

Research Article

Protective role of plant-based pollen substitute diets against *Nosema* spores in *Apis mellifera* colonies

Ashish R. Gawali*

Department of Zoology, Dr. Babasaheb Ambedkar Marathwada University, Chhatrapati Sambhajinagar (Aurangabad)- 431004 (MS), India

Bhalchandra B. Waykar

Department of Zoology, Dr. Babasaheb Ambedkar Marathwada University, Chhatrapati Sambhajinagar (Aurangabad)- 431004 (MS), India

*Corresponding author. Email: ashishgawali8888@gmail.com

Article Info

https://doi.org/10.31018/ jans.v17i1.6188 Received: September 14, 2024 Revised: February 09, 2025 Accepted: February 15, 2025

How to Cite

Gawali, A.R. and Waykar, B. B. (2025). Protective role of plant-based pollen substitute diets against *Nosema* spores in *Apis mellifera* colonies. *Journal of Applied and Natural Science*, 17(1), 193 - 199. https://doi.org/10.31018/jans.v17i1.6188

Abstract

Nosema sp. causes a significant threat to honeybee populations, making it crucial to find effective mitigation strategies. The present study examines the impact of plant-based pollen substitute diets on *Nosema* spore infection in *Apis mellifera* L. colonies. The study explores the potential of phytochemicals, which have antimicrobial properties, as natural treatments to reduce *Nosema* infection. Seven experimental groups (Diet-2 to Diet-8 along with Diet-1 (control) of colonies were fed with different pollen substitute diets containing medicinal plant leaves such as coriander (*Coriandrum sativum*), moringa (*Moringa oleifera*), tulsi (*Ocimum sanctum*) and lemongrass (*Cymbopogon citratus*); fruit powders of amla (*Embilica officinalis*), guava (*Psidium guava*) and mixed diet containing equal quantities of all plant leaves and fruit powders, along with common ingredients like defatted soya flour, skimmed milk powder, brewer's yeast, turmeric, vitamins and sugar syrup. The Diet-2 to Diet-8 were compared to a control group of colonies (Diet-1), which was only fed with sugar syrup. *Nosema* spores were counted in samples of 40 worker bees from experimental and control groups of colonies before feeding and at intervals of 8, 16, 24 and 32 days using an Improved Neubauer hemocytometer. Results showed that Diet-8 has significantly reduced the number of *Nosema* spores (50,000±28867.5 spores/bee) compared to the control group of colonies (5,50,000±28867.5 spores/bee). Diet-8 was more effective than other diets, suggesting that plant-based pollen substitute diets can help effectively manage Nosema infections and maintain healthy and disease-free honeybee colonies throughout the year.

Keywords: Apis mellifera L, Medicinal plants, Microsporidian parasite, Phytochemicals, Pollen substitute diet

INTRODUCTION

The Western honeybee *Apis mellifera* L. is a vital pollinator, contributing billions of dollars to global agricultural production annually (Kulhanek *et al.*, 2021). However, honeybee populations have declined in recent decades, largely attributed to a combination of factors, including pesticide exposure, infectious diseases and habitat loss (Kulhanek *et al.*, 2021). One of the most significant pathogens affecting honeybees is the microsporidian parasite *Nosema ceranae*, which can severely impact individual bee health and colony survival (Jousse *et al.*, 2020). *Nosema* infections disrupt gut function, reduce lifespan and impair colony-level behaviours critical for pollination services (Jousse *et al.*, 2020).

Nosema is a microsporidian, a taxonomic category of

single-celled protozoan parasites that infect a variety of animal hosts. Honeybees develop the disease by swallowing Nosema spores via food sharing or consuming faeces from an infected nestmate. Once ingested, the Nosema replicates inside midgut (stomach) cells and seizes the nutrition of the honeybee. The midgut cell membrane eventually ruptures due to the Nosema spore's subsequent rapid multiplication. This releases more spores into the honeybee's midgut and increases the likelihood that the disease will be transmitted to other bees. The symptoms of Nosema apis infection are, large numbers of dead bees occurring in the colony and diarrhoea stains at entrances of the hive, indicating gastrointestinal disorders (Mortensen et al., 2020; Bourgeois et al., 2010; Araneda et al., 2015). In temperate areas, N. apis infections generally peak in the spring, reduce during the summer and rise again in

This work is licensed under Attribution-Non Commercial 4.0 International (CC BY-NC 4.0). © : Author (s). Publishing rights @ ANSF.

the fall before lowering in the early winter (Higes *et al.*, 2010). The prevalence of Nosemosis varies with location and season of year (Mulholland *et al.*, 2012). The lifespans of infected honeybees are reduced and they frequently exhibit extreme hunger, i.e., persistently requesting to be fed by other adult honeybees (Mayack and Naug, 2009). Because *Nosema* infections may not be found outside until the colony is significantly depleted, beekeepers may find the danger of infection particularly distressing.

