitop forest (Jammu & Kashmir, India), evaluating their community structure and ecological significance. Soil samples were collected from multiple forest sites, with nematodes identified to genus level and classified by trophic groups and colonizer-persister (c-p) classes. Diversity indices (Shannon Index, Species Evenness, Maturity Index, Plant Parasitic Index) and ecological indices (Enrichment, Structure, Basal) were computed to assess soil food web conditions. Statistical analysis employed the Kruskal-Wallis test. We identified 37 nematode genera, with bacterivores dominating (58.7%), followed by omnivores (20.58%), herbivores (11%), predators (8.9%), and fungivores (2.7%). The c-p classification showed high proportions of opportunistic c-p 1 nematodes (39.0–48.1%) and stable c-p 4 (19.3–27.6%) and c-p 5 (5.8–15.3%) groups. Moderate diversity (Shannon Index: 1.93; Evenness: 0.73) and nutrient-rich conditions (Maturity Index: 2.50; Plant Parasitic Index: 3.20) were observed. High Enrichment (82.59) and Structure (92.48) indices, combined with a low Basal Index (4.17), indicated a stable food web under minimal stress. Trophic group differences were statistically significant ($p = 0.00129$). The predominance of bacterivores highlights bacterial-mediated decomposition, while nematode metrics demonstrated sensitivity to ecological gradients, underscoring their utility as bioindicators for tourism-linked disturbances. These findings advocate integrating nematode monitoring into forest management frameworks to guide soil conservation in this fragile ecosystem.

Keywords: Colonizer-persister analysis, Ecosystem health, Nematode diversity, Patnitop forest, Soil bio-indicators**INTRODUCTION**

Nematodes, members of the phylum Nematoda, represent a major component of subterranean faunal biodiversity. With approximately 27,000 documented species (Hodda, 2022; Hugot *et al.*, 2001), these organisms occupy nearly every habitable niche on the planet and account for roughly 80% of the taxonomic and functional diversity of soil-dwelling metazoans (van den Hoogen *et al.*, 2019). Soil nematodes, as intact compo-

nents of soil ecosystems, play a crucial role in nutrient cycling, organic matter decomposition, and the regulation of microbial communities (Porazinska *et al.*, 2021; van Bommel *et al.*, 2024 Ferris, 2010; Freckman, 1988). Their functional diversity is central for sustaining soil health and ecosystem reliability, particularly in forested environments (Kumar *et al.*, 2021; Pajak *et al.*, 2016). Nematodes exhibit remarkable ecological adaptability, spanning a spectrum from fast-colonizing r-strategists to persistent K-strategists, with various inter-

mediate forms existing along the colonizer-persister (c-p) continuum (Bongers, 1990; Kergunteuil *et al.*, 2016; Kouser *et al.*, 2021). On account of large trophic and functional diversity, nematodes play a leading role in regulating multiple crucial ecosystem processes including nutrient cycling, successional changes, and energy flow (Rueß, 2024; Yeates, 2003; Bakonyi *et al.*, 2007; Wan *et al.*, 2022). Studies conducted across forest gradients in China (Pang *et al.*, 2024; Shao *et al.*, 2023) have demonstrated that soil attributes, rather than climate variables, are the primary determinants of nematode community structure. Furthermore, nematode-microbe interactions form complex trophic networks that facilitate ecosystem feedback loops essential to soil health (Ruess, 2024). Nematodes are abundant soil metazoans and reliable bioindicators, reflecting ecological shifts through their sensitivity to biotic and abiotic factors (Asif *et al.*, 2021; Van Eekeren *et al.*, 2010). Their community structure is shaped more by soil properties—such as pH, moisture, and organic content—than by climatic gradients (Shao *et al.*, 2023; Pang *et al.*, 2024). Nematode-based indices have emerged as effective tools in soil ecological studies, offering insights into ecosystem condition, stress, and sustainability (Du Preez *et al.*, 2022). A number of researchers have worked on the nematode diversity in Indian forests such as natural forests of Arunachal Pradesh (Baniyamuiddin *et al.* 2007), Sal forests of Dehradun (Rizvi, 2008), and DKG forest of district Poonch, Jammu and Kashmir (Vaid *et al.* 2014). Patnitop (32.99°N, 75.23°E; elevation: 2,024 m), a region within Jammu and Kashmir's Udhampur district, is characterized by alpine meadows and high floristic diversity. Its mixed coniferous and broadleaf vegetation, coupled with its elevation-driven microclimates, renders it ecologically significant for studying Himalayan temperate ecosystems. The region is home to a range of coniferous trees such as *Cedrus deodara*, *Pinus wallichiana*, and *Abies pindrow*. This varied plant life spawns a fitting environment for studying nematode diversity in the region. The aim of the present study was (i) to determine the soil nematode diversity, functional role and community structure (ii) to assess ecosystem health and stability using ecological indices.

MATERIALS AND METHODS

Collection of samples

A survey was conducted in May and June 2023 to examine the diversity and community structure of soil nematodes in Patnitop Forest (33.08°N, 75.33°E). Soil samples were randomly collected using a trowel from various locations within the forest, totalling fifty samples. Each sample was placed in an airtight plastic bag to preserve moisture, labelled with site details, collection dates, and recorded temperatures, and then trans-

ported to the laboratory for further analysis.

