

Research Article

## Standardization of packaging material, storage durations and chemical treatments on vase life of soft shield fern *Polystichum squarrosum* (D. Don)

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### Abstract

*Polystichum* species, native to warm-temperate and montane-tropical environments (including some alpine regions), are terrestrial or rock-dwelling ferns. These plants exhibit dark green, leather-like, and lustrous fronds, which are highly valued in floral arrangements due to their aesthetic elegance and longevity. *Polystichum squarrosum* is not being cultivated commercially but collected from forests for its use as cut green in the florist industry. Hence the collected material should be handled carefully to utilize every frond. Therefore, the present study aimed to develop suitable post-harvest handling procedures for this valued cut green. A Completely Randomized Design (CRD) Factorial layout with 19 treatment modules ( $M_0$  to  $M_{18}$ ) was employed. Treatment modules included packaging material as poly mesh net bags and packing materials as corrugated fiberboard boxes, two storage durations (3 and 6 days) and chemical treatments [glycerol (4%) and Benzyl adenine ( $25\text{mg L}^{-1}$ )]. Results revealed that spraying cut fronds with 4% glycerol, wrapping them in cellophane sheets and storing them for three days ( $M_6$ ) resulted in significant improvements in appearance, weight change, vase life and water uptake. Fronds from Season-II (March-April 2020) exhibited excellent performance over Season-I (August-September 2019). The interaction between  $M_6$  and Season-II further enhanced the postharvest parameters. The combination of glycerol treatment and cellophane wrapping for three-day storage proved most effective in preserving the quality of cut fronds.

**Keywords:** Benzyl adenine, Glycerol, Packaging, *Polystichum squarrosum*, Vase life, storage

### INTRODUCTION

*Polystichum* is a diverse genus of 500 species in Dryopteridaceae family, commonly found in the temperate and subtropical regions (He and Zhang, 2022). In North-Western Himalayan regions it occurs at altitudes of 1900-2400 above msl (Saggoo and Kaur, 2018). There are 50 species under the genus *Polystichum* reported from India (Jangi *et al.*, 2021). *Polystichum squarrosum*

is being commercially utilized in floriculture industry as a cut green. It is well-known for its medicinal properties, including antibacterial effects, and for treating pyloric diseases (Singh, 1999). *P. squarrosum* ( $2n = 82$ ) is reported from various North-Western Himalayan regions viz. Nainital, Mussoorie, Shimla, Matiana, Dalhousie, Manali, Solan, Barog and Chamoli (Saggoo and Kaur, 2018). The fronds have dark green, leathery and shiny foliage valued in floriculture for their ele-

gance and long lasting nature (Yadav *et al.*, 2018). The leaves have commercial importance and are extensively used to decorate marriages, pandals and temples. They are commonly sold by the name 'Pahadi patti' in florist shops (Yadav *et al.*, 2018). Cut greens impart freshness and background contrasting colour and variety to the floral designs; hence, they constitute an important component of floricultural industry largely used as fillers in bouquets and flower arrangements. They also play a major role in enhancing the national GDP (Pacifci *et al.*, 2007, Reid and Jiang, 2012, Abou-Dahab *et al.*, 2013).

Antitranspirants reduce water loss from plant leaves by reducing stomata size and number (Goreta *et al.*, 2007). They fall into three categories: film-forming types like glycerol, reflecting materials that bounce off radiation from leaf surfaces and stomatal closing agents such as  $(\text{MgCO}_3)$  which impact leaf tissue metabolism (Kumar *et al.*, 2023; Warmund *et al.*, 2021; Lv *et al.*, 2020). Antitranspirants have different modes of action (Kociecka *et al.*, 2023; Chaves *et al.*, 2009) and their formulations prevent excess water loss without reducing  $\text{CO}_2$  uptake. They can be applied in the foliar form by reducing transpiration in three different ways (Mphande *et al.*, 2020). The first set of techniques involves the application of reflective materials through spraying to reduce the absorption of radiant energy (Mphande *et al.*, 2020). The second approach entails the utilization of antitranspirants, which are sprayed onto foliage and form transparent films which effectively obstruct the outward movement of water vapor from the leaf surface (Moroni *et al.*, 2020). An additional mechanism employed by antitranspirants involves the inhibition of stomata opening, consequently reducing the water loss from the leaf (Sow and Ranjan, 2021).

Growth regulators are recommended to prolong post-harvest longevity, benzyl adenine (BA) is one of the growth regulators that delays leaf yellowing and consequently increases leaf longevity (Zhang *et al.*, 2023). Spraying of 6- benzylaminopurine (BAP) at concentrations of  $37.5\text{--}300\text{ mg L}^{-1}$  improved postharvest durability of *Anthurium andreaeanum* 'Apalai' flower without inducing spathe blueing (Favero *et al.*, 2020). The accelerated onset of senescence poses a challenge in exporting cut foliage plant species. External factors like temperature, moisture, gases, radiation and pathogens induce senescence in cut flowers (Raza *et al.*, 2022).

