



## Optimization of fermentation conditions for producing Indian rock bee (*Apis dorsata*) mead using response surface methodology

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**Abstract** Mead is a traditional drink which results from the alcoholic fermentation of diluted honey carried out by yeast (*Saccharomyces cerevisiae* KF233529). The present investigation was carried out for the optimization of fermentation parameters for maximizing the yield of ethanol. Response Surface Methodology (RSM) based central composite design was employed to obtain best combination of temperature, fermentation time and total soluble solids (TSS). The optimum conditions for ethanol yield were temperature 28°C, TSS 15°Brix and 6 days after fermentation. The model showed that the value of  $R^2$  (0.9998) was high and p-value of interaction of variance was <0.0001. Hence the model can be said to be of highly significant.

**Keywords:** Ethanol, Fermentation time, Mead, Response Surface Methodology (RSM), Temperature, TSS

### INTRODUCTION

Mead is one of the world's oldest alcoholic beverages, containing 8-18% (v/v) of ethanol, which results from the alcoholic fermentation of diluted honey carried out by yeast. Though mead is the oldest fermented product being used by man yet it is difficult to find it commercially (Pereira *et al.*, 2009) since mead producers face several problems, like delayed and arrested fermentation, production of off-flavours by the yeast and lack of uniformity of the final product.

Honey is a natural product, a highly concentrated solution of a complex mixture of sugars. It also contains small amounts of other constituents such as minerals, proteins, vitamins, organic acids, flavonoids, phenolic acids, enzymes and other phytochemicals. The components in honey responsible for its antioxidative effect are flavonoids, phenolic acids, ascorbic acid, catalase, peroxidase and carotenoids (Turkmen *et al.*, 2006; Bertoneclj *et al.*, 2007). The colour, flavour, aroma and yeast are important quality characteristics which in turn influence the quality of mead (Gupta and Sharma, 2009). Rock bees (*Apis dorsata*) are giant bees found all over India in sub-mountainous regions up to altitude of 2700 m. This honey has higher amount of enzymes, amino acids and minerals than *A. cerana* and *A. mellifera* honey. Fermentation process has both the nonlinear and dynamic properties. Considerable attempts have been made by several researchers to propose a methodology based on mathematical models. Major problems of fermentation process are that they need a large number

of experiments and often the models are very complicated to describe the experimental observation (Hajar *et al.*, 2012).

Optimization of process condition is one of the most critical stages in the development of an efficient and economic bioprocess (Karuppaiya *et al.*, 2009). The conventional one-factor-at-a-time approach of optimization is not only tiresome but also ignores to merge interaction of each factor. One of the most common optimization used in last two decades is the Response Surface Methodology (RSM). RSM is a powerful mathematical model with a collection of statistical techniques by which interaction between multiple processes variables can be identified with fewer experimental trials. It is widely used to examine and optimize the operational variables for experimental design, model developing, and test variable and condition optimization. There are various advantages in using statistical methodologies in terms of rapid and reliable short listing of process conditions, understanding interaction among them, and a tremendous reduction in total number of experiments, resulting in saving time, glassware, chemicals and manpower (Cheynier *et al.*, 1983).

In spite of various advantages, statistical designs have been applied to only limited number of aerobic submerged and solid state fermentation and anaerobic submerged fermentation processes deal with a large number of variables, and there are several reports on the application of RSM for the production of primary and secondary metabolites through microbial fermentation (Karuppaiya *et al.*, 2009). Although

many advances in the developments of mead have been made over the last few years, particularly in terms of optimizing ethanol concentration, there is still scope for future development. The present study was aimed to find out the optimum fermentation condition for producing mead as a health drink. RSM was used for optimization of ethanol concentration less than five.

## MATERIALS AND METHODS

**Yeast strain and culture growth conditions:** *Saccharomyces cerevisiae* (KF233529) has been isolated from honey and standardized in the Department of Agricultural Microbiology was used for this study. The yeast cells were grown in Yeast peptone dextrose agar (YPD), containing glucose 20g, peptone 10g, yeast extract 5g and agar 20g per litre. The culture was routinely maintained at 4°C on slants. Before use, the culture was transferred to YPD broth and incubated for 24 h at 27° C.

**Honey:** In the present study, rock bee honey was obtained from a local bee keeper at north-east region of Dindugal district, Tamil Nadu.

