

Research Article

Fixing critical limits of boron in rice soils of Karaikal region, Puducherry union territory, India

Kowsalya, A*

Department of Soil Science and Agricultural Chemistry, Faculty of Agriculture, Annamalai University, Annamalai Nagar - 608002 (Tamil Nadu), India

M. V. Sriramachandrasekharan

Department of Soil Science and Agricultural Chemistry, Faculty of Agriculture, Annamalai University, Annamalai Nagar - 608002 (Tamil Nadu), India

P. Senthilvalavan

Department of Soil Science and Agricultural Chemistry, Faculty of Agriculture, Annamalai University, Annamalai Nagar - 608002 (Tamil Nadu), India

*Corresponding author. Email: kowsalya0395@gmail.com

Article Info

<https://doi.org/10.31018/jans.v14i2.3415>

Received: March 12, 2022

Revised: May 15, 2022

Accepted: May 21, 2022

How to Cite

Kowsalya, A. *et al.* (2022). Fixing critical limits of boron in rice soils of Karaikal region, Puducherry union territory, India. *Journal of Applied and Natural Science*, 14(2), 385 - 395. <https://doi.org/10.31018/jans.v14i2.3415>

Abstract

Boron is relatively immobile in plants and its deficiency can cause serious yield reduction in rice by retarding panicle formation. Karaikal region of Puducherry union territory includes different soils with varying fertility status thus supplying boron at optimum level is a must concern to enhance the rice productivity. A pot experiment was conducted to fix critical limits of boron in soils and rice of this Karaikal region. Forty surface soil samples (0-15 cm) were collected from Inceptisol, Entisol and Vertisol soil orders of different locations in Karaikal region. The experiment was conducted with three levels of boron viz., 0, 1 and 2 ppm applied through borax in a factorial completely randomized design with three replications. The available B content of soil was estimated with five extractants. The critical limits of hot water, hot 0.01 M CaCl₂, 0.05 M HCl, 1.0 M NH₄OAc, 0.01M CaCl₂+ 0.05 M Mannitol extractable B were found to be 0.50, 0.58, 0.46, 0.45 and 0.39 ppm in Inceptisol, 0.48, 0.59, 0.44, 0.39 and 0.48 ppm in Entisol and 0.45, 0.54, 0.45, 0.44 and 0.41 ppm in Vertisol and a critical limit of 31.0, 39.0 and 34.0 ppm B in rice plant for Inceptisol(0.5ppm), Entisol(0.48 ppm) and Vertisol(0.45 ppm) as determined by Cate and Nelson's graphical procedure. Among the extractants the hot water extractant showed the highest significant and positive correlation with Bray's percent yield(0.48,0.47 and 0.48), plant B content(0.041*, 0.019* and 0.271*) and B uptake(0.012*, 0.646* and 0.293*), respectively in Inceptisol, Entisol and Vertisol. From this study, the indicating knowledge of critical limit of boron will help to avoid yield loss of rice in the study region.

Keywords: Bray percent yield, Boron, Cate and Nelson's method, Critical limit, Rice

INTRODUCTION

Rice is the staple food for nearly half of the world's population that live in developing countries. The world's total area occupied by rice is one third to the total area planted to cereals. India has 43.78 m ha⁻¹ total rice cultivation area with average productivity of 4.05 t ha⁻¹. In Tamilnadu, rice is cultivated in an area of 1.81 m ha⁻¹ with the production of 63.08 lakh metric tonnes and average productivity of 2.8 tonnes ha⁻¹ (USDA 2020). In the Union Territory of Puducherry, the area under rice cultivation was 16,263 ha during the year 2015-2016, which accounted for 63.30 per cent of the total cropped area. Production of rice during 2015-16 was reported as 43,886 Mg as against 52,717 Mg in 2014-2015. The

average yield of rice was 2699 kg ha⁻¹ in 2016. In Karaikal, rice is grown in an area of 4447 ha with an average yield of 1727 kg ha⁻¹ with a total production of 7682 Mg (Directorate of Economics and Statistics, 2016).

The micronutrients in rice are one of the most important inputs for a better yield. The balanced use of micronutrients is essential for sustaining soil fertility and raising food grain production. Among micronutrients, boron (B) nutrition got a special importance because of the widespread deficiency of B in crops (Singh *et al.*, 2006; Singh, 2012; Prasad *et al.*, 2014; Sarangi *et al.*, 2016; Yoshinari and Takano, 2017). They further observed that, out of 33,000 soil samples, 45% samples were found to be deficient in B (0.5 mg kg⁻¹ hot water-soluble

B as the critical level). Boron deficiency is further affected by coarse texture, low pH, low organic carbon, and leaching under high rainfall (Singh, 2008; Wimmer et al., 2009; Jones et al., 2013; Dey et al., 2015;). Based on several field and green house experiments, Singh et al. (2006) and Rattan et al (2009) reported the critical value of B as 0.5 mg kg⁻¹. Crops grown on soils with available nutrient status below the critical limit are likely to be deficient and sensitive crops would show deficiency symptoms and respond well to the addition of that deficient nutrient (Wimmer et al. (2015); Powlson et al. (2011). The hot water-soluble B content of the black soil (Vertisol) was 0.6 and 0.7 mg kg⁻¹, respectively; these results are in accordance with the findings of other researchers, which revealed that the hot water-soluble B consists of a non-specifically adsorbed B fraction and it is very feebly held by divergent soil constituent's particles (Murthy et al., 2012). Consequently, with simple irrigation, this B fraction is released into soil solution and recognized as the most easily available B fraction for plant uptake (Renan and Gupta, 1999). There seems to be no report about the status of boron in soils of Karaikal region of Puducherry Union Territory. Hence, pot experiment was conducted to fix the critical limit of boron in soil and rice var. ADT-46 plant.

MATERIALS AND METHODS

A pot experiment with rice variety (ADT - 46) was conducted during 2020 in Karaikal. The 40 surface samples (0-15cm) coming under the order Inceptisol, Entisol and Vertisol were collected from different locations of Karaikal region. The initial soil samples were analysed for both mechanical and chemical compositions following standard methods (viz., Soil pH was measured in suspension of (1:2.5 soil: water) using a glass electrode standardized with pH 4.0, 7.0 and 9.2 buffer tablets attached to an Ion analyser, conductivity was measured in the same suspension using a conductivity meter and the cation exchange capacity was determined by Neutral normal ammonium acetate method (Jackson, 1973). The organic carbon content was determined by modified Chromic acid wet digestion titration method (Walkley and Black, 1934). CaCO₃ % was estimated by

Acid neutralization method (Allison and Moodie, 1965). The available nitrogen was determined by alkaline permanganate method (Subbiah and Asija, 1956). Available phosphorus (using 0.5 M NaHCO₃ of pH 8.5) was quantified by the spectrophotometer method (Olsen et al., 1954). Available potassium (using neutral normal ammonium acetate extract) was determined by Flame photometric method (Standford and English, 1949). Soils were analysed for available boron by different extractants (Table 1). There were three B treatments viz. B₀, B₁ and B₂ mg kg⁻¹. Each of the treatments was replicated thrice in a Factorial Completely Randomized Design (FCRD). The total number of earthen pots used in this study was 120. An amount of 10 kg of each soil was weighed and four rice seedlings were planted in each pot. The boron was applied through analytical grade di - sodium tetra borate (Na₂B₄O₇·10H₂O) as per treatment schedule (B₀- Control; B₁-0.04 mg/pot; B₂-0.08 mg/pot). The nutrients N, P₂O₅ and K₂O equivalent to @ 150:50:50 kg ha⁻¹ were applied as basal dose through urea, single super phosphate and muriate of potash, respectively to each pot. The plants were cut at the stage of 6 weeks, washed with distilled water and dried in an oven at 65°C for 24 hours. The extractable B content of soil (Wolf, 1971) and B content of plant (Gupta and Stewart, 1975) in the digest were determined by Spectrophotometer and B uptake was computed by dry matter production (DMP) multiplied with boron content.