Packaging materials and chemical treatments helps extend the life of cut flowers vase (Sharma *et al.*, 2010). Packaging plays a pivotal role in the flower trade, significantly impact the quality of cut flowers and foliage to provide quality products for consumers (Shinde *et al.*, 2023). Effective packaging methods are vital for prolonging storage duration and preserving flower quality during transportation (Jang *et al.*, 2021).

The key principles underlying successful packaging involve minimizing the rates of transpiration, respiration, and cell division (Khalid *et al.*, 2020). Corrugated fiber board boxes stand out among the most suitable packaging materials (Lai *et al.*, 2022). Additionally, a wide array of interior packaging materials such as paper, paperboard, transparent films, polyethylene, and other innovative materials are available, necessitating thorough evaluation for their suitability and cost-effectiveness. Packaging should minimize transpiration, respiration, and cell division to maintain long-term cut foliage storage and quality during transportation (Bhattacharjee, 1999). Therefore, a comprehensive comparison of diverse packaging options is essential to ensure the extended freshness of flowers or cut greens for secure transport to their destinations (Sarkar *et al.*, 2019). *Polystichum* is not being cultivated commercially but it is being collected from the forests for its utilization as cut green in florist industry. As a result of indiscriminate collection from forests, it has become an endangered plant species. Hence, the collected material should be handled carefully to utilize every frond. Therefore, suitable postharvest handling procedures for this valued cut green are needed. Hence, the present study aimed to assess the storage materials, storage durations, and chemical treatments that enhance the longevity of *P. squarrosum* cut fronds.

## MATERIALS AND METHODS

The present study on vase life of soft shield fern (*P. squarrosum*) was performed at the farm laboratory of the Department of Floriculture and Landscape Architecture, Dr Yashwant Singh Parmar University of Horticulture and Forestry in Nauni, Solan H.P. India, 173230. The study was conducted in two seasons, August - September 2019 (Season-I) and March - April 2020 (Season-II). Fronds from Chail forest, Solan district of Himachal Pradesh, were promptly collected and brought to the laboratory after harvesting. Treatment modules included three packaging material cellophane sheets, poly mesh net bags and corrugated fiberboard boxes (CFB), two storage durations (3 and 6 days) and spraying of glycerol (4 %) and Benzyl Adenine ( $25\text{ mg L}^{-1}$ ). The cut fronds of Soft shield fern measuring 80 cm were prepared in the morning (7-8 am) and their fresh weight was recorded simultaneously. Thereafter each cut frond was sprayed with glycerol (4 %) and Benzyl Adenine ( $25\text{ mg L}^{-1}$ ) treatment-wise. Then, the treated fronds were stored in different packaging materials (i.e. cellophane sheets, polymesh net bags and corrugated fiberboard boxes) for three and six days at ambient temperature. During the study period, the average temperature, relative humidity, and light intensity were recorded as 24.05 degrees Celsius, 61.38%, and 827.50

Lux in season-I. However, for season-II, the values were slightly different, with a temperature of 19.10 degrees Celsius, relative humidity of 62.39%, and light intensity of 734.82 Lux. The observation of weight loss and appearance of fronds were recorded after storage. Subsequently, the stored fronds and fresh fronds were placed in test tubes containing 70 ml of distilled water to assess their longevity. Freshness and color were graded (A, B, C) with corresponding points of 5, 3 and 1, respectively, based on the Royal Horticulture Society colour chart (RHS) (Voss, 2002).

