

Response of walnut (*Juglans regia* L.) leaf aqueous extracts, on seed germination and seedling growth of wheat (*Triticum aestivum* L.) in agri-silvi system of Uttarakhand

Abhishek Bahuguna^{1,4}, Sandhya Bahuguna^{2*} and Birendra Prasad³

¹Shri Venkateshwara University, NH 24, Rajabpur, Gajraula, Dist. J. P. Nagar (Uttar Pradesh), INDIA

²Shridhar University, Pilani - Chirawa Road, Pilani (Rajasthan), INDIA

³Department of Plant Breeding, GBPUA&T, Pantnagar, Udham Singh Nagar (Uttarakhand), INDIA

⁴College of Forestry and Hill Agriculture, Ranichauri, Tehri Garhwal (Uttarakhand), INDIA

*Corresponding author. E-mail: sanbhi11@gmail.com

Received: July 2, 2013; Revised received: August 11, 2013; Accepted: September 22, 2013

Abstract: Effect of different doses of aqueous extracts of walnut leaf was studied on germinating seeds and early seedling growth of wheat variety (cv. VL-616) recommended for hills under West Himalayan agri-silvi system. Seven treatments comprised of distilled water control (0%), 10%, 20%, 30%, 40%, 50% and 60% concentration of leaf extracts were treated. The effect of aqueous extracts was found inhibitive; indicate a direct proportional relationship with concentration dependent manner on seed germination and subsequent seedling growth of wheat. The wheat variety exhibited extent of phytotoxicity at 60% extracts application in comparison to untreated control. Invariably there was a decrease in first count, germination, seedling root and shoot length, seedling fresh and dry weight and vigour index with increasing aqueous extracts concentration on germinating wheat, however the shoot length was observed maximum at 60% concentration.

Keywords: Germination, Leaf extract, Vigour index, Walnut, Wheat

INTRODUCTION

Walnut (*Juglans regia* L.) is a large deciduous fruit tree with long fragrant leaves, distributed in the Himalayas between 1375-3350 m. asl, extending in the west to Afghanistan and east to Bhutan (Prasad *et al.*, 2009). In the Himalayas, the walnut is one of the first species to lose its leaves, tree becoming leafless from September to October i.e. the right time of *rabi* crop sowing in hills (Dua *et al.*, 2007). Presence of trees in agri-silvi system results in direct exposure of associated crop to continuous release of chemicals. These chemicals influence local environment, germination and growth of plants. The effects of these chemicals on other plants are known as allelopathy to be dependent on the concentration released into the soil / environment (Tharayil *et al.*, 2006; Inderjit *et al.*, 2008; Tharayil *et al.*, 2008; Kaur *et al.*, 2009). Allelopathy involves a plant's secretion of biochemical materials into the environment to inhibit germination or growth of surrounding vegetation. Allelopathic effects of trees on agriculture crops are well documented. (Tripathi *et al.*, 1996; Kohli *et al.*, 2000).

Allelochemical released by trees inhibit seed germination (Singh and Bawa, 1982), reduce plant growth through inhibitive cell division (Baker, 1996), reduce mineral

uptake, increase or decrease respiration, inhibit protein and haemoglobin synthesis (Rice, 1974). Walnut are often found growing on landscape site and when certain other landscape plants are planted near or under this shade tree, they tend to retard germination, yellow, wilt and die. The chemical responsible for walnut allelopathy is juglone (Willis, 2000; Jose, 2002).

A colourless nontoxic reduced form called harmless hydro-juglone is abundant, especially in leaves, fruit hulls, stem and roots of walnut. When exposed to air or oxidizing substances, hydrojuglone is oxidized to its toxic form, juglone (Dana and Lerner, 1990; Bertin *et al.*, 2003). Rain washes juglone from the leaves and carries it into the soil. Thus neighbouring plants of the walnut are affected by absorbing juglone through their roots (Rietveld, 1983). In previous studies, effects of juglone and walnut leaf extracts on various plant species were investigated (Einhelling, 1986; Hejl *et al.*, 1993; Kocacaliskan; Terzi, 2001).

Therefore, it is important to select tolerant varieties of wheat for planting in areas adjacent to walnut trees. To the best of our knowledge, no research has been reported on the effect of walnut leaf extracts on germinating wheat. It is hypothesized that in a walnut based agri-silvi inter cropping system, juglone released from walnut trees could be an inhibitor to the germination and subsequent

seedling growth of wheat close to the walnut trees. Thus, the objective of this work was to test this hypothesis in a laboratory experiment.