Beekeepers commonly provide pollen supplements to bee colonies when they believe that bees suffer from a lack of nutrition or the incoming resources are of poor or inadequate quality. For example, pollen diets, which often contain a protein source derived from soy, wheat, brewers' yeast, milk powder and legumes enriched with vital vitamins, are frequently fed to migratory colonies when they are utilized to pollinate crops. These diets significantly contribute to the colony's protein requirements (DeGrandi-Hoffman *et al.*, 2010).

Medicinal plants having antimicrobial properties have been explored as potential natural treatments for *Nosema* (Alberoni *et al.,* 2023). An alternative strategy is to investigate the usage of plant-based pollen substitutes as a dietary intervention to boost honeybee immunity and resistance to *Nosema* (Daisley *et al.,* 2020).

Selected medicinal plants have essential phytochemicals and other protein-rich ingredients for inhibiting the growth of *Nosema* spores. Phytochemicals play an important role in honeybee health, and plant-derived compounds may be able to inhibit *Nosema* spore development (Daisley *et al.*, 2020). Phytochemicals such as caffeic, gallic, p-coumaric acids, quercetin, thymol, resveratrol and kaempferol most effectively prolonged the honeybee's lifespan (Bernklau *et al.*, 2019).

Thymol inhibits the growth of pathogenic bacteria and fungi (Rice, 2001; Yücel & Doğaroğlu, 2005). Resveratrol is a phytoalexin produced by certain plants in response to infections caused by phytopathogenic microorganisms (Fremont, 2000; Prokhoda *et al.*, 2019). The present study aimed to develop plant-based pollen substitute diets for controlling of *Nosema* spore infection in *Apis mellifera* L. colonies.

MATERIALS AND METHODS

The study was conducted at Dr. Babasaheb Ambedkar Marathwada University, Campus, Chhatrapati Sambhajinagar. Feeding experiments were carried out on honeybee colonies (*Apis mellifera* L). Plant-based pollen substitute diets were fed to honeybee colonies from July 01, 2023, to August 10, 2023. Temperature ranges from 20.0°C to 30.0°C, while humidity ranges from 73.8% to 96.3%. Sixteen honeybee colonies were selected for the experimentation. All colonies were standardized, each consisting of eight frames, and all maintained the same strength as worker bees. A frame feeder was provided for feeding treatments.

The leaves and fruits of selected plants were collected from Chhatrapati Sambhajinagar at Latitude: 19.9007° N Longitude: 75.3116° E. The fine leaf powders of plants, such as coriander (Coriandrum sativum), moringa (Moringa oleifera), tulsi (Ocimum sanctum), lemongrass (Cymbopogon citratus) and fruit powders of amla (Embilica officinalis) and guava (Psidium guava) were prepared. The selection of plants to prepare pollen substitute diets was based on phytochemicals. These fine powders of the above plant leaves and fruits were mixed with other common ingredients separately and prepared seven feed combinations, labeled as Diet -2 to Diet-8 (Table 1). White crystal sugar was used to prepare sugar syrup. Sugar and water were used in a 1:1 ratio (i.e., 50% sugar and 50% water), then boiled for 4 to 5 minutes and filtered.

Fine powders of coriander, moringa, tulsi and lemongrass leaves and fruit powders of amla and guava were added separately together with other ingredients such as defatted soya flour, skimmed milk powder, brewer's yeast, powdered sugar, turmeric powder, vitamins A, D, E & K, citric acid, guar gum powder, potassium sorbate (E202), sodium propionate (E281) as shown in Table 1.