Nematode extraction and processing

Soil samples were processed using Cobb's sieving-decantation method followed by modified Baermann funnel extraction. Nematodes were fixed in heated (65° C) formalin-acetic acid (4:1), dehydrated through a graded glycerine-ethanol series (5% glycerine in 30% ethanol), and permanently mounted in anhydrous glycerine using established protocols (Seinhorst, 1959). Specimen integrity was verified by phase-contrast microscopy prior to analysis.

Nematode counting

Nematode abundance was quantified by preparing microscopic slides for genus-level identification. Additionally, a Syracuse counting dish was utilized to enhance counting accuracy.

Community analysis: Nematode diversity and ecological distribution were assessed using the following metrics:

Frequency (N): The number of samples in which a specific genus was detected.

Absolute Frequency (AF %): The proportion of samples containing a given genus relative to the total samples analyzed, expressed as:

$$AF = \text{Genus frequency} / \text{Total number of sample counted} \times 100 \quad (\text{Eq. 1})$$

Relative Frequency (RF) The occurrence of a genus compared to the cumulative frequency of all genera, calculated as:

$$RF = \text{Frequency of genus} / \text{Total frequency of all genera} \times 100 \quad (\text{Eq. 2})$$

Mean density (MD): The average number of individuals of a genus across all samples, determined by:

$$MD = \text{Total number of individuals of a particular genus counted in all samples} / \text{total number of sample counted} \quad (\text{Eq. 3})$$

Relative density (RD %): The mean density of a genus relative to the sum of mean densities for all genera, computed as (Tomar *et al.* 2006):

$$RD = \text{Mean density of particular genus} / \text{Sum of mean density of all nematode genera} \times 100 \quad (\text{Eq. 4})$$

Statistical analysis

Frequency (N), Absolute frequency (AF %), Density (MD), Relative density (RD%), were calculated following Tomar and Ahmad, 2009, in Microsoft Office excel version 2021. Nematode specific indices like the Maturity index (MI), Channel Index (CI), Structure Index (SI), Plant Parasitic Index (PPI), and Enrichment Index (EI) (Ferris *et al.*, 2001) were calculated using NINJA: Nematode Indicator Joint Analysis accessed on 31/07/2024, (Sieriebriennikov *et al.*, 2014). Pie Chart, Shannon diversity (H') and evenness (R software). Graph for food web analysis and c-p triangle

were plotted using NINJA: Nematode Indicator Joint Analysis. A Kruskal-Wallis test followed by Dunn's post-hoc test was carried out to determine if there was any significance in abundance among trophic groups using R software, version 4.4.1 (R Core Team, 2025).

Nematode identification process

Following fixation and dehydration, the nematodes were preserved as permanent slides using the paraffin wax ring method. These slides were then examined under an Olympus BX51 DIC microscope for detailed study. Taxonomic identification of the extracted nematodes was carried out by referencing key taxonomic works (Andrássy, 1983; Jairajpuri and Ahmad, 1992; Ahmad, 1996), enabling classification up to the genus level.

RESULTS AND DISCUSSION

Nematode community patterns and ecological distribution

In the nematode community diversity study conducted in Patnitop, 37 nematode genera were identified. Bacterivores were the most prevalent, representing 58.7% of the total genera, followed by omnivores (20.58%), herbivores (11%), predators (8.9%), and fungivores (2.7%) (Fig 1 and 2). This outcome aligns with the findings of previous studies (Dong et al., 2017; Kashyap et al., 2022). This suggests that a faster, bacteria-driven decomposition process is dominant in the study area (Sushchuk and Matveeva, 2021; Vaid et al., 2014; Nisa et al., 2021). Herbivorous nematodes varied based on parasitism type, with migratory endoparasites (e.g., *Pratylenchus* spp.), ectoparasites (e.g., *Longidorus* spp.), sedentary parasites (e.g., *Heterodera* spp.), and root hair feeders (e.g., *Tylenchus* spp.) present across all sampled localities.

Nematodes were categorized using colonizer-persister (c-p) values for free-living forms (Table 1). C-p 1 nematodes dominated (39.0%–48.1%), indicating a high pro-

portion of opportunistic species, which are often associated with disturbed or enriched environments (Ettema and Bongers, 1993; Kemenju et al., 2009) (Fig. 2). C-p 2 nematodes were less abundant (9.9%–16.1%), while c-p 3 nematodes showed moderate presence (7.7%–11.4%). Higher c-p groups (c-p 4 and c-p 5) were also recorded, with c-p 4 nematodes representing 19.3%–27.6% and c-p 5 nematodes comprising 5.8%–15.3% of the community (Fig. 2). The presence of higher c-p groups suggests some level of ecosystem stability, as these groups are associated with slower growth rates and longer life cycles, often indicating a more resilient and balanced ecosystem (Ettema and Bongers, 1993; Preez et al., 2022).

Population structure of nematode genera

Among bacterivorous nematodes, *Mesorhabditis* was the most frequent genus (Absolute Frequency, AF = 21.06%), followed by *Acrobeles* (AF = 14.99%) and *Diplogasteriana* (AF = 14.85%). *Eucephalobus* was the least frequent (AF = 0.69%). For omnivorous nematodes, *Dorylaimus* (AF = 14.85%) and *Aporcelaimus* (AF = 17.81%) were the most frequent, while *Moshajia* was the least frequent (AF = 0.77%). Among plant-parasitic nematodes, *Pratylenchus* had the highest AF (3.89%), and *Tylenchus* was the least frequent (AF = 0.52%). Predatory nematodes were dominated by *Mononchus* (AF = 3.82%), with *Crassolabium* being the least frequent (AF = 0.58%). Among fungivorous nematodes, *Aphelenchus* was the most frequent (AF = 1.21%), while *Doryllium* and *Nothotylenchus* were the least frequent (AF = 0.77% and 0.37%, respectively) (Table 1).