The critical limit of B in soil and rice plant was determined by the graphical method proposed by Cate and Nelson's (1965). This procedure plotted a scatter diagram of the Bray's per cent yield on Y-axis versus soil test values on X-axis. For evaluation of different extractants, the co-efficient of correlation between amount of boron extracted by different extractants with Bray's per cent yield, tissue boron concentration and uptake of boron by rice

crop were worked out. Bray's percent yield was calculated by using the formula

$$\text{Bray's per cent yield} = \frac{\text{Yield without Boron} \times 100}{\text{Yield at optimum Boron}} \quad \text{Eq.1}$$

Table 1. List of different extractants used for extraction of available soil boron

Name of the extractants	Soil: solution ratio	Shaking time (min)	Reference
Hot water	1:2	5	Berger and Truog (1939)
Hot 0.01 M CaCl ₂	1:2	5	Aitken et al. (1987)
0.01 M CaCl ₂ + 0.05 M Mannitol (pH 8.5)	1:2	60	Cartwright et al. (1983)
1.0 M NH ₄ OAC	1:2	60	Gupta and Stewart (1975)
0.05 M HCl	1:2	60	Cartwright et al. (1983)

RESULTS AND DISCUSSION**Initial soils characteristics**

The experiment soils had pH range of 6.16 to 8.79. The soil was alkaline with mean of 8.06, EC ranged from 0.10 to 0.87 dSm⁻¹, non-saline with a mean of 0.43 dSm⁻¹, organic carbon ranged from 3.51 to 14.78 g kg⁻¹ with mean of 7.39 g kg⁻¹, CaCO₃ ranging from 0.53 to

3.53%, soil was non-calcareous with mean of 1.96%, the texture of the soil was sandy clay, sandy clay loam and clay (Table 2). The available N ranged from 198.8 to 588 kg ha⁻¹ with mean of 335.86 kg ha⁻¹, P ranged from (7.0 to 64.1 kg ha⁻¹) with mean of 23.18 kg ha⁻¹ and K ranged from (104.7 to 413.5 kg ha⁻¹) with mean of 262.89 kg ha⁻¹. The DTPA extractable B varied from (0.34 to 0.78 mg kg⁻¹) with mean of 0.50 mg kg⁻¹ (Table 3).

Table 2. Initial characteristics of soils used for the pot experiments

Soil order	Soil (Location)	Textural class	pH	EC (dSm ⁻¹)	OC (g kg ⁻¹)	CaCO ₃ (%)
Inceptisol	Madhur	Clay	8.02	0.40	7.61	1.25
	Madhur	Clay	8.28	0.56	7.46	1.02
	Madhur	Clay	7.58	0.48	7.90	0.98
	Madhur	Clay	8.01	0.44	6.00	1.03
	Madhur	Clay	7.45	0.36	8.05	1.23
	Madhur	Sandy clay loam	6.53	0.18	6.00	0.53
	Madhur	Sandy clay loam	6.16	0.10	5.41	0.76
	Madhur	Sandy clay	7.67	0.25	6.29	0.98
	Madhur	Sandy clay	8.55	0.53	8.34	1.32
	Madhur	Sandy clay	8.33	0.34	7.90	1.27
	Madhur	Sandy clay	8.72	0.42	8.20	1.10
	Madhur	Sandy clay	8.66	0.76	8.05	0.78
	Mupaithangudy	Sandy clay	8.61	0.44	10.39	0.75
	Mupaithangudy	Sandy clay	7.83	0.21	8.34	1.36
	Mupaithangudy	Sandy clay	7.62	0.31	9.37	0.79
	Serumavilangai	Sandy clay loam	8.32	0.39	4.24	0.99
	Serumavilangai	Sandy clay loam	8.86	0.45	4.10	1.52
Entisol	Thiruvengadapuram	Sandy clay	8.34	0.49	10.83	0.78
	Thiruvengadapuram	Clay	8.45	0.87	8.93	2.15
	Kezhaponpethi	Clay	8.06	0.83	9.51	1.36
	Kezhaponpethi	Sandy clay	7.72	0.53	6.59	0.96
	Kezhaponpethi	Sandy clay	7.73	0.38	11.56	3.43
	Thiruvettakudy	Sandy clay loam	8.32	0.39	7.46	1.28
	Thiruvettakudy	Sandy clay loam	8.14	0.44	6.59	1.12
	Thiruvettakudy	Sandy clay loam	8.22	0.52	8.93	1.34
	Thiruvettakudy	Sandy clay loam	7.89	0.40	7.46	0.86
	Thiruvettakudy	Sandy clay loam	8.51	0.53	6.73	1.52
	Thiruvettakudy	Sandy clay loam	8.81	0.57	10.68	2.23
	Thiruvettakudy	Sandy clay loam	8.30	0.40	7.76	1.34
	Thiruvettakudy	Sandy clay loam	8.36	0.48	7.32	1.45
Vertisol	Mathalangudy	Clay	7.28	0.32	14.78	1.46
	Mathalangudy	Clay	7.44	0.28	10.39	1.30
	Mathalangudy	Clay	8.08	0.23	3.51	0.78
	Mathalangudy	Clay	8.44	0.42	4.68	0.83
	Mathalangudy	Clay	8.56	0.51	4.98	1.05
	Mathalangudy	Clay	8.79	0.32	6.15	0.88
	Mathalangudy	Sandy clay loam	8.45	0.78	3.95	3.26
	Mathalangudy	Sandy clay loam	7.78	0.21	3.80	2.58
	Mathalangudy	Sandy clay loam	8.25	0.58	5.12	2.67
	Mathalangudy	Sandy clay loam	7.34	0.14	4.24	3.43
	Range	SCI – SC-C	6.16-8.79	0.10-0.87	3.51 -14.78	0.53 -3.43.
	Mean		8.06	0.43	7.39	1.96

Dry matter production, B content and uptake

Rice plants significantly ($p=0.05$) responded to boron application in all the soils (Table 4). DMP improved drastically to B applied in soils which were low in B content (0.4 ppm) compared to high B content (0.85 ppm) soils. The dry matter production was highest (48.9 g/pot) with application of 2 mg kg⁻¹ of boron and was sig-

nificantly higher over other levels of boron. The plant boron content and uptake were significantly influenced in all soils where B was applied at the rate 2 mg kg⁻¹. The plant B content in boron 2 mg kg⁻¹ treated pot ranged from 25.5 – 45.5 ppm with mean value 40.4 ppm and while 1 mg kg⁻¹ ranged from 21.5 – 45.5 ppm with mean value of 34.9 ppm and in control pot B con-