**Honey-must preparation and fermentation condition for mead fermentation:** Rock bee (*A. dorsata*) honey was diluted with tap water (35g: 85mL) and mixed to homogeneity. The insoluble solids were removed by filtering to obtain a clarified honey-must. Sulphur dioxide, in the form of potassium metabisulfite, was added up to a concentration of 100mg/L of free SO<sub>2</sub> to inhibit the bacterial growth. Starter culture was prepared by pre-growing the yeast culture in YPD broth for 24hrs. Incubation was done at 27°C with gentle orbital shaker at 120 rpm. Above honey must was inoculated with 4% inoculum with an initial population of 10<sup>5</sup> Colony-Forming Units (CFU/mL). Fermentation was carried out in 250 ml Erlenmeyer flasks with 100 ml honey must. Two days after aerobic fermentation, the Erlenmeyer flasks were water sealed.

**Estimation of Ethanol:** Ethanol content was estimated using Refractometer (%).

**Experimental design and response surface methodology:** Design Expert Software Version (8.0) was used to optimize the fermentation condition for Indian rock bee mead production. This software applies the principle of RSM to determine the optimal response. Three important factors, namely temperature (A), fermentation time (B) and TSS (C), considered as operating (independent) parameters, were selected to study their effect on ethanol production. Table 1 states the actual values and the coded values of the variables

employed. Coded values of +1, 0 and -1 correspond to high, medium and low values of variables, respectively. Ethanol percentage was regarded as the response or output variable (r). The central composite design (CCD) was used to access the effects of the three input independent parameters on the desired responses and build a second order (quadratic) model for the response variable (r). The statistically designed experiments comprised 8 factorial points, 6 axial points and 6 replicates at the centre points resulting in a total of 20 experiments. The ethanol percentage was observed from each of the 20 experiments analysed by Analysis of Variance (ANOVA) to determine the optimum conditions. The regression analysis was performed to fit the response.

## RESULTS AND DISCUSSION

### Interaction of factors and honey wine fermentation:

In mead fermentations, the fermentation process is influenced by the temperature. But temperature tolerance for growth of yeast and fermentation is strongly strain dependent (Rousseau *et al.*, 1992). In the honey wine fermentation, ethanol production was high at 28°C upto 6 days. The optimum yield for ethanol production was obtained at 28°C. Ethanol production was decreased with the increasing temperature. The reason was that after a period of time (6 days) this high temperature (34.73° C) inactivated the yeast cell. It was reported that ethanol producing yeast could grow rapidly at temperature 25-33°C (Ozcelik and Denli, 1996). At less than 25°C and more than 30°C, it was not favourable temperature for our yeast strain therefore at these stress conditions ethanol productions was lowest. At 32°C, the yeast cell was moderately activated and ethanol concentration was gradually increased with time upto 10 days. Therefore the yeast cells were very much affected by temperature. Temperature controls the cell viability, growth rate, exponential phase, enzyme activity and membrane function (Torija *et al.*, 2003).

Fermentation is slow in a medium containing low sugar, whereas its speed increases in must which have 15°Brix. Above this concentration, fermentation slows. Thus, an elevated amount of sugar hinders yeast growth and decreases the ethanol concentration (D'Amato *et al.*, 2006). It is known that the high substrate concentrations may cause osmotic shock of the yeast cells and slow down the mass and heat transfer. A decline of the ethanol concentration could be noticed because of the exhaustion of the release glucose and the transition of the yeast metabolism

**Table 1.** Natural levels, codes and intervals of variation of the independent variables in the design of experiments.

Process parameters	Codes	Levels					Interval of variation
		-1.682	-1	0	+1	+1.682	
Temperature (°C)	A	21.27	24	28	32	34.72	4
Fermentation time (days)	B	-0.72	2	6	10	12.72	4
TSS (°Brix)	C	-1.81	5	15	25	31.81	10

**Table 2.** Central composite design matrix of process parameters of independent variables and their corresponding experimental and predicted yields of ethanol.

Run No.	Independent variables			Ethanol (%)	
	Temperature (°C)	Fermentation time (days)	TSS (°Brix)	Observed	Predicted
1	-1	-1	-1	0.35	0.32
2	+1	-1	-1	0.50	0.48
3	-1	+1	-1	2.05	2.06
4	+1	+1	-1	2.45	2.44
5	-1	-1	+1	2.40	2.39
6	+1	-1	+1	2.29	2.27
7	-1	+1	+1	3.51	3.52
8	+1	+1	+1	3.62	3.63
9	- $\alpha$	0	0	2.72	2.73
10	+ $\alpha$	0	0	2.93	2.95
11	0	- $\alpha$	0	0.82	0.86
12	0	+ $\alpha$	0	3.48	3.46
13	0	0	- $\alpha$	0.49	0.51
14	0	0	+ $\alpha$	3.25	3.25
15	0	0	0	4.81	4.85
16	0	0	0	4.85	4.85
17	0	0	0	4.89	4.85
18	0	0	0	4.90	4.85
19	0	0	0	4.83	4.85
20	0	0	0	4.84	4.85