MATERIALS AND METHODS

Leaves of more than ten years old walnut trees were used in obtaining the extracts because walnut trees younger than seven years old do not contain sufficient juglone to cause toxicity (Prataviera *et al.*, 1983; Piedrahita, 1984). The leaves of walnut were collected in the first week of August, 2012, since the juglone content of walnut was found to be highest in the last week of July and the first week of August (Tekintas *et al.*, 1988) and dried the leaves at 70°C in an oven for 48 h. Later mechanically crushed the dried leaves and made fine powder then soaking in distilled water at room temperature for 48 h. and mechanically stirred for one hour at end. The extracts thus obtained were filtered through Whatman No. 1 filter paper and stored in refrigerator until required. 100 g of crushed leaf powder were soaked in 1000 ml of distilled water for preparing 100% concentration of stock solution. Treatment consisted of five concentration of aqueous leaf extract (10, 20, 30, 40, 50 and 60 %) along with control on wheat variety VL-616 recommended for hills under west Himalayan agri-silvi system. 50 seeds of each treatment were placed separately in pre-sterilized petridishes with two fold filter paper at the bottom. The experiment was laid out in CRD with four replications. 10 ml. distilled water each of control and five concentration of leaf leachate were added in each petridishes on first day and 5 ml later or as and when required. Seeds were surface sterilized with 0.1% mercuric chloride solution. Petridishes were sterilized in hot air oven at 160°C prior to start the experiment. The petridishes were placed in an incubator at a temperature of 20°C. The first count and seeds germinated were counted daily for 4 and 8 days respectively. Root length, shoot length and seedling fresh weights of ten randomly selected seedlings from each treatment were recorded after ten days of the start of experiment (Anon, 1985). Seedling dry weight was measured after subjecting the samples in an air oven at 80°C for 24 h and attained constant weight for three consecutive readings. Vigour index I was calculated as a product of germination and seedling length, however, vigour index II was worked out by multiplying germination per cent with seedling dry weight (Abdul-Baki and Anderson, 1973).

RESULTS AND DISCUSSION

The effect of walnut leaf extracts on seed germination and seedling vigour characteristics are presented in table 1. There was significant gradual decrease in the germination percentage at first and final count with the increase in walnut leaf extracts and higher value (60.33

and 97.33 %) was observed for control, while application of 60% leaf extract resulted significantly lowest per cent germination (19.67 and 66.33 %) respectively. The result also depicted that each treatments of leaf extract concentration were differed significantly to each other with respect to germination for both first and final count. Thus there was an inhibitory effect on germination with increase in leaf extract concentration. This is in conformity with the findings of Orcutt and Nilsen (2000). Reduction in root, shoot and seedling length across increasing concentration of walnut leaf extracts up to 60% was noticed. Each treatment of walnut leaf extract had significant influence on root, shoot and seedling length over control (0%). The maximum root, shoot and seedling length of 14.52, 15.07 and 29.59 cm was observed for control, while lowest value of 3.44, 4.89 and 8.33 cm was measured for 60% treatment respectively. The reduction in seedling growth may be attributed to inhibitive cell division due to walnut leaf extracts. In the present study, walnut leaf extracts containing juglone significantly prevented root, shoot and as well as seedling elongation. Kocacaliskan and Terzi (2001) demonstrated that both juglone and walnut leaf extracts inhibit germination and seedling growth of several plant species such as watermelon, tomato, garden crest and cucumber (Tekintas *et al.*, 1988), for tomato and bean (Neave and Dawson 1989), wheat and corn (Jose and Gillespie, 1998), wheat (Prasad *et al.*, 2011). An inhibitory effect was noticed in the fresh and dry weight of seedling with the increase in leaf extract concentration from control to 60% and same trend was calculated in terms of vigour index I and II (Table 1). Least fresh weight (0.97 g) was observed for 60% concentration, while maximum seedling fresh weight of 2.73 g was observed from control treatment. The significantly maximum dry weight value of 0.45 g was recorded in untreated control, while significantly least (LSD<0.05) results (0.17 g) was observed at minimum concentration of leaf extracts (60%). Vigour index (Germination % × seedling length) and (Germination % × dry weight of seedling) is a real reflection of seedling vigour of seed/seed lot which were extremely reduced as the walnut aqueous leaf extracts concentration increased and statistically maximum value for vigour index I and II (2881.02 and 44.32) was computed for untreated control over all other treatments, while least value (552.36 and 11.07) was calculated also for 60% leaf extract concentration respectively. In several previous studies, it was determined that walnut leaf extracts decreased seed germination, seedling length along with seedling fresh and dry weight for various crops. Vigour index (I and II) is a multiple criteria of germination with seedling length and dry weight of seedling. Therefore, these indexes were markedly inhibited by the walnut leaf extract. This result is in close agreement with the findings of Kocacaliskan

Table 1. Effect of aqueous leaf extracts of *J. regia* L. on first count, germination (Final count), root-shoot elongation fresh & dry weight of seedling and vigour index I and II in wheat cv. VL- 616.