There were 19 treatment modules and details are  $M_0$  = Control (Fresh fronds collected from forest and placed in test tube containing water),  $M_1$  = Fronds packed in Corrugated fiberboard boxes and stored for three days,  $M_2$  = Fronds packed in Corrugated fiberboard boxes and stored for six days,  $M_3$  = Fronds wrapped in cellophane sheets with three days storage,  $M_4$  = Fronds wrapped in cellophane sheets with six days storage duration,  $M_5$  = Poly mesh net bags as packaging material with three days storage,  $M_6$  = Poly mesh net bags as packaging material with six days storage,  $M_7$  = cut fronds sprayed with glycerol (4%) and packed in Corrugated fiberboard boxes and stored for three days,  $M_8$  = fronds sprayed with glycerol (4%) and packed in Corrugated fiberboard boxes and stored for six days,  $M_9$  = fronds sprayed with glycerol and wrapped in cellophane sheets with three days storage,  $M_{10}$  = spraying the fronds with glycerol @ 4% and wrapped in cellophane sheets with six days storage,  $M_{11}$  = fronds sprayed with glycerol @ 4% and kept in Poly mesh net bags with three days storage,  $M_{12}$  = spraying the fronds with glycerol 4% and kept in Poly mesh net bags with six days storage,  $M_{13}$  = Fronds sprayed with Benzyl Adenine ( $25\text{mg L}^{-1}$ ) and packed in Corrugated fiberboard boxes with three days storage,  $M_{14}$  = Fronds sprayed with Benzyl Adenine ( $25\text{mg L}^{-1}$ ) and packed in Corrugated fiberboard boxes with three six storage,  $M_{15}$  = Fronds sprayed with Benzyl Adenine ( $25\text{mg L}^{-1}$ ) and kept in Poly mesh net bags with three days storage,  $M_{16}$  = Fronds sprayed with enyl Adenine ( $25\text{mg L}^{-1}$ ) and kept in Poly mesh net bags with six days storage,  $M_{17}$  = Fronds sprayed with Benzyl Adenine @  $25\text{mg L}^{-1}$  and wrapped in cellophane sheets with storage duration three days and  $M_{18}$  = BA - Fronds sprayed with Benzyl Adenine @  $25\text{mg L}^{-1}$  and wrapped in cellophane sheets with storage duration six days.

#### Statistical analysis

By employing a Completely Randomized Design (CRD), the data for quantitative estimation of various physico-chemical attributes were analyzed. All the data recorded was subjected to statistical analysis using MS-Excel and OP-Stat to determine the significance of the results obtained.

## RESULTS AND DISCUSSION

### Appearance (Freshness and colour)

The data in Table 1 shows that treatment module  $M_9$  i.e. spraying of cut fronds with Glycerol @ 4 % then wrapping in cellophane sheets and 3 days storage, resulted in the best appearance score (4.67) for cut fronds, while module  $M_2$  (i.e. fronds packed in CFB Boxes with 6 days storage) had the lowest score (1.73). Cut fronds from Season-II ( $S_2$ ) exhibited a better appearance (3.36) than those from Season-I ( $S_1$ , 3.23). The interaction of  $M_9 \times S_2$  gave the best appearance (4.80), while  $M_2 \times S_1$  had the lowest (1.47). The treatment module  $M_9$ , involving spraying fronds with 4% glycerol and wrapping them in cellophane sheets for 3 days of storage, improved turgidity and minimized water evaporation, creating a favorable micro-climate for physiological processes. The results were supported by the findings of Punetha and Trivedi (2018) in the rose cultivar 'Naranjo'. Fronds from Season-II outperformed Season-I due to physiological maturity and accumulated food reserves, while Season-I fronds suffered from succulence caused by prolonged rainy conditions. The results were supported from the findings of (Wahba *et al.*, 2017) in *Cynara cardunculus* L and (Gololo *et al.*, 2016) in *Barleria dinteri*, *Grewia flava* and *Jatropha lagarinthoides* that seasonal changes affected the quantity of inherent phytochemicals in the plant leaves.

### Chlorophyll content (mg/g)

From the Table 1, treatment modules significantly affected the chlorophyll content of cut fronds. Maximum chlorophyll content (0.65 mg/g) occurred in  $M_{17}$  in which Benzyl adenine (BA) @  $25\text{mg L}^{-1}$  was sprayed on cut fronds and then fronds wrapped in cellophane sheets with 3-day storage which was at par with the treatment module  $M_{18}$ . Minimum chlorophyll content (0.30 mg/g) was in  $M_2$  (i.e. fronds stored in CFB Boxes with 6-day storage duration). Season-II exhibited higher chlorophyll content (0.45 mg/g) than Season-I (0.42 mg/g).  $M_{17} \times S_2$  interaction showed maximum chlorophyll content (0.66 mg/g) while  $M_2 \times S_1$  had minimum (0.29 mg/g).  $M_{17}$  (BA spraying with cellophane wrapping) in 3-day storage was found most effective in maintaining high chlorophyll content in both seasons. The maximum chlorophyll content in cut fronds treated with BA ( $25\text{mg L}^{-1}$ ) and wrapped in cellophane sheets for three days is attributed to the slowdown of senescence and chlorophyll degradation by application of Benzyl adenine due to the factor that Benzyl adenine application reduces the yellowing of leaves (Favero *et al.*, 2020). Packaging provides a barrier against air flow and minimizes transpiration losses, creating a favorable micro-climate for frond freshness (Lufu *et al.*, 2020). Glycerol (4%) application also delays chlorophyll degra-