Table 3. Available nutrient status of soils used for pot experiments

Soil order	Soil (Location)	Available N (Kg ha ⁻¹)	Available P (Kg ha ⁻¹)	Available K (Kg ha ⁻¹)	Available B (mg kg ⁻¹)
Inceptisol	Madhur	305.2	25.8	344.6	0.73
	Madhur	322.0	20.2	345.5	0.85
	Madhur	378.0	14.2	288.9	0.71
	Madhur	523.6	16.7	342.4	0.47
	Madhur	380.8	11.3	322.4	0.40
	Madhur	327.6	25.9	104.7	0.61
	Madhur	240.8	17.2	178.0	0.45
	Madhur	288.4	17.4	181.2	0.78
	Madhur	330.4	21.2	257.3	0.43
	Madhur	266.0	44.2	210.2	0.65
	Madhur	344.4	48.8	216.2	0.72
	Madhur	386.4	17.8	255.6	0.48
	Mupaithangudy	291.2	25.8	251.1	0.85
	Mupaithangudy	341.6	20.8	365.1	0.69
	Mupaithangudy	347.2	20.9	188.3	0.81
	Serumavilangai	277.2	9.2	184.6	0.53
	Serumavilangai	257.6	9.8	185.6	0.68
Entisol	Thiruvengadapuram	302.4	26.6	220.5	0.47
	Thiruvengadapuram	319.2	12.6	275.2	0.41
	Kezhaponpethi	288.4	17.5	180.2	0.73
	Kezhaponpethi	260.4	17.6	310.2	0.56
	Kezhaponpethi	338.8	19.9	431.5	0.78
	Thiruvettakudy	330.4	35.1	301.9	0.83
	Thiruvettakudy	414.4	24.3	234.8	0.46
	Thiruvettakudy	299.6	17.1	331.8	0.65
	Thiruvettakudy	445.2	12.6	296.0	0.73
	Thiruvettakudy	260.4	9.2	253.1	0.81
	Thiruvettakudy	277.2	12.6	331.4	0.68
	Thiruvettakudy	294.0	7	349.0	0.41
Thiruvettakudy	291.2	7.2	314.1	0.75	
Vertisol	Mathalangudy	364.0	27.1	360.1	0.48
	Mathalangudy	459.2	30.6	111.4	0.64
	Mathalangudy	271.6	25.7	220.5	0.73
	Mathalangudy	585.2	64.1	355.0	0.50
	Mathalangudy	327.6	53.9	272.1	0.73
	Mathalangudy	198.8	20.6	281.7	0.46
	Mathalangudy	238.0	18.5	202.8	0.86
	Mathalangudy	355.6	28.6	211.0	0.72
	Mathalangudy	588.0	54.6	276.0	0.74
	Mathalangudy	316.4	16.9	173.5	0.77
	Range	198.8 – 588	7.0 – 64.1	104.7 – 431.5	0.34 – 0.78
	Mean	335.86	23.18	262.89	0.50

Table 4. Effect of added B on DMP, B content and uptake by rice and Bray's per cent yield (Mean value of three replications)

Soil (Location)	Dry matter weight production (g/ pot)			Bray's per cent yield			Boron content (ppm)			Boron uptake (mg/pot)		
	B ₀	B ₁	B ₂	B ₀	B ₁	B ₂	B ₀	B ₁	B ₂	B ₀	B ₁	B ₂
Inceptisol	Madhur	31.8	34.2	39.8	85.9	17.2	21.5	25.5	0.5	0.7	1.0	
	Madhur	38.3	48.9	54.8	73.9	18.6	29.3	32.6	0.7	1.4	1.8	
	Madhur	38.7	42.1	48.3	85.6	20.9	26.6	31.8	0.8	1.1	1.5	
	Madhur	30.6	37.5	39.6	79.4	24.5	28.6	32.9	0.7	1.1	1.3	
	Madhur	34.0	38.9	42.9	83.2	20.9	26.5	35.8	0.7	1.0	1.5	
	Madhur	34.3	38.4	44.2	83.0	27.4	31.4	36.8	0.9	1.2	1.6	
	Madhur	45.3	52.8	63.3	78.0	25.3	32.6	38.6	1.1	1.7	2.4	
	Madhur	43.6	54.4	72.8	68.5	28.9	32.8	39.9	1.3	1.8	2.9	
	Madhur	33.3	35.8	63.2	67.3	31.6	35.6	43.7	1.1	1.3	2.8	
	Madhur	33.6	42.6	56.9	67.6	29.7	38.9	44.8	1.0	1.7	2.5	
	Madhur	45.1	54.9	61.2	77.6	31.6	34.8	37.6	1.4	1.9	2.3	
	Madhur	25.4	31.9	50.9	61.4	32.5	35.1	44.8	0.8	1.1	2.3	
Mupaithangudy	36.9	44.5	49.9	78.1	35.6	38.4	42.9	1.3	1.7	2.1		
Mupaithangudy	48.7	52.1	61.8	85.5	38.6	41.2	52.5	1.9	2.1	3.2		
Mupaithangudy	39.2	42.2	52.8	82.5	28.4	32.5	43.5	1.1	1.4	2.3		
Serumavilangai	39.0	45.6	49.9	81.7	25.7	32.0	41.6	1.0	1.5	2.1		
Serumavilangai	42.7	49.8	50.2	85.5	28.4	33.8	43.7	1.2	1.7	2.2		
Entisol	Thiruvengadapuram	35.0	35.2	39.3	94.0	21.8	35.6	40.2	0.8	1.3	1.6	
	Thiruvengadapuram	30.2	36.9	37.9	80.7	18.0	32.3	38.5	0.5	1.2	1.5	
	Kezhaponpethi	33.7	41.9	46.2	76.4	20.9	35.9	41.3	0.7	1.5	1.9	
	Kezhaponpethi	33.9	43.4	46.2	75.6	27.4	38.4	42.3	0.9	1.7	2.0	
	Kezhaponpethi	45.8	59.2	60.5	76.4	28.2	39.1	45.6	1.3	2.3	2.8	
	Thiruvettakudy	27.1	30.6	50.0	67.3	32.6	38.3	44.5	0.9	1.2	2.2	
	Thiruvettakudy	28.9	49.5	56.7	54.5	31.6	35.3	38.3	0.9	1.7	2.2	
	Thiruvettakudy	28.7	36.1	38.9	76.4	32.9	41.2	43.7	0.9	1.5	1.7	
	Thiruvettakudy	27.5	32.1	38.7	77.8	28.9	37.4	40.8	0.8	1.2	1.6	
	Thiruvettakudy	34.1	54.6	64.4	57.3	35.6	41.9	43.0	1.2	2.3	2.8	
	Thiruvettakudy	38.0	42.7	47.8	84.0	35.8	45.5	49.2	1.4	1.9	2.3	
	Thiruvettakudy	32.5	36.2	43.9	81.1	33.7	40.9	44.4	1.1	1.5	2.0	
Thiruvettakudy	28.5	31.7	45.8	73.6	37.2	42.6	47.0	1.1	1.4	2.2		
Vertisol	Mathalangudy	32.2	37.5	42.6	80.4	23.3	29.0	31.5	0.8	1.1	1.3	
	Mathalangudy	31.5	36.1	39.8	83.0	28.6	32.2	35.3	0.9	1.2	1.4	
	Mathalangudy	27.6	35.4	40.7	72.5	31.7	38.9	41.6	0.9	1.4	1.7	
	Mathalangudy	33.6	41.8	43.9	78.4	29.5	40.5	43.7	1.0	1.7	1.9	
	Mathalangudy	33.5	38.6	41.1	84.0	26.9	31.7	37.8	0.9	1.2	1.6	
	Mathalangudy	34.3	37.7	39.7	88.6	28.4	32.0	38.3	1.0	1.2	1.5	
	Mathalangudy	34.8	38.0	40.2	89.1	31.5	34.4	41.7	1.1	1.3	1.7	
	Mathalangudy	37.5	42.7	49.3	81.6	30.8	32.6	37.7	1.4	1.9	1.9	
	Mathalangudy	41.1	49.2	53.3	80.1	26.6	32.9	38.8	1.1	1.6	2.1	
	Mathalangudy	36.3	42.0	44.7	83.7	32.4	35.7	41.1	1.2	1.5	1.8	
	Range	25.4-48.7	30.6-59.2	37.9-72.8	54.5-94.0	17.2-38.6	21.5-45.5	25.5-52.5	0.5-1.9	0.7-2.3	1.0-3.2	
	Mean	36.2	41.9	48.9	78.0	28.6	34.9	40.4	1.0	1.6	2.0	