Method of feeding

A total of sixteen colonies were selected for experimentation, each with eight frames, fully covered with bees on both sides. The total number of bees per frame was = 4000. Therefore, considering the previous assessment, the total number of bees was = 08 X 4000 = 32,000 bees/colony. These colonies were assigned to eight groups, each group with two colonies. Sixteen colonies were allotted as eight groups; each group contained two colonies. Colonies were then labelled as Diet-1 (Control), Diet-2 (Coriander), Diet-3 (Moringa), Diet-4 (Tulsi), Diet-5 (Lemongrass), Diet-6 (Amla), Diet-7 (Guava) and Diet-8 (Mixed diet). The feeders of all the colonies were washed with water and sun-dried properly. The 10 gm of diet powder + 100 ml sugar syrup (50%) this proportion of diet was provided separately for seven different groups of colonies. The first group of colonies was fed with Diet-1 (Control) 50% sugar syrup. a second group of colonies was fed Diet-2, a third group of colonies was fed Diet-3, a fourth group of colonies was fed Diet-4, a fifth group of colonies was fed Diet-5, a sixth group of colonies was fed Diet-6, a seventh group of colonies fed with Diet-7 and an eighth group of colonies fed with Diet-8. All diets were provided at 07.00 P.M. every day from July 01, 2023 to August 10, 2023.

Method of Nosema spore counting:

The honeybee samples for Nosema spore count were

collected from the control and each experimental colony. From each frame, 05 bees were collected randomly, regardless of age. A total of 40 worker bees were collected from each colony (05 bees X 08 frames = 40 bees per colony) and preserved temporarily in Isopropyl alcohol in a plastic screw-cap container. Each sample container was labeled with the date of collection and colony code.

From each colony sample, 40 honeybees with 10 mL of distilled water were crushed using a mortar and pestle, and then 30 mL of distilled water (1 mL of water per bee) was added (Abdel-Baki *et al.*, 2016).

The crushed bees' samples was then transferred to a 100 ml beaker. Then a few drops of crushed samples were loaded on an improved Neubauer hemocytometer using a micropipette (every time the tip was replaced) and observed under a compound microscope at 400X magnification.

Then, out of 25 blocks of the Neubauer hemocytometer, only five blocks without debris were selected for counting *Nosema* spores and the total number of *Nosema* spores in the chamber grid was counted (Mortensen *et al.*, 2020). *Nosema* spores were counted four times at intervals of 8 days. The first count was done before feeding the artificial diet as an initial reading. Subsequent counts were performed 8, 16, 24 and 32 days after feeding.

Calculation of the Nosema spore count

The measurement of observed *Nosema* spore was done using a Neubauer hemocytometer and estimation of counted *Nosema* spore was done using formulae and found several spores per bee (Observed spore count from 05 blocks multiplied by 04 million divided by 80 squares counted is equal to the number of spores per bee.

Nosema spores counted from 5 blocks, each with 16 squares (5 \times 16 = 80). If those 05 blocks were 39 \times 4,000,000/ 80= 19,50,000 (or 1.95 million) spores per bee using the formula.

Number of spores/bees =

Raw spore count from 5 blocks \times 4,000,000

No. of spores/bee = $\frac{00 \times 4,000,000}{80}$ = 50,000 spore/bee (i.e., 50.000 was considered as zero (Nil) spores per bee)

Statistical analysis

Analysis of data with one-way ANOVA using the Post Hoc-Duncan multiple range test (DMRT) by SPSS 29.0.2.0. The results were denoted as (Mean \pm Standard Error) and the separation of means described with

Table 1. Composition of plant-based pollen substitute diets (/100 gm)	ition of pl	ant-based	pollen st	ubstitute (diets (/100	(mg (
Diet Codes	DSF	SMP	ВΥ	PS	τь	Vit.	CA	GGP	E202	E281	CLP	MLP	TLP	LLP	AFP	GFP
Diet-1 Control	1 Liter o	1 Liter of 50 % Sugar syrup	lar syrup													
Diet-2	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	6.0g	ı	ı			ı
Diet-3	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	·	6.0g	ı	ı	ı	ı
Diet-4	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	,	ı	6.0g			ı
Diet-5	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	,	ı	ı	6.0g	ı	ı
Diet-6	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	·	ı	ı	ı	6.0g	ı
Diet-7	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g		ı	,	,	ı	6.0g
Diet-8	45g	15g	6.4g	20g	0.7g	0.8g	5.0g	0.5g	0.3g	0.3g	1.0g	1.0g	1.0g	1.0g	1.0g	1.0g
Abbreviations: DSF: Defatted Soya flour, SMP: Skimmed milk powder, BY: Brewer's yeast, PS: Powdered sugar, TP: Turmeric powder, Vit.: Vitamin A, D, E & K, CA: Citric acid, GGP: Guar gum powder, E202: Potassium sorbate, E281: Sodium propionate, CLP: Coriander Leaves Powder, MLP: Moringa Leaves Powder, TLP: Tulsi Leaves Powder, LLP: Lemongrass Leaves Powder, AFP: Amla fruit powder, GFP: Guava fruit powder	: Defatted assium sor der, GFP:	Soya flour, bate, E281 Guava fruit	SMP: Skit I: Sodium t powder	mmed milk propionat	<pre>< powder, E a, CLP: Cc</pre>	3Y: Brewei vriander Le	r's yeast, F ∋aves Pow	oS: Powder der, MLP: I	ed sugar, TP Moringa Lea [,]	: Turmeric po ves Powder,	owder, Vit.: ['] TLP: Tulsi	Vitamin A, Leaves Pc	D, E & K, ()wder, LLF	CA: Citric a	acid, GGP: ass Leave	Guar gum s Powder,