**Table. 1.** Effect of post-harvest treatment modules on seasonal appearance (freshness & colour), chlorophyll content and fresh weight of cut fronds of *Polystichum squarrosum*

Treatment modules	Appearance (Freshness & colour)			Chlorophyll content (mg/g)			Fresh weight (g)		
	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean
M <sub>0</sub>	3.53	3.60	3.57	0.38	0.46	0.42	29.85	29.99	29.92
M <sub>1</sub>	2.47	2.60	2.53	0.31	0.33	0.32	29.80	30.04	29.92
M <sub>2</sub>	1.47	2.00	1.73	0.29	0.32	0.30	29.89	29.95	29.92
M <sub>3</sub>	4.07	4.40	4.23	0.57	0.58	0.57	29.87	29.95	29.91
M <sub>4</sub>	3.53	3.60	3.57	0.40	0.45	0.42	29.90	29.90	29.90
M <sub>5</sub>	3.67	3.80	3.73	0.43	0.46	0.45	29.89	30.00	29.95
M <sub>6</sub>	2.17	2.20	2.18	0.32	0.35	0.34	29.50	29.98	29.74
M <sub>7</sub>	3.60	3.60	3.60	0.44	0.46	0.45	29.94	29.97	29.96
M <sub>8</sub>	2.40	2.40	2.40	0.35	0.37	0.36	29.87	29.99	29.93
M <sub>9</sub>	4.53	4.80	4.67	0.61	0.63	0.62	29.64	29.96	29.80
M <sub>10</sub>	3.60	3.73	3.67	0.45	0.48	0.47	29.39	29.82	29.61
M <sub>11</sub>	4.07	4.20	4.13	0.42	0.44	0.43	29.37	29.75	29.56
M <sub>12</sub>	3.07	3.13	3.10	0.35	0.36	0.35	29.96	29.94	29.95
M <sub>13</sub>	3.20	3.20	3.20	0.38	0.38	0.38	29.83	29.82	29.82
M <sub>14</sub>	2.20	2.53	2.37	0.32	0.34	0.33	29.94	30.01	29.98
M <sub>15</sub>	3.33	3.40	3.37	0.40	0.42	0.41	29.78	29.95	29.87
M <sub>16</sub>	2.17	2.20	2.18	0.32	0.34	0.33	29.74	29.86	29.80
M <sub>17</sub>	4.33	4.40	4.37	0.65	0.66	0.65	29.86	29.27	29.57
M <sub>18</sub>	3.87	4.07	3.97	0.63	0.64	0.64	29.98	29.87	29.92
Mean	3.23	3.36		0.42	0.45		29.79	29.90	
CD <sub>0.05</sub>	Modules		0.15	Modules		0.02	Modules		NS
	Seasons		0.05	Seasons		0.01	Seasons		NS
	Modules × Seasons		0.21	Modules × Seasons		0.03	Modules × Seasons		NS

Season-I- August-September (2019); Season-II- March – April (2020)

dition and enhance freshness (Nguyen *et al.*, 2021).

#### Weight of fronds after storage/weight change (%)

The data in Table 2 shows that treatment modules significantly affected the weight change (%) of cut fronds. M<sub>9</sub> (i.e. glycerol spraying, cellophane wrapping and 3-day storage) showed the minimum weight change (3.33%), superior to all other treatments. M<sub>2</sub> (i.e. fronds stored in Corrugated Fiber Board boxes and stored for 6-days) exhibited the maximum weight change (35.66%). Minimum weight change of cut fronds (15.35%) was found in Season-II over Season-I (20.59%). In interaction M<sub>9</sub> × S<sub>2</sub> (glycerol spraying, cellophane wrapping and 3-day storage in Season-II) had the minimum weight change (2.80%) whereas, in M<sub>2</sub> × S<sub>1</sub> (fronds stored in Corrugated Fiber Board boxes, 6-day storage in Season-II) had the maximum weight change (40.04%). The minimum weight change in cut fronds treated with glycerol (4%) and wrapped in cellophane sheets for three days is due to low transpiration loss and maintained relative humidity during storage. During the season-II, the low temperatures significantly contributed to minimal physiological weight loss. This is because water loss is reduced in colder conditions, positively impacting the physiological weight of the cut fronds. (Pranuthi *et al.*, 2018) Similar results of low physiological weight loss were found in rose cv. 'Kiro' under low temperature conditions. Moreover, the maximum weight loss observed during prolonged storage can be attributed to the process of respiration or the

breakdown of complex compounds into simple compounds with low molecular weight (Anggraeni, 2021). Senescence processes in cut stems lead to consuming stored food in jujube fruit (Yang *et al.*, 2021).