[B₀-0 ppm ; B₁- 1ppm; B₂- 2 ppm (B₀- Control; B₁-0.04 mg/pot; B₂-0.08 mg/pot)]

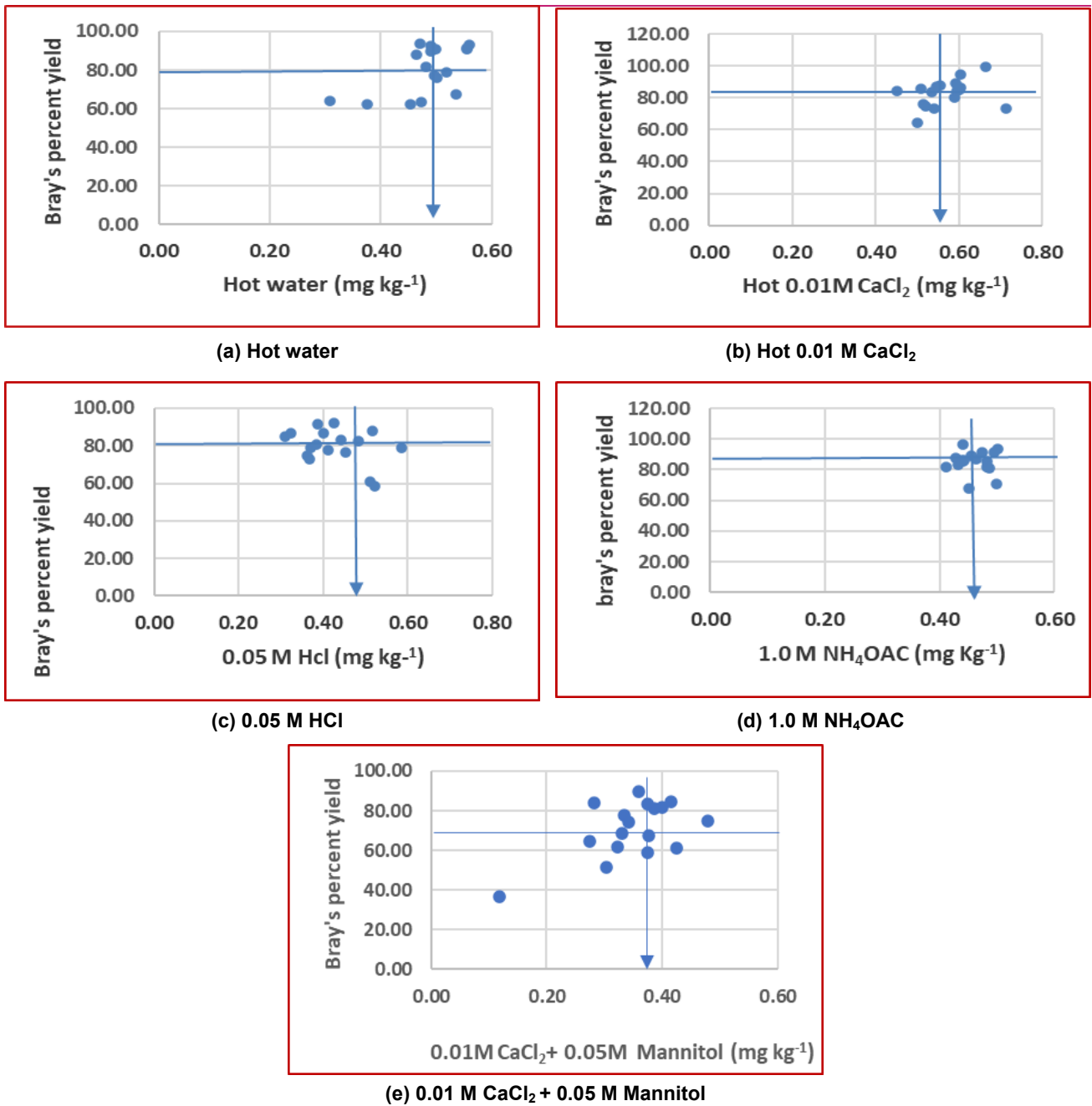


Fig. 1. Critical limit of B in *Inceptisol* among various extractants

tent ranged from 17.2-38.6ppm with mean value of 28.5ppm. The B uptake in B 2 mg kg⁻¹ treated pots ranged from 1.0 - 3.2 mg/pot) with mean value of 2.0 mg/pot, while 1 mg kg⁻¹ treated pots ranged from (0.7 – 2.3 mg/pot) with mean value of 1.5 mg/pot, in control pot, B uptake ranged from (0.5–1.9 mg/pot) with mean value of 1.0 mg/pot. The Bray's per cent yield ranged from (54.5 – 94.0) with mean value of 78 %. Boron deficiency is the second most important micronutrient constraint in soils in India after that of zinc (Zn)(Rehman et al.,2018). Hence, there was response to application of boron as noticed in the present study. The application of B resulting in increased DMP might be due to its favourable effect on the cell-dividing metabolic pathways

as reported by Hatwar et al., (2003). Positive influence of boron on DMP and boron uptake by rice silty clay, clay and calcareous soil was reported by Khan et al.(2006); Rehman et al.,2016; Ramesh and Rani (2017); Laik et al.(2021), respectively.