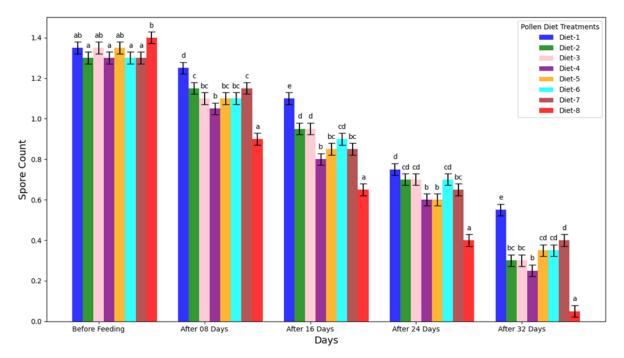
homogeneous subsets of means was noted by small alphabetical letters. The same letters meant 'No significant difference' among the means of subsets. P < 0.05 indicates the significant differences among the means in those subsets.

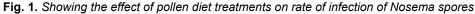
RESULTS AND DISCUSSION

The effect of plant-based pollen diet treatments on the rate of Nosema infection was observed. The infection rate was calculated before feed treatment and after feeding at 8-day intervals up to 32 days until the spores disappeared totally from the bees as shown in Table 2 and Fig. 1. The highest Nosema spore count was recorded control colonies fed with in Diet-1 (5,50,000±28867.5 spore/bee) after 32 days of feeding. The lowest rate of Nosema spore count was seen in the experimental group of colonies fed with Diet-8 (50,000±28867.5 spore/bee) as compared to control Diet-1 and other diets. However, Nosema spores were significantly reduced (all p values < 0.05). It was observed that feeding only sugar syrup was not adequate and healthy for the bees. A lowered rate of Nosema spore count was observed in colonies fed with Diet-8 (4,00,000±28867.5 spore/bee) after 24 days and Nosema spores were found to disappear totally after 32 days of feeding Diet-8 (50,000±28867.5 spore/bee), with a significant decrease in Nosema spores (all p values < 0.05). This value (50,000 Nosema spores/bee) is considered zero spores count in honeybees. The performance of Diet-4, containing tulsi leaf powder (2,50,000±28867.5 spore/bee) was found to be best in decreasing the infection rate significantly compared to

the other diets except Diet-8. The Diet-2 containing coriander leaf powder (3,00,000±28867.5 spore/bee) and Diet-3 with moringa leaf powder (3,00,000±28867.5 spore/bee) have shown similar Nosema spore counts with significantly lowered rate of Nosema infection after 32-days of feeding with a significant decrease in Nosema spores (all p values < 0.05). Similarly, Diet-5 containing lemongrass leaf powder (3,50,000±28867.5 spore/bee) and Diet-6 containing amla fruit powder (3,50,000±28867.5 spore/bee) have shown similar Nosema spore counts with a significant decrease of Nosema spores (all p values < 0.05). The Diet-7 containing guava fruit powder (4,00,000±28867.5 spore/ bee) has shown the lowest response in decreasing Nosema spore count as compared to Diet-2, Diet-3, Diet-4, Diet-5 and Diet-6 after 32 days of feeding with a significant decrease in Nosema spores count (all p values < 0.05).