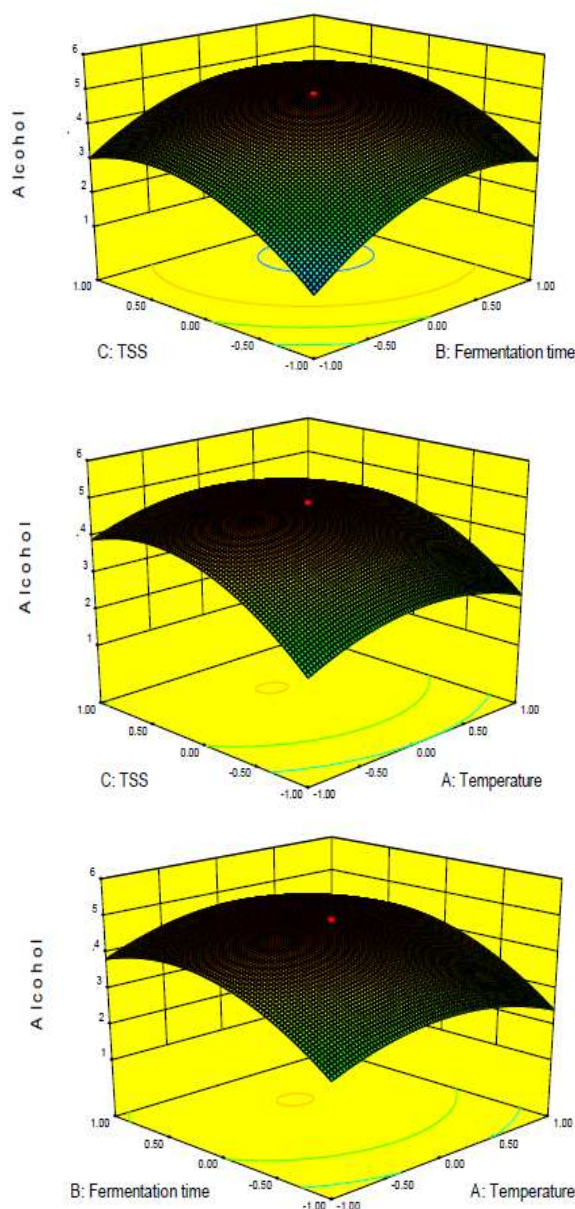
**Table 3.** Analysis of variance (ANOVA) for ethanol production using CCD.

Source	Sum of squares	Degrees of freedom	Mean square	F value	P value
Model	48.02	9	5.34	4810.82	<0.0001
Temperature (A)	0.06	1	0.06	53.86	<0.0001
Fermentation time (B)	8.17	1	8.17	7367.51	<0.0001
TSS (C)	9.04	1	9.04	8152.00	<0.0001
AB	0.02	1	0.02	24.90	0.0005
AC	0.03	1	0.03	34.09	0.0002
BC	0.18	1	0.18	165.02	<0.0001
A <sup>2</sup>	7.32	1	7.32	6601.84	<0.0001
B <sup>2</sup>	13.05	1	13.05	11762.67	<0.0001
C <sup>2</sup>	15.90	1	15.90	14337.80	<0.0001
Residual	0.01	10	1.109E-003		
Lack of fit	4.95E-003	5	9.914E-004	0.81	0.5895
Pure error	6.13E-003	5	1.227E-003		
Corrected total	48.03	19			
R <sup>2</sup> = 0.9998	Adjusted R <sup>2</sup> = 0.9996			C.V (%) = 1.11	

towards utilization of ethanol as a carbon source. Glucose and fructose utilization was almost completed within 6 days of fermentation time. The glucose and fructose consumption was in accordance with the results of ethanol concentration since the glucose and fructose was consumed as a carbon source by the yeast. Substrate inhibition significantly effect on

ethanol yield and their results concerning the substrate inhibition were in agreement with the results in this study (Nikolic *et al.*, 2009).