Treatment	First count (%)	Germination / final count (%)	Root length (cm)	Shoot length (cm)	Seedling length (cm)	Seedling fresh weight (g)	Seedling dry weight (g)	Vigour index I	Vigour index II	Relative growth index (RGI)
0% (control)	60.33	97.33	14.52	15.07	29.59	2.73	0.45	2881.02	44.32	62.00
10%	54.33	94.00	12.97	13.37	26.34	2.47	0.40	2476.72	37.32	57.80
20%	49.00	91.00	11.32	11.87	23.19	2.45	0.32	2094.61	28.55	54.25
30%	42.67	90.33	10.11	11.01	21.12	2.33	0.26	1922.72	23.96	46.88
40%	34.33	82.33	8.05	8.82	16.88	1.73	0.24	1388.90	19.53	41.72
50%	27.50	73.67	5.51	7.68	13.20	1.51	0.20	972.32	14.96	37.30
60%	19.67	66.33	3.44	4.89	8.33	0.97	0.17	552.36	11.07	29.66
CD @5%	3.30	2.70	1.13	0.93	1.89	0.66	0.13	181.63	13.83	3.88
CV	4.58	1.81	6.91	5.13	5.46	8.68	11.38	5.90	10.77	4.71
Sem	1.08	0.89	0.37	0.30	0.62	0.21	0.04	59.88	4.56	1.28
gm	41.11	85.00	9.41	10.38	19.80	2.02	0.29	1755.52	25.67	47.08

CD- Critical Difference; CV- Critical Variance; Sem- Stander Mean Error

and Terzi (2001) in watermelon, tomato, garden crest and alfalfa, Prasad *et al.* (2011) in cauliflower.

Germination rate traits in terms of relative growth index (RGI) was significantly reduced by walnut leaf extracts containing juglone and maximum value (62.00) for RGI were calculated in control (0%) treatment, while least value (29.66) was recorded respectively in undiluted extracts, however, the value for each and every treatment differed significantly with respect to RGI. RGI express the power of germination i.e. germination spread over the time. These findings support the earlier work where retard germination rate and percentage were observed following walnut leaf extracts and juglone of various plant species (Reynolds, 1987 and Rietveld, 1983). The delayed and unsynchronized germination might be attributed to interfere metabolic activities in the walnut leaf extracts subjected seeds (Terzi *et al.*, 2003).

Conclusion

As a conclusion, our results clearly revealed that aqueous leaf extracts of walnut had inhibitory effects on germinating wheat. Although allelopathic effects of walnut leaf extracts have been examined previously, no studies related to the effect of walnut leaf extract have been reported for wheat to date. However, wheat seed exhibited extent of tolerance and might be option in walnut intercropping under West Himalaya Agri-silvi system as a rabi cereal. Therefore, walnut leaf extracts phytotoxicity cannot be ruled out when examining the cause for observed reduction in seed germination and growth in wheat under walnut intercropping Agri-silvi system. A future study is necessary to get more detail information about walnut-wheat allelopathic relationships by using more wheat varieties.