In terms of dominance, *Mesorhabditis* (Mean Density, MD = 21.06%; Relative Density, RD = 10.93%) was the most dominant bacterivorous genus, while *Eucephalobus* (MD = 0.69%; RD = 0.36%) was the least dominant. Among omnivores, *Aporcelaimus* (MD = 17.81%; RD = 9.25%) was the most dominant, and *Moshajia* (MD = 0.77%; RD = 0.40%) was the least dominant. For plant-parasitic nematodes, *Pratylenchus* (MD = 3.89%; RD = 2.02%) was the most dominant, while *Tylenchus* (MD = 0.52%; RD = 0.27%) was the least dominant. Among predators, *Mononchus* (MD = 3.82%; RD = 1.98%) was the most dominant, and *Crassolabium* (MD = 0.58%; RD = 0.30%) was the least dominant. For fungivores, *Aphelenchus* (MD = 1.21%; RD = 0.63%) was the most dominant (Table 1).

Functional Indices

Functional indices provided insight into the health and functional dynamics of the soil ecosystem (Table 2, Fig. 4). The Shannon Diversity Index was 1.93, with a Species Evenness of 0.73, indicating a moderately diverse and evenly distributed nematode community. This suggests a well-distributed variety of species,

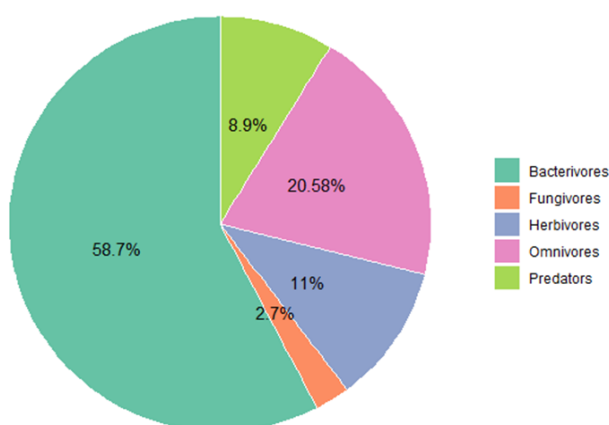


Fig.1. Nematode generic diversity

Table 1. Population structure of soil-inhabiting nematodes in Patnitop forest

Nematode Genera	C-p value	F	AF	RF	MD	RD
Bacterivores						
<i>Acrobeles</i>	2	21	56.7568	3.79747	8.66	4.49637
<i>Bursilla</i>	1	11	29.7297	1.98915	12.69	6.58879
<i>Cephalobus</i>	2	16	43.2432	2.89331	4.4	2.28453
<i>Diplogasteriana</i>	1	21	56.7568	3.79747	14.99	7.78297
<i>Eucephalobus</i>	2	9	24.3243	1.62749	0.69	0.35826
<i>Mesorhabditis</i>	1	17	45.9459	3.07414	21.06	10.9346
<i>Panagrolaimus</i>	1	7	18.9189	1.26582	2.52	1.30841
<i>Plectus</i>	2	12	32.4324	2.16998	4.41	2.28972
<i>Protorhabditis</i>	1	12	32.4324	2.16998	12.97	6.73416
<i>Rhabditis</i>	1	19	51.3514	3.4358	13.06	6.78089
<i>Teratocephalus</i>	3	14	37.8378	2.53165	17.16	8.90966
Omnivores						
<i>Aporcelaimus</i>	5	19	51.3514	3.4358	17.81	9.24714
<i>Dorylaimus</i>	4	16	43.2432	2.89331	14.85	7.71028
<i>Mesodorylaimus</i>	4	18	48.6486	3.25497	3.53	1.83281
<i>Metaporcelaimus</i>	5	8	21.6216	1.44665	1.97	1.02285
<i>Moshajia</i>	4	14	37.8378	2.53165	0.77	0.39979
Herbivores						
<i>Helicotylenchus</i>	0	20	54.0541	3.61664	3.8	1.973
<i>Hemicriconemoides</i>	0	20	54.0541	3.61664	1.45	0.75286
<i>Mesocriconema</i>	0	19	51.3514	3.4358	3.06	1.58879
<i>Paralongidorus</i>	0	15	40.5405	2.71248	3.13	1.62513
<i>Pratylenchus</i>	0	19	51.3514	3.4358	3.89	2.01973
<i>Psilenchus</i>	0	19	51.3514	3.4358	1.85	0.96054
<i>Rotylenchus</i>	0	15	40.5405	2.71248	1.13	0.58671
<i>Tylenchus</i>	0	11	29.7297	1.98915	0.52	0.26999
<i>Xiphinema</i>	0	15	40.5405	2.71248	3.2	1.66147
Predators						
<i>Crassolabium</i>	4	12	32.4324	2.16998	0.58	0.30114
<i>Discolaimus</i>	4	8	21.6216	1.44665	2.29	1.18899
<i>Eudorylaimus</i>	4	19	51.3514	3.4358	2.51	1.30322
<i>Labronema</i>	4	10	27.027	1.80832	0.6	0.31153
<i>Microdorylaimus</i>	4	14	37.8378	2.53165	0.86	0.44652
<i>Mononchus</i>	4	18	48.6486	3.25497	3.82	1.98339
<i>Nygolaimoides</i>	5	18	48.6486	3.25497	2.24	1.16303
<i>Prionchulus</i>	4	19	51.3514	3.4358	2.6	1.34995
Fungivores						
<i>Aphelenchus</i>	2	14	37.8378	2.53165	1.21	0.62825
<i>Doryllium</i>	4	11	29.7297	1.98915	0.77	0.39979
<i>Nothotylenchus</i>	2	11	29.7297	1.98915	0.37	0.19211
<i>Tylencholaimus</i>	4	12	32.4324	2.16998	1.18	0.61267