The genus *Nosema* produces microspore-forming parasites that cause Nosemosis, a disease that affects worker bees, queens and drones by attacking the middle intestine's epithelial lining (Botías *et al.*, 2012; Bollan *et al.*, 2013). The unicellular microsporidium *Nosema apis* was the only recognized causative agent of Nosemosis in honeybees (*Apis mellifera*) (Nabian *et al.*, 2011). The *Nosema ceranae* can infect honeybee larvae and reduce longevity in adult bees (Eiri *et al.*, 2015; Palmer-Young *et al.*, 2017). According to Porrini *et al.*, 2011, Rinderer and Dell Elliott, 1977 certain proteins can raise Nosemosis. However, in the commercial diets they tested, there was no correlation between the protein content and the observed *Nosema* levels in the bees.





Gawali, A.R. and Waykar, B.B. / J. Appl. & Nat. Sci. 17(1), 193 - 199 (2025)

	0	•						. ,
Days	Diet-1 (Control)	Diet-2	Diet-3	Diet-4	Diet-5	Diet-6	Diet-7	Diet-8
No. of spores before feeding	1,350,000 ± 28867.5 ^{ªb}	1,300,000 ± 28867.5ª	1,350,000 ± 28867.5 ^{ab}	1,300,000 ± 28867.5ª	1,350,000 ± 28867.5 ^{ab}	1,300,000 ± 28867.5ª	1,300,000 ± 28867.5ª	1,400,000 ± 28867.5 ^b
After 08- days	1,250,000 ± 28867.5 ^d	1,150,000 ± 28867.5 ^c	1,100,000 ± 28867.5 ^{bc}	1,050,000 ± 28867.5 ^b	1,100,000 ± 28867.5 ^{bc}	1,100,000 ± 28867.5 ^{bc}	1,150,000 ± 28867.5 ^c	9,00,000 ± 28867.5 ^ª
After 16- days	1,100,000 ± 28867.5 [°]	9,50,000 ± 28867.5 ^d	9,50,000 ± 28867.5 ^d	8,00,000 ± 28867.5 ^b	8,50,000 ± 28867.5 ^{bc}	9,00,000 ± 28867.5 ^{cd}	8,50,000 ± 28867.5 ^{bc}	6,50,000 ± 28867.5ª
After 24- days	7,50,000 ± 28867.5 ^d	7,00,000 ± 28867.5 ^{cd}	7,00,000 ± 28867.5 ^{cd}	6,00,000 ± 28867.5 ^b	6,00,000 ± 28867.5 ^b	7,00,000 ± 28867.5 ^{cd}	6,50,000 ± 28867.5 ^{bc}	4,00,000 ± 28867.5ª
After 32- days	5,50,000 ± 28867.5 [°]	3,00,000 ± 28867.5 ^{bc}	3,00,000 ± 28867.5 ^{bc}	2,50,000 ± 28867.5 ^b	3,50,000 ± 28867.5 ^{cd}	3,50,000 ± 28867.5 ^{cd}	4,00,000 ± 28867.5 ^d	50,000 ± 28867.5 ^ª

Table 2. Showing the effect of pollen diet treatments on the Nosema spore count in honeybees. (Number of spores/ bee)

Data on the same alphabet means, 'No significant difference' among the means of subsets; P < 0.05 indicates the significant differences among the means in those subsets

In the present study, a plant-based pollen diet was rich in essential phytochemicals that significantly improved overall health, improved immunity and lowered microspore count. The results showed that plant-based pollen substitute diets have significantly decreased spores after feeding (all p values < 0.05). The medicinal plants have phytochemicals (i.e., phenol, flavonoid, alkaloid, terpenoid, p-coumaric acid and quercetin) that are responsible for the decreased rate of Nosema infection. The results of the present study correlated with (Maistrello et al., 2008), who also used medicinal plants with phytochemicals such as resveratrol (Polygonurn cuspidatum), lysozyme, vetiver oil (Vetiveria zizanioides) and thymol (Thyme). Phytochemicals showed positive effects, but they cannot show any toxic impact on adult bees and have a high consumption rate. All the compounds show significant effects in treating Nosema disease. Therefore, plant-based pollen diets were found to be a suitable treatment against Nosema disease.