Evaluation of different extractants

The boron extracted by different extractants differed significantly among the soils (Table 5). The B extracted by hot water (0.35-0.58, 0.35–0.63 and 0.34–0.58 ppm) with mean value of (0.48, 0.47 and 0.48ppm), Hot 0.01 M CaCl₂ (0.44 – 0.71, 0.45 – 0.62 and 0.44 – 0.63ppm) with mean value of (0.53, 0.52 and 0.55ppm), 0.05 HCl (0.41 – 0.59, 0.36 – 0.60 and 0.40 – 0.35 ppm) with

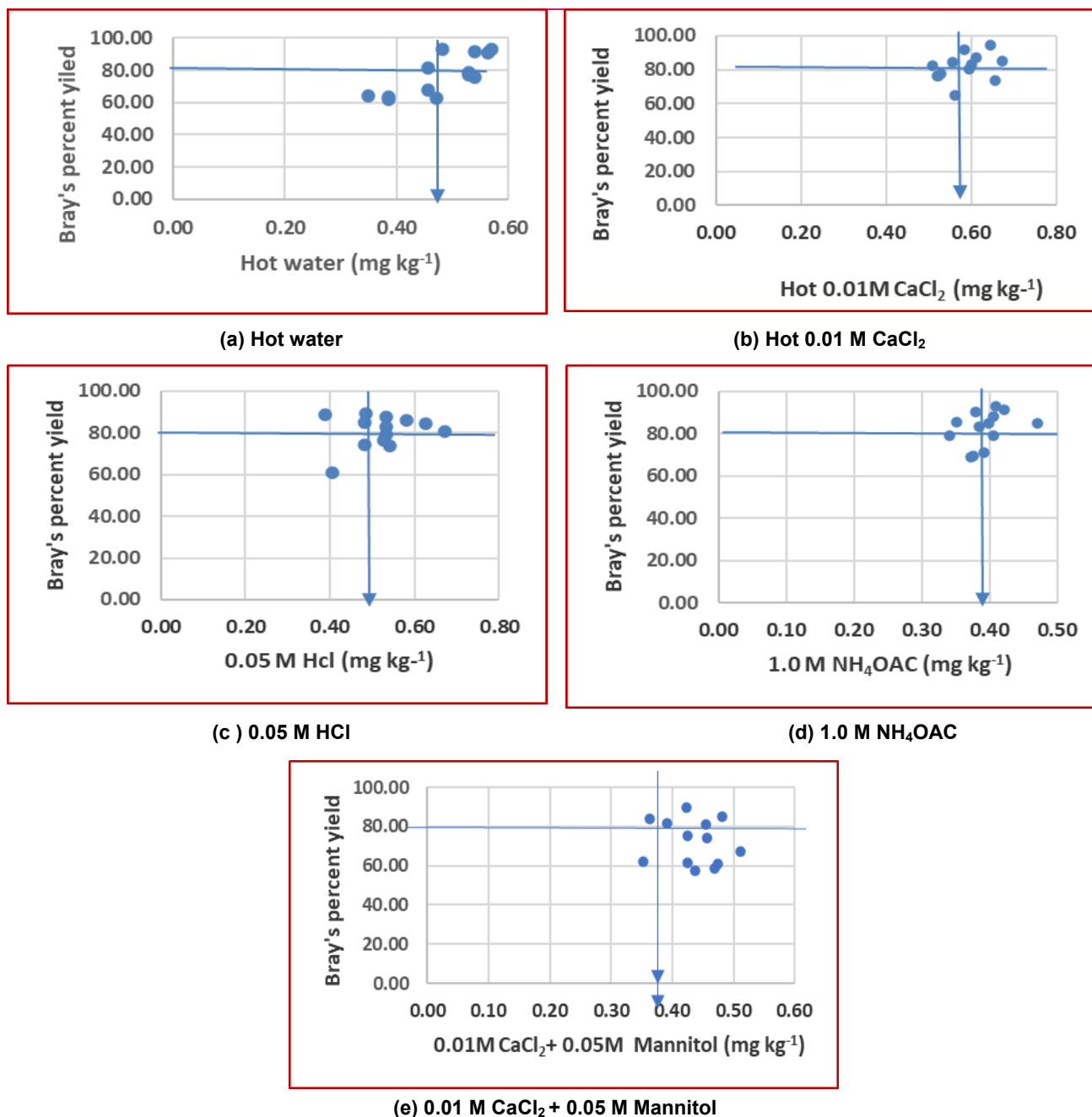


Fig. 2. Critical limit of B in Entisol among various extractants

mean value of { 0.50, 0.49 and 0.48 ppm}, 1.0 M NH₄OAc (0.41 – 0.49, 0.41 – 0.48 and 0.41 – 0.48ppm) with mean value of { 0.45, 0.45 and 0.47ppm} and 0.01M CaCl₂+ 0.05M Mannitol (0.37 – 0.48, 0.21 – 0.51 and 0.19 – 0.53ppm) with mean value of {0.42, 0.37 and 0.37ppm} in Inceptisol, Entisol and Vertisol, respectively. Based on the amount of B extracted by different extractants, the relative efficiency was of the following order Hot 0.01 M CaCl₂> 0.05M HCl > Hot water > 1.0 M NH₄OAc>0.01M CaCl₂+ 0.05M Mannitol (pH 8.5) in Inceptisol, Entisol and Vertisol. The amount of B extracted by different extractants were correlated with Bray's per cent yield, plant tissue and boron uptake by rice crop (Table 5). Among the extractants,hot

water correlated the highest with Bray's per cent yield (r = 0.629**, 0.762** and 0.587**) which gave positive and significant relationship as compared to other extractants in Inceptisol, Entisol and Vertisol and also hot water recorded significant positive correlation with B content (0.041*, 0.019* and 0.271*) and uptake (0.012*, 0.646* and 0.293*), respectively in Inceptisol, Entisol and Vertisol. Hence hot water was considered as the best B extractant for influencing available B in soils. The hot-water-soluble boron (HWS-B) extraction procedure was used as a benchmark to see any relative variation in available B in comparison with alternate extractants(0.0 M HCL, 0.018M CaCl₂, 1M NH₄OAc, 0.25 M sorbitol-DTPA,0.05 M Mannitol prepared in