#### Vase life (days)

The data in Table 2 indicate that treatment modules and seasons significantly influenced the vase life of cut fronds. The maximum vase life (19.14 days) was observed in M<sub>9</sub> (glycerol spraying, cellophane wrapping and 3-day storage), while the minimum (7.87 days) was in M<sub>2</sub> (CFB boxes, 6-day storage) which was at par with M<sub>6</sub> (8.47 days). Longer vase life (13.33 days) was observed in Season-II as compared to that in Season-I (10.19 days). The M<sub>9</sub> × S<sub>2</sub> interaction showed significantly maximum vase life (21.14 days) in cut fronds with glycerol spraying and cellophane wrapping for three days in Season-II, while M<sub>2</sub> × S<sub>1</sub> had the minimum (6.88 days) in Season-I.

The glycerol treatment might have affected stomata conductivity, reducing water loss and maintaining turgidity, thus prolonging vase life. Cellophane wrapping create a favorable atmosphere, slowing respiration, transpiration, and cell division processes (Sarkar *et al.*, 2019). However, due to physiological changes during storage, rapid senescence occurred after a specific period in *Nephrolepis exaltata* (Abou-Dahab *et al.*, 2013).

#### Water uptake (ml)

**Table. 2** Effect of treatment modules on weight change (%), vase life (days) and water uptake/consumed (ml) by *Polystichum squarrosum*

Treatment modules	Weight Change (%)			Vase life (days)			Water uptake (ml)		
	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean	Season-I (S <sub>1</sub> )	Season-II (S <sub>2</sub> )	Mean
M <sub>0</sub>	15.82	12.34	14.08	8.25	12.32	10.29	33.00	25.83	29.42
M <sub>1</sub>	19.62	11.01	15.32	8.32	10.42	9.37	35.33	26.86	31.10
M <sub>2</sub>	40.04	31.28	35.66	6.88	8.85	7.87	24.53	15.90	20.22
M <sub>3</sub>	7.31	3.19	5.25	13.57	17.57	15.57	38.33	29.16	33.75
M <sub>4</sub>	17.06	10.97	14.02	10.53	13.43	11.98	35.93	26.27	31.10
M <sub>5</sub>	15.93	11.98	13.95	10.07	11.78	10.93	39.00	28.87	33.94
M <sub>6</sub>	30.43	27.22	28.83	6.97	9.97	8.47	21.60	11.26	16.43
M <sub>7</sub>	18.01	11.98	14.99	10.35	14.35	12.35	27.00	18.15	22.58
M <sub>8</sub>	29.17	20.67	24.92	8.30	11.29	9.80	24.33	17.09	20.71
M <sub>9</sub>	3.86	2.80	3.33	17.14	21.14	19.14	17.80	10.10	13.95
M <sub>10</sub>	9.72	8.84	9.28	13.23	17.23	15.23	20.47	14.20	17.33
M <sub>11</sub>	19.57	15.61	17.59	11.34	15.34	13.34	27.27	18.16	22.71
M <sub>12</sub>	27.29	24.36	25.83	9.53	10.52	10.02	19.07	11.17	15.12
M <sub>13</sub>	26.46	19.26	22.86	9.61	12.28	10.95	38.60	28.38	33.49
M <sub>14</sub>	33.69	21.45	27.57	7.59	10.59	9.09	33.87	23.27	28.57
M <sub>15</sub>	23.99	17.61	20.80	9.55	13.22	11.38	39.93	29.51	34.72
M <sub>16</sub>	31.40	26.69	29.04	7.45	10.05	8.75	30.40	23.41	26.91
M <sub>17</sub>	8.45	5.12	6.79	14.17	18.17	16.17	43.73	34.01	38.87
M <sub>18</sub>	13.33	9.26	11.30	10.83	14.83	12.83	39.73	30.14	34.94
Mean	20.59	15.35		10.19	13.33		31.05	22.20	
CD <sub>0.05</sub>	Modules		1.41	Modules		0.84	Modules		0.05
	Seasons		0.46	Seasons		0.27	Seasons		1.26
	Modules × Seasons		1.99	Modules × Seasons		1.19	Modules × Seasons		1.78

Season-I- Experiment conducted during August-September (2019); Season-II- Experiment conducted during March – April (2020)