Medicinal plants such as coriander, moringa, tulsi, lemongrass, amla and guava have almost all types of phytochemicals (phytosterol, phenolic acid, flavonoid, alkaloids, terpenoids, p-coumaric acids, caffeic acid, gallic acid and quercetin) that fulfill the requirement of nutrition and disease resistance capacity (Bhat *et al.*, 2014; Gopalakrishnan *et al.*, 2016; Singh and Chaudhuri, 2018; Vidhani *et al.*, 2016; Asaolu *et al.*, 2009; Mishra and Mahanta, 2014; Sachan *et al.*, 2013; Tanwar *et al.*, 2014). Phytochemicals such as caffeic acid, gallic acid, p-coumaric acids and kaempferol prolonged the honeybee's lifespan most effectively. It is necessary to boost growth, improve immunity and increase the resistance of bees against diseases (Bernklau *et al.*, 2019).

The decreased *Nosema* spore counts correlate significantly (all p values < 0.05) with the overall performance

and results of Bhat *et al.* (2014); Gopalakrishnan *et al.* (2016); Singh and Chaudhuri (2018); Vidhani *et al.* (2016); Asaolu *et al.* (2009); Mishra and Mahanta (2014); Sachan *et al.* (2013); Tanwar *et al.* (2014); Bernklau *et al.* (2019).

Analysis of variance (ANOVA) showed a significant difference (p < 0.05) in *Nosema* spore count among the different *Apis mellifera* L. colonies fed with plant-based pollen substitute diets and the control colonies. The various small alphabet letters within every column of Table 2 represent *Nosema* spore counts with statistically significant differences (P < 0.05).

Conclusion

This study has evaluated the impact of plant-based pollen substitute diets on the Nosema spore infection rate in Apis mellifera L. colonies. The number of spores per Diet treatment can vary based on bee nutrition. Plant-based pollen diets containing proteins and essential phytochemicals influenced microspores and reduced the Nosema infection. Diet-8, containing equal quantities of all the medicinal plants, lowered the number of Nosema spores observed in A. mellifera L. The mean differences are denoted as small alphabetical letters within every column representing statistically significant differences (P < 0.05). In addition, all ingredients such as defatted soya flour, skimmed milk powder, brewer's yeast, powdered sugar; turmeric powder; vitamins A, D, E and K; citric acid, guar gum powder; potassium sorbate and sodium propionate; the leaf and fruit powders of selected medicinal plants including leaf powders of coriander, moringa, tulsi, lemongrass and fruit powders of amla and guava with 50% sugar syrup have shown significant results in decreasing Nosema spore count and also to enhance immunity and overall efficiency of bees as compared to the control diet. Nutritionally, medicinal plants may fulfil the requirement of all phytochemicals essential for growth, reproduction and maintaining resistance against diseases. Hence, mixed Diet-8 can be recommended for the commercial beekeeping practice to improve honeybee colonies' health and overall performance.

ACKNOWLEDGEMENTS

The authors are thankful to the Department of Zoology, Dr. Babasaheb Ambedkar Marathwada University, Chhatrapati Sambhajinagar, for their assistance in research work.

Conflict of interest

The authors declare that they have no conflict of interest.

REFERENCES

- Abdel-Baki, A. A. S., Mares, M. M., Dkhil, M. A., & Al-Quraishy, S. (2016). First detection of *Nosema* sp., microsporidian parasites of honeybees (*Apis mellifera*) in Riyadh city, Saudi Arabia. *Journal of King Saud University-Science*, 28(4), 396-399. https://doi.org/10.1016/ j.jksus.2016.05.005
- Alberoni, D., Di Gioia, D., & Baffoni, L. (2023). Alterations in the microbiota of caged honeybees in the presence of *Nosema ceranae* infection and related changes in functionality. *Microbial Ecology*, 86(1), 601-616. https:// doi.org/10.1007/s00248-022-02050-4
- Araneda, X., Cumian, M., & Morales, D. (2015). Distribution, epidemiological characteristics and control methods of the pathogen *Nosema ceranae* Fries in honey bees *Apis mellifera* L.(Hymenoptera, Apidae). *Archivos de medicina veterinaria*, 47(2), 129-138. http:// dx.doi.org/10.4067/S0301-732X2015000200002
- Asaolu, M. F., Oyeyemi, O. A., & Olanlokun, J. O. (2009). Chemical compositions, phytochemical constituents and in vitro biological activity of various extracts of Cymbopogon citratus. *Pakistan Journal of Nutrition*, 8(12), 1920-1922. https://doi.org/10.3923/pjn.2009.1920.1922
- Bernklau, E., Bjostad, L., Hogeboom, A., Carlisle, A., & HS, A. (2019). Dietary phytochemicals, honey bee longevity and pathogen tolerance. *Insects*, 10(1), 14. https:// doi.org/10.3390/insects10010014
- Bhat, S., Kaushal, P., Kaur, M., & Sharma, H. K. (2014). Coriander (Coriandrum sativum L.): Processing, nutritional and functional aspects. *African Journal of plant science*, 8 (1), 25-33. http://dx.doi.org/10.5897/AJPS2013.1118
- Bollan, K. A., Hothersall, J. D., Moffat, C., Durkacz, J., Saranzewa, N., Wright, G. A., & Connolly, C. N. (2013). The microsporidian parasites *Nosema* ceranae and *Nosema* apis are widespread in honeybee (*Apis mellifera*) colonies across Scotland. *Parasitology Research*, 112, 751-759. https://doi.org/10.1007/s00436-012-3195-0
- Botías, C., Martín-Hernández, R., Meana, A., & Higes, M. (2012). Critical aspects of the *Nosema spp*. diagnostic sampling in honey bee (*Apis mellifera* L.) colonies. *ParasitologyRresearch*, 110, 2557-2561. https://