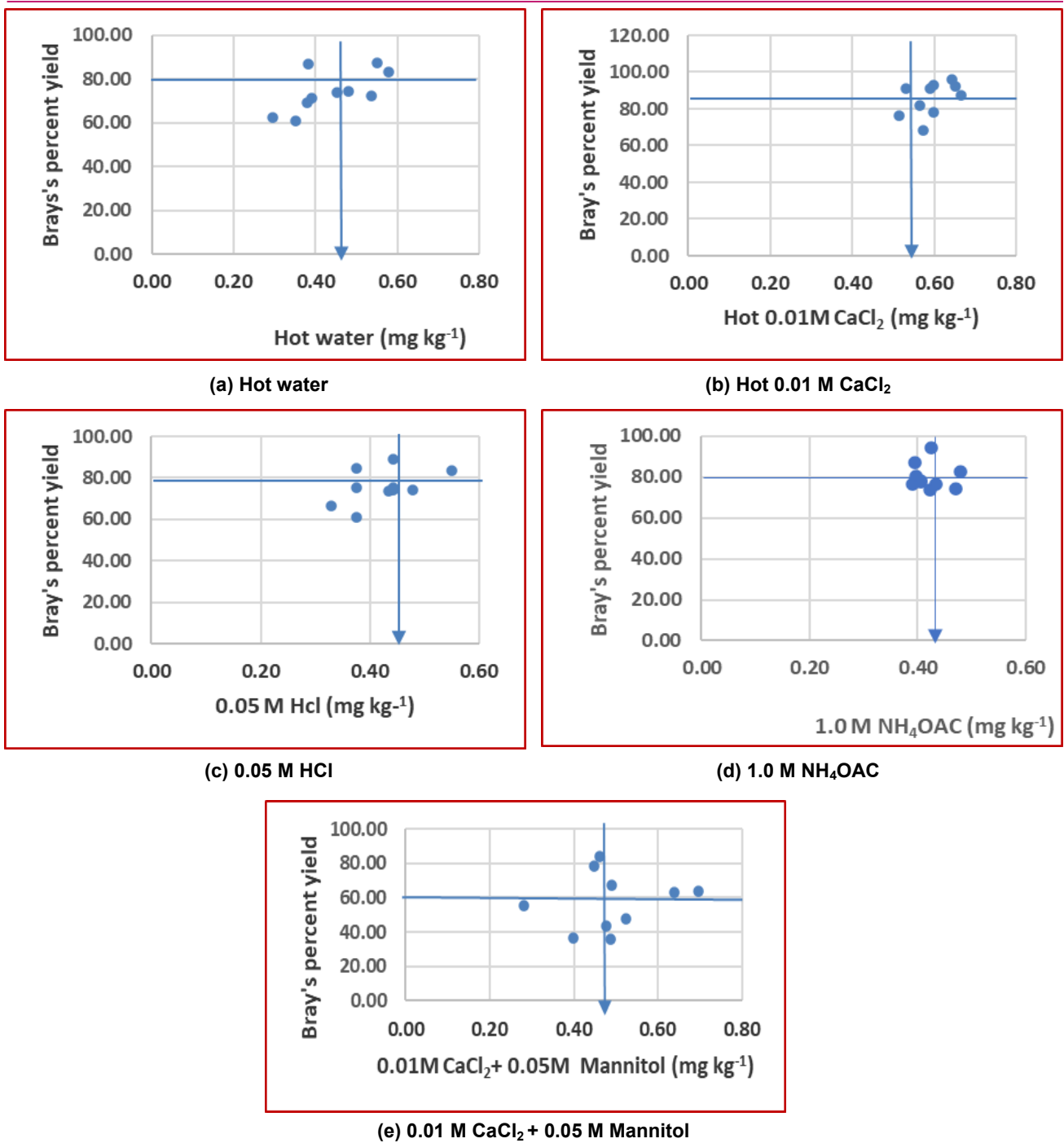


Fig. 3. Critical limit of B in Vertisol among various extractants

0.01M CaCl₂, 0.005 M AB-DTPA)(Niaz et al., 2011;Sofi Khurshid et al., 2017).

Critical limits of boron

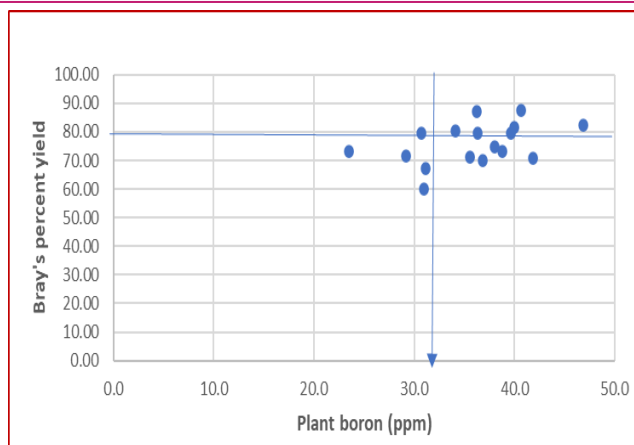
The critical limits for soil boron extracted by hot water, Hot 0.01 M CaCl₂, 0.05M HCl, 1.0 M NH₄OAc and 0.01M CaCl₂+ 0.05M Mannitol (pH 8.5) solution in Inceptisol, Entisol and Vertisol worked out to be 0.50, 0.58, 0.46, 0.45 and 0.39 ppm Inceptisol (Fig.1 a-e), 0.48, 0.59, 0.44, 0.39 and 0.38 ppm(Entisol (Fig.2 a-e) and 0.45, 0.54, 0.45, 0.44 and 0.41ppm Vertisol (Fig.3

a-e). The critical limits of B were fixed following the procedure of graphical method (Cate and Nelson,1965). From the correlation coefficient between B extractants and Bray's per cent yield, the critical limits of 0.50, 0.48 and 0.45 ppm obtained with hot water is the critical limits of B for rice soils of Karaikal region. The present result was in agreement with earlier workers (Marupaka et al.,2022; Samreen et al.,2022))and they reported that HWSBoron method was given better results than the other extractants tried. A critical limit of 31.0, 39.0 and 34.0 ppm in rice plants for Inceptisol,

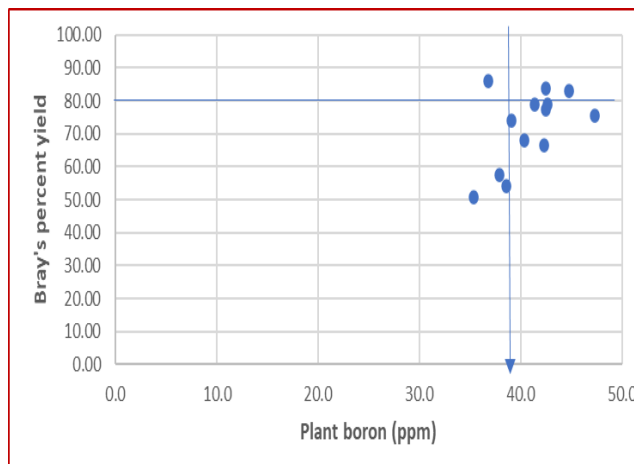
Table 5. Boron extracted and its relationship with Bray's per cent yield, plant boron and uptake (Mean value of three replications)

Extractant	Boron extracted(ppm)			Critical limits in soils			Correlation (r) with soil Boron at vegetative stage											
	Entisol			Entisol			Inceptisol			Vertisol								
	Range	Mean	Range	Mean	Range	Mean	BPY (ppm)	BPY (ppm)	Boron uptake Conc.	Boron uptake Conc.	Boron uptake Conc.	Boron uptake Conc.						
Hot 0.01 M CaCl ₂	0.44-0.71	0.53	0.45-0.62	0.52	0.44-0.63	0.55	0.60	0.208*	0.58	0.340*	0.57	0.457*	0.167*	0.082*	0.139*	0.175*	0.05*	0.059*
Hot water	0.35-0.58	0.48	0.35-0.63	0.47	0.34-0.58	0.48	0.49	0.629**	0.48	0.762**	0.45	0.587**	0.041*	0.012*	0.019*	0.646*	0.271*	0.293*
0.05M HCl	0.41-0.59	0.50	0.36-0.60	0.49	0.40-0.35	0.48	0.46	0.534**	0.44	0.585**	0.45	0.482*	0.543**	0.535**	0.478*	0.081*	0.226*	0.236*
1.0 M NH ₄ OAC	0.41-0.49	0.45	0.41-0.48	0.45	0.41-0.48	0.47	0.45	0.258*	0.39	0.372*	0.44	0.555**	0.522**	0.296*	0.084*	0.023*	0.270*	0.208*
0.01M CaCl ₂ +	0.37-0.48	0.42	0.21-0.51	0.37	0.19-0.53	0.37	0.39	0.582**	0.38	0.167*	0.41	0.160*	0.200*	0.299*	0.199*	0.279*	0.178*	0.240*
Mannitol(pH 8.5)																		