C-p value = Colonizer-persister value, N = Frequency, AF = Absolute frequency, RF = Relative frequency, MD = Mean density, RD = Relative density

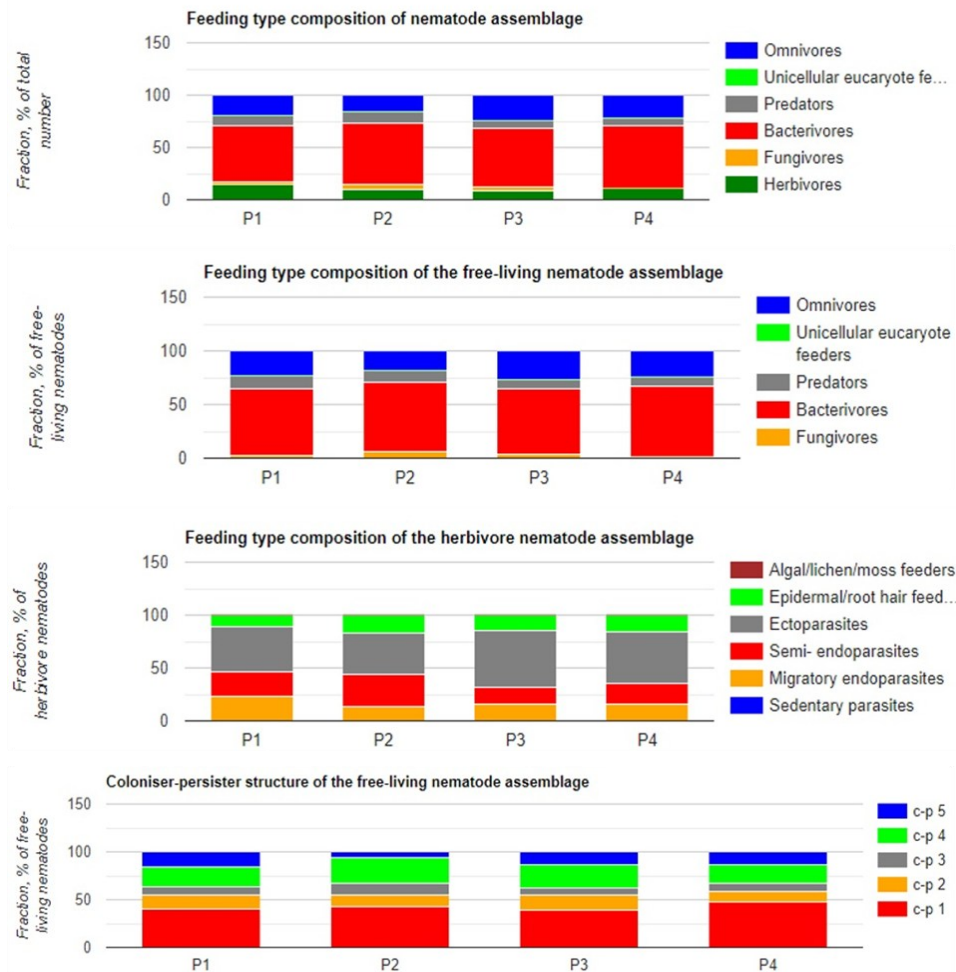


Fig 2. Proportion of soil nematode communities that occurred in Patnitop forest of district Udhamapur

which is essential for fostering ecosystem functions such as nutrient cycling and the stability of food webs. Studies have shown that nematode diversity and abundance can vary significantly with elevation. For instance, in the subtropical forests of Huangshan Mountain, nematode diversity increased with elevation, likely due to increased soil moisture (Ding *et al.*, 2024). The Species Evenness of 0.73 indicates a relatively even distribution of species, which is consistent with findings from the Mezin National Nature Park, where nematode communities showed varied distribution across different habitats (Жиліна and Шевченко, 2024). The Maturity Index (MI), recorded at 2.50, reflects a community skewed toward colonizer (r-strategist) nematodes, suggesting moderate environmental disturbance and a low successional status (Bongers and Ferris, 1999). Complementing this, the Enrichment Index (EI = 82.59) indicates a highly enriched system, likely influenced by recent organic inputs or nutrient flushes, which tend to promote bacterial-feeding nematodes (Shaw *et al.*, 2019; Tomar and Ahmad, 2009). Despite signs of enrichment, the Structure Index (SI = 92.48) indicates a well-developed, complex food web, characteristic of a functionally stable and structured community, a pattern

consistent with old-growth or mature forest soils (Förster *et al.*, 2024; Kitagami and Matsuda, 2024). The Plant Parasitic Index (PPI = 3.20) indicates a moderate herbivorous load, characterized by 11% herbivorous nematode abundance, a level typically observed in stable forest ecosystems (Neher, 2001; Ferris, 2010). Furthermore, the low Basal Index (BI = 4.17) and Channel