The data in Table 2 showed a significant influence of treatment modules and seasons on water uptake by cut fronds. Treatment module M<sub>9</sub> (glycerol spraying, cellophane wrapping, 3-day storage) had the minimum water uptake (13.95 ml/stem), while M<sub>17</sub> (BA spraying, cellophane wrapping, 3-day storage) had the maximum (38.87 ml/stem) water uptake. Season-II (S<sub>2</sub>) exhibited lower water uptake (22.20 ml/stem) compared to Season-I (S<sub>1</sub>, 31.05 ml/stem). In the M<sub>9</sub> × S<sub>2</sub> interaction, the minimum water uptake (10.10 ml/stem) occurred (glycerol spraying, cellophane wrapping for three days in Season-II), and the maximum occurred (43.73 ml/stem) in M<sub>17</sub> × S<sub>1</sub> interaction (BA spraying, cellophane wrapping for three days in Season-I). Glycerol treatment maintained water balance as an osmolyte, promoting normal water absorption (Rida and El-Gedaway, 2022; Revathi *et al.*, 2021) while cellophane wrapping created optimal conditions for healthy physiological systems (Punetha and Trivedi, 2018). Higher water uptake in Season-I might be due to increased respiration at higher temperatures (Dusenge *et al.*, 2019). Lower water uptake in fronds stored for 6 days could be attributed to decreased water absorption ability, aligning with the findings of Patel *et al.* (2008) in spider lily.

## Conclusion

The treatment module M<sub>9</sub> (glycerol spraying, cello-

phane wrapping, 3-day storage) resulted in the best appearance, maximum chlorophyll content, minimum weight change and longest vase life of cut *Polystichum squarrosum* fronds. Season-II (March-April 2020) fronds performed better than Season-I (August-September 2019). The combination of glycerol treatment and cellophane wrapping for three-day storage proved most effective in preserving the quality of cut fronds.

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**Conflict of Interest**

The authors declare that they have no conflict of interest.