The 3D response surface plots described by the regression model were drawn to illustrate the effects of interaction of each independent variable (temperature, fermentation time and TSS) on the response variable



**Fig. 1.** Response surface curve showing the effects fermentation parameters on rock bee mead ethanol content (%).

(Bocchini *et al.*, 2002). The response surface plots with 3D response surface of the calculated model are shown in Fig. which indicates the relationship between the response and the experimental data. The ethanol yield was significantly affected by temperature, fermentation time and TSS where temperature produced greater effect. The point prediction tool of the software was used to determine the optimum values of the factors for maximum ethanol production: temperature 28°C, fermentation time 6 days and TSS 15°Brix. The result confirmed that the model was adequate for reflecting the expected optimization Hajar *et al.* (2012) also studied the fermentation parameters (pH, temperature, inoculums concentration, sugar concentration and time) for maximizing ethanol

production. They reported that it could be achieved at the conditions when inoculum concentration 6-14% (v/v), pH (4.0-6.0), sugar concentration (14-22° Brix), temperature (24-32°C) and time of incubation (30-54 hrs). Ghosh *et al.* (2012) optimized the process condition for palm wine fermentation using response surface methodology. In this study temperature, TSS and fermentation time were considered as independent variables.

**RSM analysis for the mead fermentation:** The mead fermentation was carried out by controlling various fermentation parameters which were important for production of ethanol. The average of the triplicate measurements of the ethanol concentration are shown in Table 2. Optimum ethanol concentration 4.84 % was determined at the optimum condition of 28°C temperature, 15° Brix and after 6 days. Statistical significance of honey wine fermentation model is explained by analysis of variance (ANOVA). Nature of fit of the regression model is determined by the adjusted co-efficient of determination ( $R^2$  adj). The high value of  $R^2$  adj 0.9996 indicates the goodness of fit of the regression equation. The predicted co-efficient of determination ( $R^2$  pred) value was 0.990.

The probability of p-value for models of less than 0.05 indicate that models were significant, p-value less than 0.0001 indicate the models were highly significant. So our model p value was <0.0001 it was highly significant. The words lack of fit refers to the fact that the simple linear regression model may not adequately fit the data. Our p value for lack of fit of model was insignificant it indicated that our experimental model system was statistically significant. Values for actual and predicted responses were very close because the correlation value,  $R^2 = 99.96\%$  that means the experimental data could be accepted (Samah, 2008).

Applying the multiple regression analysis on the experiment, the response variables and the test variables are related by following second order polynomial equation:

$$\text{Final equation in terms of coded factors} = + 4.85 + 0.06*A + 0.77*B + 0.81*C + 0.05*A*B - 0.06*A*C - 0.15*B*C - 0.71*A^2 - 0.95*B^2 - 1.05*C^2$$

Final equation in terms of actual factors =  $-37.86 + 2.51*$  temperature +  $0.86*$  fermentation time +  $0.46*$  TSS +  $3.67E-003*$  temperature \* fermentation time -  $1.71E-003*$  temperature \* TSS -  $3.78*$  fermentation time \* TSS -  $0.04*$  temperature  $0.05*$  Fermentation time<sup>2</sup> -  $0.01*$  TSS<sup>2</sup> Table 3 shows the response of the variables temperature, fermentation time, TSS, temperature<sup>2</sup>, fermentation time<sup>2</sup>, TSS<sup>2</sup> and fermentation time x TSS were highly significant with p-value of less than 0.0001. For temperature x TSS and temperature x fermentation time, p value <0.05 and therefore this value was significant. All the linear ( $A^2$ ,  $B^2$  and  $C^2$ ) and interactive (BC) effect of variables were highly significant for ethanol production (Table 4), as understood from their respective p values (p<0.0001).

**Table 4.** Significance of the regression coefficients of the model

Factor	Coefficient estimate	Degrees of freedom	Standard error	p value
Intercept	4.85	1	0.01	<0.0001
Temperature (A)	0.06	1	9.012E-003	<0.0001
Fermentation time (B)	0.77	1	9.012E-003	<0.0001
TSS (C)	0.81	1	9.012E-003	<0.0001
AB	0.05	1	0.01	0.0005
AC	-0.06	1	0.01	0.0002
BC	-0.15	1	0.01	<0.0001
A <sup>2</sup>	-0.71	1	8.773E-003	<0.0001
B <sup>2</sup>	-0.95	1	8.773E-003	<0.0001
C <sup>2</sup>	-1.05	1	8.773E-003	<0.0001

## Conclusion

This study optimized the ethanol yield using RSM. The RSM allowed a rapid screening of the important influence factors and development of a polynomial model to optimize the process parameters for enhancing ethanol yield. Data obtained from experiment were analysed with RSM software (Version 8) gave the optimum ethanol yield 4.85% was determined at the optimum condition of temperature 28°C, TSS 15°Brix and 6 days after fermentation. The significant regression equation or model at the 5% level with correlation value 99.96% was also obtained. Since none had attempted so far, to optimize fermentation condition for getting maximum mead yield, the current study would be the pioneering report.

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