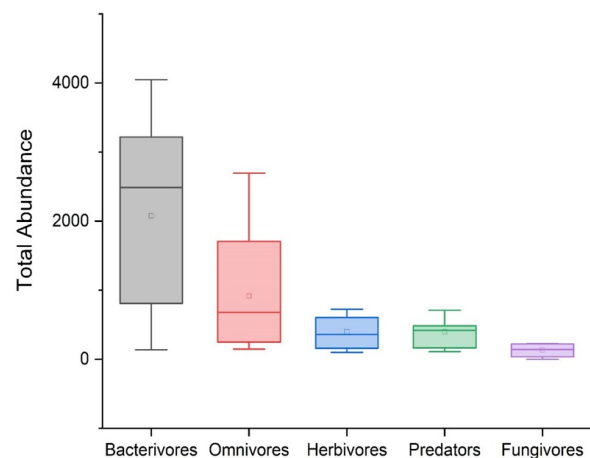


Fig. 3. Total mean abundance of different trophic groups studied in Patnitop forest

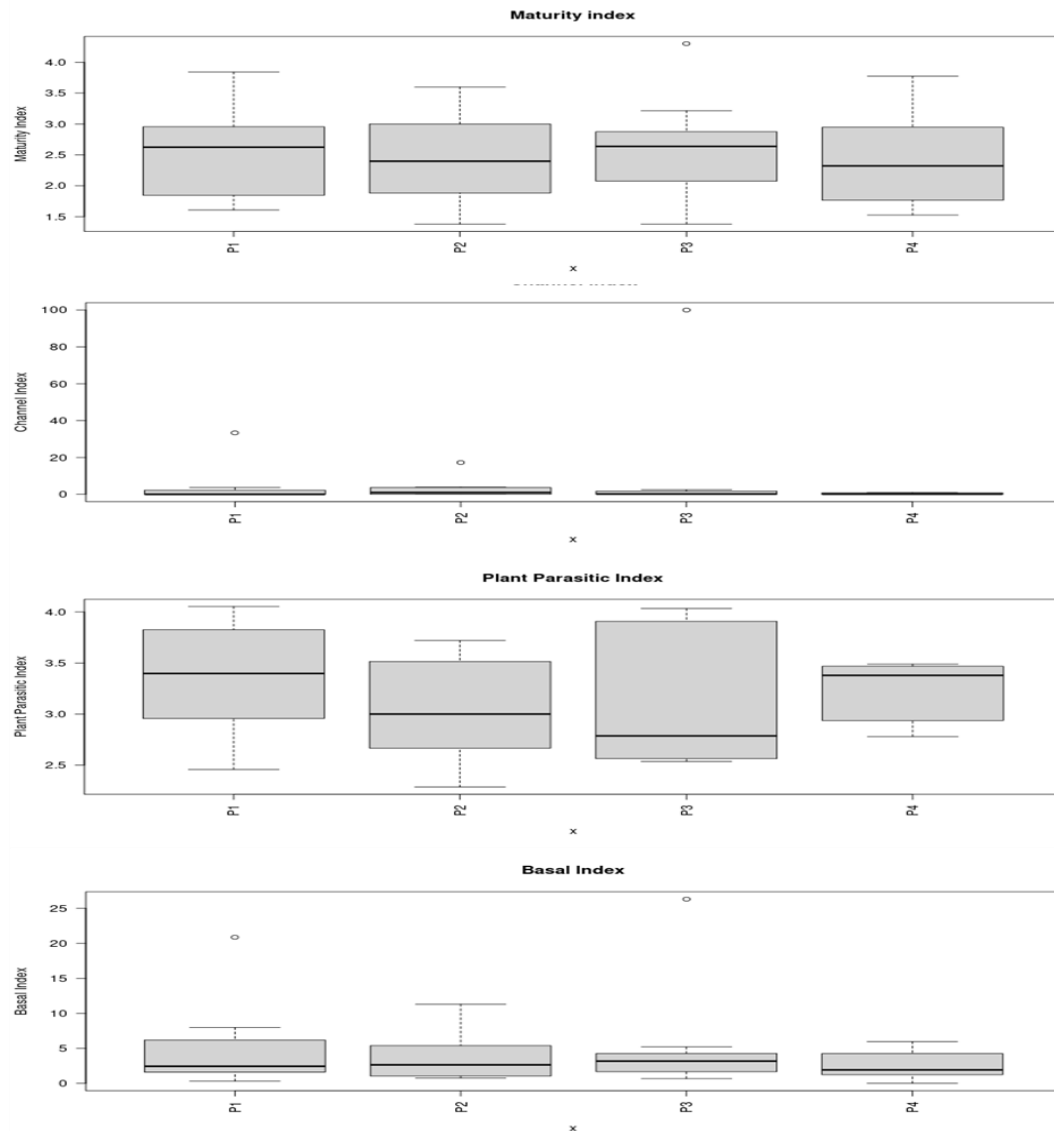


Fig. 4. Soil nematode Functional indices in Patnitop forest (MI (Maturity Index), CI (channel Index), PPI (Plant Parasitic Index), BI (Basal index))

Index (CI = 4.57) denote a minimal presence of stress-tolerant taxa and a dominant bacterial decomposition pathway, respectively. Collectively, these indices confirm that while nutrient enrichment is evident, the nematode community retains structural integrity and functional resilience, highlighting a balanced yet dynamic forest soil environment (Fig. 4). A Kruskal-Wallis test revealed significant differences among trophic groups (Kruskal-Wallis chi-squared = 17.902, df = 4, p-value = 0.00129). Dunn's post-hoc test showed that bacterivores differed significantly in abundance compared to fungivores, herbivores, and predators (Fig. 3). This highlights the functional diversity within the nematode community and their roles in nutrient cycling and ecosystem stability.