**REFERENCES**

- Abou-Dahab, T. A., El-Kady, A. F., Khenizy, S. A. & El-Ebrashi, E. F. (2013). Impact of various pulsing and holding solutions on the quality and longevity of *Nephrolepis exaltata* (L.) Schott cut foliage under room temperature. *Journal of Horticultural Science and Ornamental Plants*, 5(2), 89-9. <https://doi.org/10.5829/idosi.jhsop.2013.5.2.1117>
- Angraeni, R. S. (2021). Antibacterial (*Staphylococcus aureus* and *Escherichia coli*) and Antifungal (*Saccharomyces cerevisiae*) Activity Assay on Nanoemulsion Formulation of Ethanol Extract of Mangosteen Leaves (*Garcinia mangostana* L.) as Fruit Preservative. *Journal of Food and Pharmaceutical Sciences*, 9(1), 1-1. <https://doi.org/10.22146/jfps.1008>
- Bhattacharjee, S. K. (1999). Postharvest technology of flowers and ornamental plants. *New India Publishing*.
- Chaves, M. M., Flexas, J. & Pinheiro, C. (2009). Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of Botany*, 103(4), 551-560. <https://doi.org/10.1093/aob/mcn125>
- Dusenge, M. E., Duarte, A. G. & Way, D. A. (2019). Plant carbon metabolism and climate change: elevated CO<sub>2</sub> and temperature impacts on photosynthesis, photorespiration and respiration. *New Phytologist*, 221(1), 32-49. <https://doi.org/10.1111/nph.15283>
- Favero, B. T., Lutken, H., Dole, J. M. & Lima, G. P. P. (2020). *Anthurium andraeanum* senescence in response to 6-benzylaminopurine: Vase life and biochemical aspects. *Postharvest Biology and Technology*, 161, 111084. <https://doi.org/10.1016/j.postharvbio.2019.111084>
- Gololo, S. S., Shai, L. J., Agyei, N. M. & Mogale, M. A. (2016). Effect of seasonal changes on the quantity of phytochemicals in the leaves of three medicinal plants from Limpopo province, South Africa. *Journal of Pharmacognosy and Phytotherapy*, 8(9), 168-172. <https://doi.org/10.5897/JPP2016.0408>
- Goreta, S., Leskovar, D. I. & Jifon, J. L. (2007). Gas exchange, water status, and growth of pepper seedlings exposed to transient water deficit stress are differentially altered by antitranspirants. *Journal of the American Society for Horticultural Science*, 132(5), 603-610. <https://doi.org/10.21273/JASHS.132.5.603>
- He, H. & Zhang, L. B. (2022). *Polystichum gonggashanense* (Dryopteridaceae): a new fern from Sichuan, China. *Phytotaxa*, 559(2), 207-212. <https://doi.org/10.11646/phytotaxa.559.2.10>
- Jangi, F., Ebadi, M. T. & Ayyari, M. (2021). Qualitative changes in hyssop (*Hyssopus officinalis* L.) as affected by cold plasma, packaging method and storage duration. *Journal of Applied Research on Medicinal and Aromatic Plants*, 22, 100289. <https://doi.org/10.1016/j.jarmap.2020.100289>
- Khalid, S., Majeed, M., Ullah, M. I., Shahid, M., Riasat, A. R., Abbas, T., Aatif, H. M. & Farooq, A. (2020). Effect of storage conditions and packaging material on postharvest quality attributes of strawberry. *Journal of Horticulture and Postharvest Research*, 3(2), 195-208. <https://doi.org/10.22077/jhpr.2019.2826.1093>
- Kociecka, J., Liberacki, D. & Strozecki, M. (2023). The Role of Antitranspirants in Mitigating Drought Stress in Plants of the Grass Family (Poaceae)-A Review. *Sustainability*, 15(12), 9165. <https://doi.org/10.3390/su15129165>
- Kumar, R. & Singh, K. (2006). Effect of pre-storage pulsing on storage life of gladiolus cut spikes. *Journal of Ornamental Horticulture*, 9(4), 258-261.
- Kumar, V., Kumar, R., Khajuria, S., Khajuria, S., Gupta, S., Sinha, A. K., Raina, V., Jamwal, M., Dhotra, B. & Khajuria, V. (2023). Response of trickle irrigation and some antitranspirants on fruit yield and quality of Kinnow mandarin (*Citrus reticulata* Blanco) under rainfed condition. *The Pharma Innovation Journal*, 2023; 12(7), 3338-3342.
- Lai, N. Y. G., Kuah, A. T., Kim, C. H. & Wong, K. H. (2022). Toward sustainable express deliveries for online shopping: Reusing packaging materials through reverse logistics. *Thunderbird International Business Review*, 64 (4), 351-362. <https://doi.org/10.1002/tie.22259>
- Lambers, H., Chapin, F. S., & Pons, T. L. (2008). Plant physiological ecology. Springer Science, New York, pp. 99.
- Lufu, R., Ambaw, A. & Opara, U. L. 2020. Water loss of fresh fruit: Influencing pre-harvest, harvest and postharvest factors. *Scientia Horticulturae*, 272(102), 109519. [10.1016/j.scienta.2020.109519](https://doi.org/10.1016/j.scienta.2020.109519)
- Lv, G. L., Wanf, J. X., Feng, S. L., Yao, L. X. & Dang, Q. N. (2020). Effects of different types of anti-transpirants on water use efficiency of four herbaceous plants. *Acta Agrestia Sinica*, 28(3), 712-719. [10.11733/j.issn.1007-0435.2020.03.015](https://doi.org/10.11733/j.issn.1007-0435.2020.03.015)
- Moroni, F. J., Gascon-Aldana, P. J. & Rogiers, S. Y. (2020). Characterizing the efficacy of a film-forming anti-transpirant on raspberry foliar and fruit transpiration. *Biology*, 9(9), 255. <https://doi.org/10.3390/biology9090255>
- Mphande, W., Kettlewell, P. S., Grove, I. G. & Farrell, A. D. (2020). The potential of antitranspirants in drought management of arable crops: A review. *Agricultural Water Management*, 236, 106143. <https://doi.org/10.1016/j.agwat.2020.106143>
- Nguyen, T. H., Boonyariththongchai, P., Buanong, M., Supapvanich, S. & Wongs-Aree, C. (2021). Edible coating of chitosan ionically combined with κ-carrageenan maintains the bract and postharvest attributes of dragon fruit (*Hylocereus undatus*). *International Food Research Journal*, 28(4), 682-694. <https://doi.org/10.1016/j.scienta.2021.109916>
- Pacifici, S., Ferrante, A., Mensuali-Sodi, A. & Serra, G. (2007). Postharvest physiology and technology of cut eucalyptus branches: a review. *Journal of Agromedicine*, 137, 124-131.
- Patel, G. D., Patel, R. B., Patel, B. N., Singh, A., & Dhaduk, B. K. (2008). Effect of wrapping films and cold storage on post harvest life of spiderlily (*Hymenocallis littoralis*). *Journal of Ornamental Horticulture*, 11(3), 224-227.
- Pranuthi, P., Suseela, T., Swami, D.V., Suneetha, D. S. & Vani, V. S. (2018). Effect of Different Packing and Storage