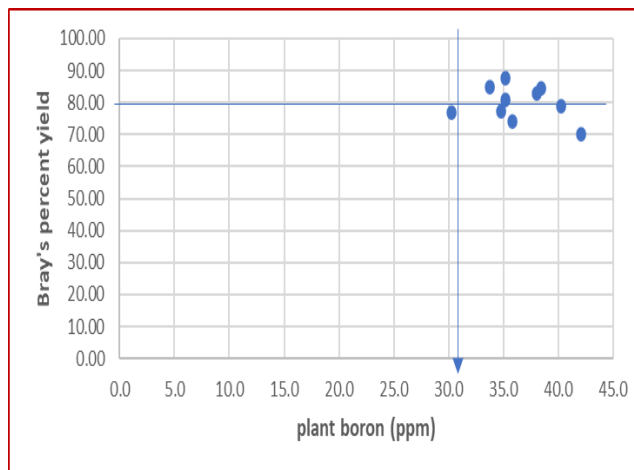
[Bo- 0 ppm ; B1- 1ppm; B2- 2 ppm (B₀- Control; B₁-0.04 mg/pot; B₂-0.08 mg/pot)]



Inceptisol



(b) Entisol



(c) Vertisol

Fig. 4. Critical limit of rice plant B

Entisol and Vertisol were determined (Fig.4). Based on the critical limit the percent boron deficient soils in Karaikal region was 67.5% and sufficient soil was 32.5%. With respect to soil orders, B deficient soil were 30.5, 37.5 and 20.9 % and B sufficient soil were 25.4, 31.4 and 24.2% in Inceptisol, Entisol and Vertisol respectively (Table 6).

Table 6. Response of rice to boron application in deficient and non-deficient soil (Mean value of three replications)

Boron status	Number of soils	Average DMP g pot ⁻¹			% Response
		B ₀	B ₁	B ₂	
Inceptisol					
Deficient < 0.59 mg kg ⁻¹	11	35.46	44.92	53.89	39.3
Sufficient > 0.59 mg kg ⁻¹	6	34.60	40.40	51.63	33.0
Entisol					
Deficient < 0.59 mg kg ⁻¹	8	32.91	41.12	49.04	37.0
Sufficient > 0.59 mg kg ⁻¹	5	32.10	40.24	44.81	32.5
Vertisol					
Deficient < 0.59 mg kg ⁻¹	7	34.59	40.28	44.14	22.0
Sufficient > 0.59 mg kg ⁻¹	3	33.37	38.99	42.08	21.5

[B₀- 0 ppm ; B₁- 1ppm; B₂- 2 ppm (B₀- Control; B₁-0.04 mg/pot; B₂-0.08 mg/pot)]

Conclusion

The study concluded that hot water was the better extractant for assessing the available boron status for Karaikal region soils. It is expected that rice crop will respond to B application when the soil contained less than 0.50 (Inceptisol), 0.48 (Entisol) and 0.45 (Vertisol) and rice plant with B concentration 31.0, 39.0 and 34.9 ppm. Based on the critical limit, the percent boron deficient soils in Karaikal region was 67.5% and sufficient soil was 32.5%. From the study, farmers of Karaikal region can optimize the boron application to rice soils.

ACKNOWLEDGEMENTS

The authors acknowledge the Department of Soil Science and Agricultural Chemistry, Faculty of Agriculture, Annamalai University, Annamalai Nagar for providing research facilities cum support for the conduct of the experiment.

Conflict of interest

The authors declare that they have no conflict of interest.

REFERENCES

- Aitken, R.L., Jeffrey, A.J. & Compton, B.L. (1987). Evaluation of selected extractants for boron in some Queensland soils. *Australian Journal of Soil Research*, 25, 263- 267.
- Allison, L.E., & Moodie. C.D. (1965). Carbonate: In C.A. Black et al. (ed.) *Methods of soil analysis. Part 2. 2nd Ed. Agron. Monogr.* 9. ASA, CSSA, and SSSA, Madison, WI.1379-1400. <https://doi.org/10.2134/agronmonogr.9.2.c40>
- Berger, K.C. & Truog, E. (1939). Boron determination in soils and plants using the quinalizarin reaction. *Ind. Eng. Chem. Anal. Ed.* 11,10, 540-553. <https://doi.org/10.1021/ac50138a007>
- Cartwright, B., Tiller, K.G., Zarcinas, B.A. & Spuoncer, L.R. (1983). The chemical assessment of the boron status of soils. *Australian Journal of Soil Research*, 21(3), 321-332. DOI: <https://doi.org/10.1071/sr9830321>
- Cate R.B. & Nelson, L.A. (1965). Graphical procedure for critical limits of nutrients. *Proc. Soil Sci. Soc. Am.* 89: 658.
- Dey, A., Dwivedi, B.S., Datta, S.P., Meena, M.C., & Agarwal, B.K. (2015). Soil boron status: impact of lime and fertilizers in an Indian long-term field experiment on a Typic Paleustalf. *Acta Agriculturae Scandinavica, Section B-Soil & Plant Science*, 65(1), 54–62.
- Directorate of Economics & Statistics (2016). Puducherry and Public Works Department, Directorate of Economics & Statistics Puducherry, 2016 - <http://agri.puducherry.gov.in/apstat.htm>
- Gupta, S.K. & Stewart, J.W.B. (1975). The extraction and determination of plant-available boron in soils, *Schweiz Landw. Forsch.*, 14, 153-169.
- Hatwar, G.P., Gondane, S.M., Urkade, S.M., & Ahukar, V. (2003). Effect of micronutrients on growth and yield of chilli. *Journal of Soils and Crops*. 13, 123–125.
- Jackson, M.L. (1973). *Soil chemical Analysis*. Prentice Hall of India (Pvt). Ltd., New Delhi.
- Jones, D.L., Cross, P., Withers, P.J., DeLuca, T.H., Robinson, D.A., Quilliam, R.S., Harris, I.M., Chadwick, D.R., & Edwards, G. (2013). Review: Nutrient stripping: The global disparity between food security and soil nutrient stocks. *Journal of Applied Ecology*. 50(4), 851–862.
- Khan, R., Gurmani, A.H., Gurmani, A.R., & Sharifzia, M. (2006). Effect of boron application on rice yield under wheat rice system. *Int. J. Agric. Biol.*, 8, 805–808.
- Laik, R., Singh, S.K., Pramanick, B, Kumari, V., Nath, D., Dessoky, E.S., Attia, A.O., Hassan, & M.M.; Hossain, A. (2021). Improved method of boron fertilization in rice (*Oryza sativa* L.)–Mustard (*Brassica juncea* L.) cropping system in upland calcareous soils. *Sustainability*, 13, 5037. <https://doi.org/10.3390/su13095037>
- Marupaka, V., Mylavarapu, R.S., Bergeron, J., Smidt, S.J., Hochmuth, G.J., & Santen, E.V. (2022). Comparing boron soil testing methods for coastal plain sandy Soils. *Communications in Soil Science and Plant Analysis*, DOI: <https://doi.org/10.1080/00103624.2022.2046041>
- Murthy, Y.L.N., Padmavathi, P. & Haripriya, Ch.V. (2012). Critical limit of boron for safflower (*Carthamus tinctorius* L.) grown in semi-arid Vertisol. *Journal of Oilseeds Research*, 29, 310-311.
- Niaz, A., Ahmad, W., Zia, M.H., and Ranjith, A.M. (2011).