Nematodes as potential soil bio-indicators

Nematodes were assessed as bioindicators of soil health using food web diagnostics (Fig. 6). Over 80% of

the samples were characterized by fertile or suppressive soils with moderate C:N ratios and bacterial-fungal properties. A c-p triangle analysis (Fig. 5) indicated that most soil samples were close to enrichment (c-p 1) and stability conditions (c-p 3–5). The predominance of c-p 1 nematodes suggests a soil environment undergoing disturbance or enrichment, potentially due to human activities, such as tourism, or natural disturbances. In contrast, cp-4 represents stress-tolerant persisters, characterized by slower growth rates and longer life cycles, which often suggest a more resilient and balanced ecosystem. The present findings indicate that the prevalence of cp-1 nematodes suggests a soil environment experiencing either disturbance or enrichment. This may be attributed to human activities, particularly tourism, given that Patnitop is recognized as a prime tourist destination, or it could be a result of natural dis-

Table 2. Summary of nematode diversity indices in Patnitop forest

S.No.	Ecological Indices	Values
1	Shannon Diversity	1.93
2	Species Evenness	0.73
3	Maturity Index	2.50
4	Plant Parasitic Index	3.20
5	Channel Index	4.57
6	Basal Index	4.17
7	Enrichment Index	82.59
8	Structure Index	92.48

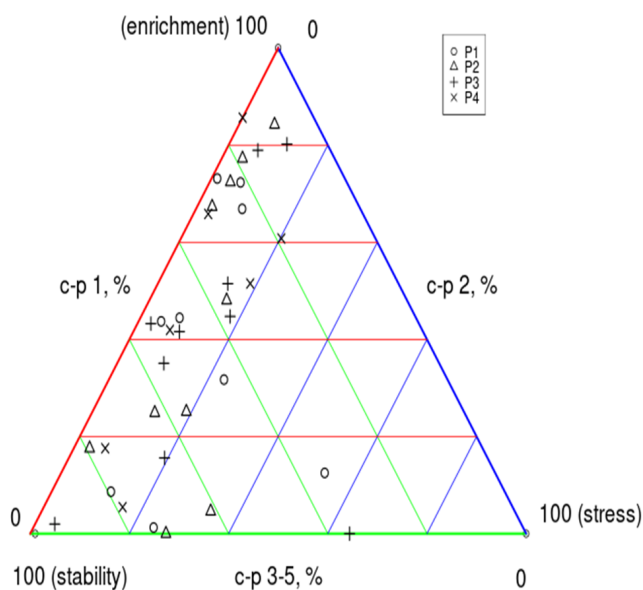
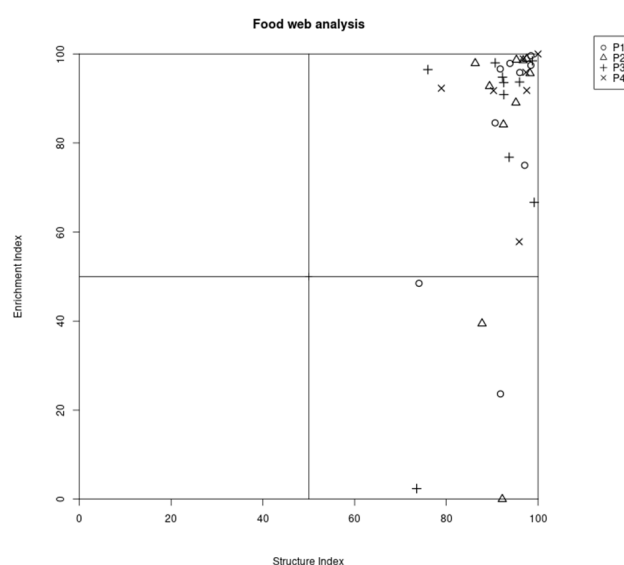
turbances. Supporting this, recent studies have shown that tourist trampling in forest ecosystems significantly alters soil microbial communities, often promoting bacterial growth and shifting community composition toward human-associated taxa (Shang *et al.*, 2022; Li *et al.*, 2022). Such microbial shifts have been linked to increased nitrogen mineralization and nutrient cycling, potentially creating favorable conditions for colonizer-persister group 1 (c-p 1) nematodes. These microbial and nutrient changes, induced by tourism-driven disturbance, may partially explain the observed nematode community structure. Conversely, the presence of cp-4 nematodes suggests a more stable and mature ecosystem, characterized by a balanced nutrient cycle and strong soil health (Ettema and Bongers, 1993; Du Preez *et al.*, 2022).

Conclusion

The study highlights the dominance of bacterivorous nematodes in the Patnitop forest, suggesting that bacteria play a significant role in decomposing organic matter. The presence of both opportunistic (c-p 1) and stable (c-p 4–5) nematode groups suggests a dynamic soil environment influenced by natural and anthropogenic factors. The functional indices, including the Shannon Diversity Index, Maturity Index, and Enrichment Index, offer valuable insights into the health and stability of the soil ecosystem. Nematodes serve as effective bio-indicators of soil health, reflecting the impacts of disturbances and nutrient dynamics. These findings underscore the importance of sustainable forest management and long-term monitoring to maintain ecosystem stability and biodiversity. Future research should integrate nematode-based indicators with other soil health metrics to enhance conservation strategies and promote the sustainability of forest ecosystems.