REFERENCES

- Abdul-Baki, A.A. and Anderson, J.D. (1973). Vigour determination by multiple criterion. *Crop Science*, 13: 630-633.
- Anon (1985). International rules for seed testing. *Seed Science and Technology*, 13: 365-513.
- Baker, H.G. (1996). Volatile growth inhibitors produced by *Eucalyptus globulus*. *Madrono S., Francisco*, 18: 207-210.
- Bertin, C., Yang, X. and Weston, L.A. (2003). The role of root exudates and allelochemicals in the rhizosphere. *Plant Soil*, 256: 67-83.
- Dana, M.N. and Lerner, B.R. (1990). Black walnut toxicity. Purdue University Extension, West Lafayette, IN. pp 8.
- Dua, V.K., Govindakrishnan, P.M. and Lal S.S., (2007). Evaluation of wheat - potato relay intercropping system in the mid hills of Shimla. *Indian J. Agric. Res.*, 41(2): 142-145.
- Einhellung, F.A. (1986). Mechanism and mode of action of allelochemicals. In: Putman, A.R. and Tang C.S. (eds). *The Science of Allelopathy*, Wiley Inter Science, New York, pp. 171-188.
- Hejl, A.M., Einhellung, F.A. and Rasmussen, J.A. (1993). Effects of juglone on growth, photosynthesis and respiration. *Journal of Chemical Ecology*, 19: 559-568.
- Inderjit, Pollock, S.L. Callaway, R.M. and Holben, W. (2008). Phytotoxic effects of (±)-catechin in vitro, in soil, and in the field. *PLoS ONE* 3, e2536.
- Jose, S. (2002). Black walnut allelopathy : Current state of the science. In *Chemical ecology of plants: Allelopathy in Aquatic and Terrestrial Ecosystems*. Eds. Inderjit, Malik UA pp. Birknauser- Verlag. AG, Basal.
- Jose, S. and Gillespie, A. R. (1998). Allelopathy in black walnut (*Juglans nigra* L.) alley cropping: II. Effect of juglone on hydroponically grown corn (*Zea mays* L.) and soybean (*Glycine max* L.Merr.) growth and physiology. *Plant and Soil*, 203: 199-205.
- Kaur, H. *et al.*, (2009). Taking ecological function seriously: soil microbial communities can obviate allelopathic effects of released metabolites. *PLoS ONE* 4, e4700.
- Kocacaliskan, I. and Terzi, I. (2001). Allelopathic effects of walnut leaf extracts and juglone on seed germination and seedling growth. *Journal of Horticultural Science and Biotechnology*, 76: 436-440.
- Kohli, R.K., Singh, H.P., Batish, D.R. and Arora, V. (2000). Allelopathic interactions in some forests and plantation systems. In: *Man and Forest* (eds.) Kohli, R.K., Singh H.P., Vij. S.P., Dhir, K.K., Batish, D.R. and Khurana, D.K., Dept of Botany, P.U. Chandigarh: 135-154.
- Neave, I.A. and Dawson, J.O. (1989). Juglone reduces growth, nitrogenase activity and root respiration of actinorhizal back older seedlings. *Journal of Chemical Ecology*, 15: 1823-1836.
- Orcutt, D.M. and Nilsen, E.T. (2000). Physiology of plants under stress. *Soil and bioitic factors*. Wiley, New York.
- Piedrahita, O. (1984). Black walnut toxicity. Ministry of Agriculture and Food, Ontario, Canada. Factsheet 11: 7-8.
- Prasad, B., Lavania S. K. and Sah, V.K. (2011). Biassay study on effect of walnut leaf extracts on seed germination and seedling vigour of cauliflower (*Brassica oleracea* var. botrytis). *Indian Journal of Agro-Forestry*. 13(1): 103-106.
- Prasad, B., Prasad, R. and Sah, V.K. (2011). Effects of aqueous extracts of walnut (*Juglans regia* L.) leaf on germinating wheat (*Triticum aestivum* L.) in West Himalayan agri-silvi system. *Journal of Non Timber Forestry Products*, 18(1): 31-34.
- Prasad, B., Sah, V.K. and Prasad, R. (2009). Response of Aqueous Leaf Extracts of Walnut (*Juglans regia* Linn) on Germinating Wheat in North West Himalayan Agri-silvi System, New Delhi, 4th World Congress on Conservation Agriculture, February 4-7, 170-171.
- Prataviaera, A.G., Kuniyuki, A.H. and Ryogo, K. (1983). Growth inhibitors in xylem exudates of Persian walnut (*Juglans regia* L.) and their possible role in graft failure. *Journal of the American Society of Horticultural Science*, 108: 1043-1050
- Rice, E.L. (1974). Allelopathy. Academic Press, New York, pp. 422.
- Rietveld, W.J. (1983). Allelopathic effects of juglone on germination and growth of several herbaceous and woody species. *Journal of Chemical Ecology*, 9: 295-308.
- Singh, R. and Bawa, R. (1982). Effects of leaf leachates from *E. globulus* and *Aesculus indica* on germination of *Glaucium*

- flavum*. *Indian Journal of Ecology*, 9(1): 21-28.
- Tekintas, E., Tanrisever, A. and Mendilcioglu, K. (1988). Juglone'un tohum cimlenmesine etkileri. Ege Universi tesi, Ziraat Fakultesi, *Dergisi*, 25: 203-213.
- Terzi, I., Kocacaliskan, I., Benlioglu O. and Solak, K. (2003). Effect of juglone on growth on cucumber seedlings with respect to physiological and anatomical parameters. *Acta Physiologia Plantarum*, 25: 353-356.
- Tharayil, N., Bhowmik, P.C. and Xing, B. (2006). Preferential sorption of phenolic phytotoxins to soil: implications for the availability of allelochemicals. *J. Agric. Food Chem.* 54, 3033–3040.
- Tharayil, N., Bhowmik, P.C. and Xing, B. (2008). Bio-availability of allelochemicals as affected by companion compounds in soil matrices. *J. Agric. Food Chem.* 56, 3706–3713.
- Tripathi, S., Tripathi, A. and Banerjee, S.K. (1996). Comparative study of chemical nature and role of leaf and root leachates on crop productivity. *Advanced for Research India*, 14: 183-194.
- Willis, R.J. (2000). *Juglans spp.*, Juglone and allelopathy. *Allelopathy Journal*, 17: 1-55.