doi.org/10.1007/s00436-011-2760-2

- Bourgeois, A. L., Rinderer, T. E., Beaman, L. D., & Danka, R. G. (2010). Genetic detection and quantification of *Nosema apis* and *N. ceranae* in the honey bee. *Journal of Invertebrate Pathology*, 103(1), 53-58. https:// doi.org/10.1016/j.jip.2009.10.009
- Daisley, B. A., Pitek, A. P., Chmiel, J. A., Al, K. F., Chernyshova, A. M., Faragalla, K. M., & Reid, G. (2020). Novel probiotic approach to counter Paenibacillus larvae infection in honey bees. *The ISME journal*, 14(2), 476-491. https://doi.org/10.1038/s41396-019-0541-6
- DeGrandi-Hoffman, G., Chen, Y., Huang, E., & Huang, M. H. (2010). The effect of diet on protein concentration, hypopharyngeal gland development and virus load in worker honey bees (*Apis mellifera L.*). *Journal of insect physiology*, 56(9), 1184-1191. https://doi.org/10.1016/ j.jinsphys.2010.03.017
- Eiri, D. M., Suwannapong, G., Endler, M., & Nieh, J. C. (2015). *Nosema ceranae* can infect honey bee larvae and reduces subsequent adult longevity. *PLoS One*, 10(5), e0126330. https://doi.org/10.1371/journal.pone.0126330
- Fremont, L. (2000). Biological effects of resveratrol. *Life* sciences, 66(8), 663-673. https://doi.org/10.1016/s0024-3205(99)00410-5
- Gopalakrishnan, L., Doriya, K., & Kumar, D. S. (2016). Moringa oleifera: A review on nutritive importance and its medicinal application. *Food science and human wellness*, 5(2), 49-56. https://doi.org/10.1016/ j.fshw.2016.04.001
- Higes, M., Martín-Hernández, R., & Meana, A. (2010). Nosema ceranae in Europa: eine neu auftretende Nosemose Typ C. Apidologie, 41, 375-392. https:// doi.org/10.1051/apido/2010019
- 16. Jousse, C., Dalle, C., Abila, A., Traïkia, M., Diogon, M., Lyan, B., & Delbac, F. (2020). A combined LC-MS and NMR approach to reveal metabolic changes in the hemolymph of honeybees infected by the gut parasite *Nosema ceranae*. *Journal of invertebrate pathology*, 176, 107478. https://doi.org/10.1016/j.jip.2020.107478
- Kulhanek, K., Steinhauer, N., Wilkes, J., Wilson, M., Spivak, M., Sagili, R. R., & VanEngelsdorp, D. (2021). Survey-derived best management practices for backyard beekeepers improve colony health and reduce mortality. *PLoS One*, 16(1), e0245490. https://doi.org/10.1371/ journal.pone.0245490
- Maistrello, L., Lodesani, M., Costa, C., Leonardi, F., Marani, G., Caldon, M., & Granato, A. (2008). Screening of natural compounds for the control of *Nosema* disease in honeybees (*Apis mellifera*). *Apidologie*, 39(4), 436-445. https://hal.science/hal-00891923v1
- Mayack, C., & Naug, D. (2009). Energetic stress in the honeybee Apis mellifera from Nosema ceranae infection. Journal of invertebrate pathology, 100(3), 185-188. https:// doi.org/10.1016/j.jip.2008.12.001
- Mishra, P., & Mahanta, C. L. (2014). Comparative analysis of functional and nutritive values of amla (*Emblica officinalis*) fruit, seed and seed coat powder. *American Journal* of Food Technology, 9(3), 151-161. http:// dx.doi.org/10.3923/ajft.2014.151.161
- Mortensen, A. N., Jack, C. J., McConnell, M., Teigen, L., & Ellis, J. (2020). How to Quantify *Nosema* spores infection rate in a Honey Bee Colony. *Bulletin ENY-167*, 1-5.