Conflict of interest

The authors declare that they have no conflict of interest.

**Fig. 5.** c-p (colonizer-persister) triangle depicting soil status regarding nematode c-p groups' evolution in Patnitop forest**Fig. 6.** Food web diagnostic profile

REFERENCES

- Ahmad, W. (1996). Plant parasitic nematodes of India. *Aligarh. India*.
- Andrássy, I. (1983). A taxonomic review of the suborder *Rhabditina* (Nematoda: Secernentia) (p. 241pp).
- Asif, M., Jahan, R., & Mahboob, M. (2021). Biological indicative assessment of nematodes in evaluating different terrestrial habitats.
- Bakonyi, G., Nagy, P., Kovacs-Lang, E., Kovacs, E., Barabas, S., Répási, V., & Seres, A. (2007). Soil nematode community structure as affected by temperature and moisture in a temperate semiarid shrubland. *Applied Soil Ecology*, 37(1-2), 31-40.
- Bongers, T., & Ferris, H. (1999). Nematode community structure as a bioindicator in environmental monitor-

- ing. *Trends in Ecology & Evolution*, 14(6), 224-228.
6. Bongers, T. (1990). The maturity index: an ecological measure of environmental disturbance based on nematode species composition. *Oecologia*, 83, 14-19.
 7. Ding, K., Qiang, Z., Hu, Z., Cheng, S., Sun, R., Fang, H., & Ma, C. (2024). Elevational Gradients of Soil Nematode Communities in Subtropical Forest Ecosystems. *Forests*, 15(12), 2149.
 8. Dong, K., Moroenyane, I., Tripathi, B., Kerfahi, D., Takahashi, K., Yamamoto, N., & Adams, J. (2017). Soil nematodes show a mid-elevation diversity maximum and elevational zonation on Mt. Norikura, Japan. *Scientific Reports*, 7(1), 3028.
 9. Du Preez, G., Daneel, M., De Goede, R., Du Toit, M. J., Ferris, H., Fourie, H., & Schmidt, J. H. (2022). Nematode-based indices in soil ecology: Application, utility, and future directions. *Soil Biology and Biochemistry*, 169, 108640.
 10. Ettema, C. H., & Bongers, T. (1993). Characterization of nematode colonization and succession in disturbed soil using the Maturity Index. *Biology and Fertility of Soils*, 16, 79-85.
 11. Ettema, C. H., & Bongers, T. (1993). Characterization of nematode colonization and succession in disturbed soil using the Maturity Index. *Biology and Fertility of Soils*, 16, 79-85.
 12. Ferris, H., Bongers, T., & de Goede, R. G. (2001). A framework for soil food web diagnostics: extension of the nematode faunal analysis concept. *Applied soil ecology*, 18(1), 13-29.
 13. Ferris, H. (2010). Contribution of nematodes to the structure and function of the soil food web. *Journal of nematology*, 42(1), 63.
 14. Ferris, H. (2010). Form and function: metabolic footprints of nematodes in the soil food web. *European Journal of Soil Biology*, 46(2), 97-104.
 15. Ferris, H., & Benavides, I. V. (2024). Opinions and Suggestions on Nematode Faunal Analysis. *Journal of nematology*, 56(1).
 16. Förster, A., Hohberg, K., Rasche, F., & Emmerling, C. (2024). Nematode community structure suggests perennial grain cropping cultivation as a nature-based solution for resilient agriculture. *Journal of Sustainable Agriculture and Environment*, 3(3), e12112.
 17. Freckman, D. W. (1988). Bacterivorous nematodes and organic-matter decomposition. *Agriculture, Ecosystems & Environment*, 24(1-3), 195-217.
 18. Hugot, J. P., Baujard, P., & Morand, S. (2001). Biodiversity in helminths and nematodes as a field of study: an overview. *Nematology*, 3(3), 199-208.
 19. Hodda, M. (2022). Phylum Nematoda: a classification, catalogue and index of valid genera, with a census of valid species. *Zootaxa*, 5114(1), 1-289.
 20. Jairajpuri, M. S., & Ahmad, W. (1992). *Dorylaimida: free-living, predaceous and plant-parasitic nematodes*. Brill.
 21. JW, K., NK, K., GK, M., BM, R., & PM, W. (2009). Nematode community structure as influenced by land use and intensity of cultivation.
 22. Kashyap, P., Afzal, S., Rizvi, A. N., Ahmad, W., Uniyal, V. P., & Banerjee, D. (2022). Nematode community structure along elevation gradient in high altitude vegetation cover of Gangotri National Park (Uttarakhand), India. *Scientific reports*, 12(1), 1428.
 23. Kergunteuil, A., Campos-Herrera, R., Sánchez-Moreno, S., Vittoz, P., & Rasmann, S. (2016). The abundance, diversity, and metabolic footprint of soil nematodes is highest in high elevation alpine grasslands. *Frontiers in Ecology and Evolution*, 4, 84.
 24. Kimenju, J. W., Karanja, N. K., Mutua, G. K., Rimberia, B. M., & Wachira, P. M. (2009). Nematode community structure as influenced by land use and intensity of cultivation. *Tropical and subtropical agroecosystems*, 11(2), 353-360.
 25. Kitagami, Y., & Matsuda, Y. (2023). Distribution and characterization of nematodes in above-ground microhabitats in a natural pristine cedar forest in Yakushima Island, Japan. *Canadian Journal of Zoology*, 102(3), 264-271.
 26. Kouser, Y., Shah, A. A., & Rasmann, S. (2021). The functional role and diversity of soil nematodes are stronger at high elevation in the lesser Himalayan Mountain ranges. *Ecology and evolution*, 11(20), 13793-13804.
 27. Li, Q., Dai, M., & Luo, F. (2022). Influence of tourism disturbance on soil microbial community structure in Dawei Mountain national forest park. *Sustainability*, 14(3), 1162.
 28. Neher, D. A. (2001). Role of nematodes in soil health and their use as indicators. *Journal of nematology*, 33(4), 161.
 29. Nisa, R. U., Tantray, A. Y., Kouser, N., Allie, K. A., Wani, S. M., Alamri, S. A., ... & Shah, A. A. (2021). Influence of ecological and edaphic factors on biodiversity of soil nematodes. *Saudi journal of biological sciences*, 28(5), 3049-3059.
 30. Porazinska, D. L., De Mesquita, C. P. B., Farrer, E. C., Spasojevic, M. J., Suding, K. N., & Schmidt, S. K. (2021). Nematode community diversity and function across an alpine landscape undergoing plant colonization of previously unvegetated soils. *Soil Biology and Biochemistry*, 161, 108380.
 31. Pająk, M., Błońska, E., Frąc, M., & Oszust, K. (2016). Functional diversity and microbial activity of forest soils that are heavily contaminated by lead and zinc. *Water, Air, & Soil Pollution*, 227, 1-14.
 32. Pang, S., Hua, B., Yang, W., Zhang, S., Guan, Y., Bai, K., & Zhang, X. (2024). Soil properties override climatic factors to shape soil nematode diversity in the eastern forest transect of China. *Global Ecology and Conservation*, 54, e03061.
 33. Ruess, L. (2024). Nematodes and their trophic interactions in the soil microbiome. *Understanding and utilizing soil microbiomes for a more sustainable agriculture*, 29.
 34. R Development Core Team. (2015). R: A language and environment for statistical computing. R Foundation for Statistical Computing. <https://www.R-project.org/>
 35. Seinhorst, J. W. (1959). A rapid method for the transfer of nematodes from fixative to anhydrous glycerin.
 36. Shang, Q., Liu, Y., & Li, Q. (2022). Effects of tourism trampling on soil nitrogen mineralization in *Quercus variabilis* Blume forests varies with altitudes in the climate transition zone. *Forests*, 13(9), 1467.
 37. Shao, Y., Wang, Z., Liu, T., Kardol, P., Ma, C., Hu, Y., & Fu, S. (2023). Drivers of nematode diversity in forest soils across climatic zones. *Proceedings of the Royal Society B*, 290(1994), 20230107.
 38. Shaw, E. A., Boot, C. M., Moore, J. C., Wall, D. H., & Baron, J. S. (2019). Long-term nitrogen addition shifts the soil