- Conditions on Biochemical Parameters in Extending the Vase Life of Cut Carnation cv. Kiro. *International Journal of Current Microbiology and Applied Sciences*, 7(8), 1348-1355. <https://doi.org/10.20546/ijcmas.2018.712.158>
25. Punetha, P. & Trivedi, H. (2018). Analysis of antitranspirant chemicals in relation to the post-harvest attributes of cut rose cv. Naranjo. *International Journal of Chemical Studies*, 6(2), 1745-1749.
  26. Raza, A., Charagh, S., García-Caparrós, P., Rahman, M.A., Ogwugwa, V.H., Saeed, F. & Jin, W. (2022). Melatonin-mediated temperature stress tolerance in plants. *GM Crops & Food*, 13(1), 196-217. <https://doi.org/10.1080/21645698.2022.2106111>
  27. Reid, M. S., & Jiang, C. Z. (2012). Postharvest biology and technology of cut flowers and potted plants. *Horticultural Reviews*, 40, 1-54. <https://doi.org/10.1002/9781118351871.ch1>
  28. Revathi, P., Arun, M. N., Sukumar, K., Dhruva, S. & Bindu, G. S. (2021). Enhancing the water stress tolerance in soybean (*Glycine max* L.) through anti-transpirants and mulch. *Legume Research-An International Journal*, 44 (11), 1315-1321. 10.18805/LR-4216
  29. Rida, M. E. D. F. & El-Gedawey, H. (2022). Influence of some anti-transpirants on the longevity, quality and cold dry transportation of *Thaumatococcus bipinnatifidum* (selloum) cut leaves. *Scientific Journal of Flowers and Ornamental Plants*, 9(2), 115-132. 10.21608/SJFOP.2022.251807
  30. Saggoo, M. I. S. & Kaur, M. (2018). An infertile triploid population of *Polystichum squarrosum* from Himachal Pradesh. *Cytologia*, 83(2), 181-186. <https://doi.org/10.1508/cytologia.83.181>
  31. Sarkar, B., Tayyab, M., Kim, N. & Habib, M. S. (2019). Optimal production delivery policies for supplier and manufacturer in a constrained closed-loop supply chain for returnable transport packaging through metaheuristic approach. *Computers and Industrial Engineering*, 135, 987-1003. <https://doi.org/10.1016/j.cie.2019.05.035>
  32. Sharma, B. P., Beshir, H. M., Chaudhary, S. V. S. & Dilta, B. S. (2010). Effect of various wrapping materials and storage durations on post harvest life of asiatic hybrid lily cv.'apeldoorn'. *Annals of Horticulture*, 3(1), 69-74.
  33. Sharma, P. & Thakur, N. (2020). Effect of pulsing and storage methods for extending vase life of cut flowers. *International Journal of Chemical Studies*, 8(6), 1320-1328. <https://doi.org/10.22271/chemi.2020.v8.i6s.10943>
  34. Shinde, S. P., Chaudhari, S. R. & Matche, R. S. (2023). A way forward for a sustainable active packaging solution for prolonging the freshness and shelf life of *Rosa hybrida* L. cut flowers. *Postharvest Biology and Technology*, 204, 112475. <https://doi.org/10.1016/j.postharvbio.2023.112475>
  35. Singh, H. B. (1999). Potential medicinal pteridophytes of India and their chemical constituents. *Journal of Economic and Taxonomic Botany*, 23(1), 63-78.
  36. Sow, S. & Ranjan, S. (2021). Antitranspirants: A novel tool for combating water stress under climate change scenario. *Food and Scientific Report*, 2, 29-31.
  37. Voss, D. H., (2002). The Royal Horticultural Society Colour Chart.
  38. Wahba, H. E., Sarhan, A. Z., Salama, A. B., Sharaf-Eldin, M. A. & Gad, H. M. (2017). Effect of seasonal variation on the growth and chemical composition of *Cynara cardunculus* L. plants. *J. Mater. Environ. Sci*, 8, 318-323.
  39. Warmund, M. R., Trinklein, D. H., Ellersieck, M. R. & Smeda, R. J. (2021). Antitranspirants partially mitigate auxin herbicide injury on tomato plants. *HortScience*, 56 (8), 932-939. <https://doi.org/10.21273/HORTSCI15888-21>
  40. Yadav, P., Kashyap, B., Dhiman, S.R., Sharma, P., Gupta, Y.C. & Sharma, M. (2018). Effect of glycerine on drying of cut foliage of *Polystichum squarrosum* (D Don) Fee. *International Journal of Farm Sciences*, 8(3), 51-57. 10.5958/2250-0499.2018.00081.2
  41. Yang, L., Wang, X., He, S., Luo, Y., Chen, S., Shan, Y., Wang, R. & Ding, S. (2021). Heat shock treatment maintains the quality attributes of postharvest jujube fruits and delays their senescence process during cold storage. *Journal of Food Biochemistry*, 45(10), 1-16. <https://doi.org/10.1111/jfbc.13937>
  42. Zhang, L., Shi, X., Hou, H., Lin, Q., Zhu, S. & Wang, G. (2023). 6-Benzyladenine Treatment Maintains Storage Quality of Chinese Flowering Cabbage by Inhibiting Chlorophyll Degradation and Enhancing Antioxidant Capacity. *Plants*, 12(2), 334. <https://doi.org/10.3390/plants12020334>