- Relative efficiency of different extractants for available boron estimation in alkaline calcareous soils, *Communications in Soil Science and Plant Analysis*, 42, 1934-1944. DOI: <https://doi.org/10.1080/00103624.2011.591468>
17. Olsen, S.R., Cole, C.V., Watanabe, F.S., & Dean, J.A. (1954). Estimation of available phosphorus in soils by extraction with sodium bicarbonate. Circular, Vol-939 (p.19). Washington, DC: US Department of Agriculture.
 18. Powelson, D.S., Gregory, P.J., Whalley, W.R., Quinton, J.N., Hopkins, D.W., Whitmore, A.P., Hirsch, P.R., & Goulding, K.W.T. (2011). Soil management in relation to sustainable agriculture and ecosystem services. *Food Policy*, 36, S72–S87.
 19. Prasad, R., Kumar, D., Shivay, Y.S., & Rana, D.S. (2014). Boron in Indian agriculture—A review. *Indian journal of Agronomy*, 59(4), 511–517.
 20. Rattan, R.K., Patel, K.P., Manjaiah, K.M., & Datta, S.P. (2009). Micronutrients in soil plant animal and human health. *J. Indian Soc. Soil Sci.*, 57, 546–558.
 21. Rehman, A., Farooq, M., Nawaz, A., & Ahmad, R. (2016). Improving the performance of short duration basmati rice in water-saving production systems by boron nutrition. *Annals of Applied Biology*, 168(1):19-28. DOI: <https://doi.org/10.1111/aab.12237>
 22. Rehman, A., Farooq, M., Rashid, A., Nadeem, F., Bell, R.W., & Siddique, K.H.M. (2018). Boron nutrition of rice in different production systems—A review. *Agronomy for Sustainable Development*, 38:25 DOI: <https://doi.org/10.1007/s13593-018-05048>
 23. Remesh, R. & Rani, B. (2017). Effect of boron application through soil and foliar methods on the yield attributes and nutrient uptake of wet land rice. *Agric. Update*, 12, 301–304. DOI : [https://doi.org/10.15740/HAS/AU/12.TECHSEAR\(1\)2017/301-304](https://doi.org/10.15740/HAS/AU/12.TECHSEAR(1)2017/301-304)
 24. Renan, L. & Gupta, U.C. (1991). Extraction of soil boron for predicting its availability to plants. *Communications in Soil Science and Plant Analysis*, 22, 1003-1012.
 25. Samreen, T., Rashid, S., Nazir, M.Z., Riaz, U., Noreen, S., Nadeem, F., Kanwal, S., Munir, H., & TulMuntaha, S. (2022). Co-application of Boron, Sulphur, and Biochar for enhancing Growth and Yield of Brassica nopus under calcareous Soil. *Communications in Soil Science and Plant Analysis*, 53:9,1050-1067. DOI: [HTTPS://DOI.ORG/10.1080/00103624.2022.2043339](https://doi.org/10.1080/00103624.2022.2043339)
 26. Sarangi, D.R., Jena, D. and Chatterjee, A.K. (2016). Determination of critical limit of boron for rice, groundnut and potato crops in red and laterite soils of Odisha. *International Journal of Bio-resource and Stress Management*, 7(4):933-938. DOI: [HTTPS://DOI.ORG/10.23910/IJBSM/2016.7.4.1558](https://doi.org/10.23910/IJBSM/2016.7.4.1558)
 27. Singh, M.V. (2008). Micronutrient deficiencies in crops and soils in India. In: *Micronutrient Deficiencies in Global Crop Production*. Springer Netherlands, 93–125.
 28. Singh, M.V. (2012). Zinc and boron deficiency in Indian soils and advances in amelioration techniques. In: *Proceeding of Workshop on Micronutrient Deficiencies in Crops of Odisha and their Management*, Bhubaneswar Chapter of ISSS, OUAT, Bhubaneswar, India, 16–28.
 29. Singh, R.N., Singh, S., & Kumar, B. (2006). Interaction effect of sulphur and boron on yield, nutrient uptake and quality of characters of soya-bean (*Glycine max L. merill*) grown in acidic upland soil. *J. Indian Soc. Soil Sci.* 54, 516–518.
 30. Sofi Khurshid A., Ali Tahir., Uddin, K.M., & Bhat, M.A. (2017). Evaluation of suitable extractants for boron and establish its critical limits in soil and wheat plant grown under temperate condition of Kashmir Valley. *SKUAST Journal of Research*, 19(3), 143-146.
 31. Stanford, S. & English, L. (1949). Use of flame photometer in rapid soil tests of K and Ca. *Agronomy Journal*, 41: 446-447.
 32. Subbiah, B.V. & Asija, G.L. (1956). A rapid procedure for the estimation of the available nitrogen in soil. *Current Science*, 25, 259-260.
 33. USDA 2020. <https://www.ers.usda.gov>, o...PDF web results Rice outlook: May 2020-USDA ERS.
 34. Walkley, A. & Black, I.A. (1934). An estimation of the wet acid method for determining soil organic matter and a proposed modification of the chromic and titration method. *Soil Science*, 37, 29-39.
 35. Wimmer, M.A., Lohnit, G., Bassil, E., Muhling, K.H., & Goldbach, H.E. (2009). Membrane-associated, boron-interacting proteins isolated by boronate affinity chromatography. *Plant Cell and Physiology*, 50, 1292-1304. DOI: <https://doi.org/10.1093/pcp/pcp073>
 36. Wimmer, M.A., Goldberg, S., & Gupta, U.C. (2015). Boron. In: Barker, A.V., Pilbeam, D.J. (Eds.), *Handbook of Plant Nutrition*, Second Edition. CRC Press, 305.
 37. Wolf, B. 1971. The determination of boron in soil extracts, plant materials, composts, manures, water, and nutrient solutions. *Communications in Soil Science and Plant Analysis*, 2, 363-374.
 38. Yoshinari, A., & Takano, J. (2017). Insights into the mechanism underlying boron homeostasis in plants. *Frontiers in Plant Science*, 8, 1951. DOI: <https://doi.org/10.3389/fpls.2017.01951>