- nematode community to bacterivore-dominated and reduces its ecological maturity in a subalpine forest. *Soil Biology and Biochemistry*, 130, 177-184.
39. Sieriebriennikov, B., Ferris, H., & de Goede, R. G. (2014). NINJA: An automated calculation system for nematode-based biological monitoring. *European Journal of Soil Biology*, 61, 90-93.
40. Sushchuk, A. A., & Matveeva, E. M. (2021). Soil nematodes of coniferous forests in the Finnish-Russian Friendship Nature Reserve. *Nature Conservation Research. Заповедная наука*, 6(S1), 76-88.
41. Tomar, V. V. S., & Ahmad, W. (2009). Food web diagnostics and functional diversity of soil inhabiting nematodes in a natural woodland. *Helminthologia*, 46, 183-189.
42. Tomar, V. V. S., Mohammad Baniyammuddin, M. B., & Wasim Ahmad, W. A. (2006). Community structure of soil inhabiting nematodes in a mango orchard at Aligarh, India.
43. Van Den Hoogen, J., Geisen, S., Routh, D., Ferris, H., Traunspurger, W., Wardle, D. A., & Crowther, T. W. (2019). Soil nematode abundance and functional group composition at a global scale. *Nature*, 572(7768), 194-198.
44. Vaid, S., Shah, A. A., Ahmad, R., & Hussain, A. (2014). Diversity of soil inhabiting nematodes in Dera Ki Gali forest of Poonch district, Jammu and Kashmir, India. *International Journal of Nematology*, 24(1), 97-102.
45. van Bommel, M., Arndt, K., Endress, M. G., Dehghani, F., Wirsching, J., Blagodatskaya, E., & Ruess, L. (2024). Under the lens: Carbon and energy channels in the soil micro-food web. *Soil Biology and Biochemistry*, 199, 109575.
46. Van Eekeren, N., De Boer, H., Hanegraaf, M., Bokhorst, J., Nierop, D., Bloem, J., & Brussaard, L. (2010). Ecosystem services in grassland associated with biotic and abiotic soil parameters. *Soil biology and biochemistry*, 42(9), 1491-1504.
47. Wan, B., Liu, T., Gong, X., Zhang, Y., Li, C., Chen, X., & Liu, M. (2022). Energy flux across multitrophic levels drives ecosystem multifunctionality: Evidence from nematode food webs. *Soil Biology and Biochemistry*, 169, 108656.
48. Yeates, G. W. (2003). Nematodes as soil indicators: functional and biodiversity aspects. *Biology and Fertility of soils*, 37, 199-210.