https://doi.org/10.32473/edis-in1123-2016

- Mulholland, G. E., Traver, B. E., Johnson, N. G., & Fell, R. D. (2012). Individual variability of *Nosema ceranae* infections in *Apis mellifera* colonies. *Insects*, 3(4), 1143-1155. https://doi.org/10.3390/insects3041143
- Nabian, S., Ahmadi, K., Shirazi, M. N., & Sadeghian, A. G. (2011). First detection of *Nosema ceranae*, a microsporidian protozoa of European honeybees (*Apis mellifera*) in Iran. *Iranian journal of parasitology*, 6(3), 89-95. https:// pmc.ncbi.nlm.nih.gov/articles/PMC3279895/
- 24. Palmer-Young, E. C., Tozkar, C. Ö., Schwarz, R. S., Chen, Y., Irwin, R. E., Adler, L. S., & Evans, J. D. (2017). Nectar and pollen phytochemicals stimulate honey bee (Hymenoptera: Apidae) immunity to viral infection. *Journal* of economic entomology, 110(5), 1959-1972. https:// doi.org/10.1093/jee/tox193
- Porrini, M. P., Sarlo, E. G., Medici, S. K., Garrido, P. M., Porrini, D. P., Damiani, N., & Eguaras, M. J. (2011). Nosema ceranae development in Apis mellifera: influence of diet and infective inoculum. Journal of Apicultural Research, 50(1), 35-41. http://dx.doi.org/10.3896/ IBRA.1.50.1.04
- Prokhoda, I. A., Eliseeva, E. V., Polesskaya, O. P., Potseluev, E. I., & Zabenko, D. V. (2019). Management of the life cycle of the innovation apiproduct from drone larvae and its introduction in the food industry. In *IOP Conference Series: Earth and Environmental Science* (Vol. 274, No. 1, p. 012122). IOP Publishing. http://dx.doi.org/10.1088/1755-1315/274/1/012122
- Rice, R. N. (2001). *Nosema* disease in honeybees. Genetic variation and control. Report n. 01/46, Australian Government, Rural Industries Research and Development

Corporation.

- Rinderer, T. E., & Dell Elliott, K. (1977). Worker honey bee response to infection with *Nosema apis*: influence of diet. *Journal of Economic Entomology*, 70(4), 431-433. https://doi.org/10.1093/jee/70.4.431
- Sachan, N. K., Gangwar, S. S., Sharma, R., & Kumar, Y. (2013). An investigation into phytochemical profile and neutraceutical value of amla (*Emblica officinalis*) fruits. *Int J Modern Pharm Res*, 2(1), 1-12.
- Singh, D., & Chaudhuri, P. K. (2018). A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). *Industrial Crops and Products*, 118, 367-382. https://doi.org/10.1016/j.indcrop.2018.03.048
- Tanwar, B., Andallu, B., & Chandel, S. (2014). Influence of processing on physicochemical and nutritional composition of *Psidium Guajava* L.(Guava) products. *International Journal of Agriculture and Food Science Technology*, 5(2), 47-54.
- 32. Vidhani, S. I., Vyas, V. G., Parmar, H. J., Bhalani, V. M., Hassan, M. M., Gaber, A., & Golakiya, B. A., (2016). Evaluation of some chemical composition, minerals fatty acid profiles, antioxidant and antimicrobial activities of Tulsi (*Ocimum sanctum*) from India. *American Journal of Food Science and Technology*, 4(2), 52-57. http:// dx.doi.org/10.12691/ajfst-4-2-5
- 33. Yücel, B., & Doğaroğlu, M. (2005). The impact of Nosema apis Z. infestation of honey bee (Apis mellifera L.) colonies after using different treatment methods and their effects on the population levels of workers and honey production on consecutive years, 1142-1145. http:// dx.doi.org/10.3923/pjbs.2005.1142.1145