Effect of ethrel spray on the ripening behaviour of mango (*Mangifera indica* L.) variety 'Dashehari'


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Abstract: An experiment was carried out to investigate the effect of post harvest spray of different concentrations (100, 200, 400, 600 and 800 ppm) of ethrel (2-chloroethyl phosphonic acid) on ripening and colour development in ‘Dashehari’ mango fruits harvested in second week of June, 2015. The treated fruits were assessed for physico-chemical parameters such as physiological loss in weight (%), firmness (Kg/cm²), TSS (°Brix), titrable acidity (%), total carotenooids (mg/100g) and peel chlorophyll (mg/100g) and observations were recorded at 2 days interval during 8 days storage at ambient temperature. Changes in total soluble solids (8.5 to 23.23° Brix), total carotenooids (0.807 to 7.12 mg/100g) and PLW (14.58%) showed increasing trends up to 8 days during storage whereas fruit firmness (8.5 to 0.68 Kg/cm²), titrable acidity (1.26 to 0.08%) and total peel chlorophyll (5.2 to 0.14 mg/100g) showed decreasing trends. At the end of the storage period for 8 days, Ethrel spray at 600 ppm induced uniform ripening with attractive yellow colour within 4 days while untreated control fruits failed to ripen uniformly and remain light green even after 8 days of storage. Ripening advances by 4 days in fruits sprayed with 600 ppm ethrel compared to unsprayed control fruits.

Keywords: ‘Dashehari’ mango, Ethrel Spray, Ripening, Uniform colour

INTRODUCTION

Mango is one of the choicest fruit of India popularly known as ‘king of fruits’ continues to dominate the Indian fruit basket contributing 36 per cent to total fruit area and 20.3 per cent to total fruit production. It is grown over an area of 2.5 million ha with an annual production of 18.45 million tonnes in the country (Anonymous, 2014). Dashehari is one of the important variety of mango, largely being grown in northern states of India. It is known for its excellent flavor, fragrance and taste but suffers from poor colour development and uneven ripening causes economic losses to growers and traders. Being a climacteric fruit, uniform ripening and colour development can be induced by artificial means in mango. Now a day’s calcium carbide (CaC₂) popularly known as ‘masala’ is being indiscriminately used for artificial ripening of mango. However, it possesses impurities of arsenic and phosphorus hydrde which are extremely harmful for human health (Hossain et al., 2015). Further, calcium carbide is banned under Prevention of Food Adulteration (PFA) rules, 1955 and also under Food Safety and Standards (Prohibition and Restrictions on sales) regulations, 2011, its use is still unabated. Therefore, the need for alternative safe methods of artificial ripening is of the utmost importance for providing good quality and safe fruits for consumers.

The Food Safety and Standards (Prohibition and Restriction on Sales) Regulations, 2011, permitted use of Ethrel/Ethylene gas for the artificial ripening of fruits. Early and uniform ripening and colour development can be achieved by dipping of fruits in diluted ethrel (2 - Chloroethyl phosphonic acid) solution which is recommended for a number of climacteric fruits including mango (Venkatesan and Tamilmani, 2013, Gupta et al., 2015), banana (Kulkarni et al., 2011), Tomato (Dhall and Singh, 2013), papaya (Singh et al., 2012) and guava (Mahajan et al., 2008). Dipping of fruits in ethrel solution is cumbersome process and required more time and labour. Another method of ripening of mango is through exposure of fruits to ethylene gas in modern ripening chambers which requires huge investment and is not economical for farmers or small traders. Therefore, an alternative simple method of post harvest application of ethrel on fruits through hand spray is standardized for induction of uniform ripening and colour development in ‘Dashehari’ mango.

MATERIALS AND METHODS

Mango (*Mangifera indica* L. ‘Dashehari’) fruit were obtained from the experimental orchard of the ICAR-Central Institute for Subtropical Horticulture, Lucknow India. Mature green mango fruit with light-cream coloured pulp and total soluble solids of 8° – 9° Brix, titrable acidity 1.20-1.40%, were harvested in morning
hours. Immediately after harvesting, all fruit were brought to the handling and storage laboratory of ICAR-CISH. Healthy fruits (n = 720) of a uniform size, without any defect and free from visual blemishes, cuts, pests, or diseases were washed in water and spread on the floor. The experiments involved six treatments, each with three replications. The 720 fruits were divided into six lots of 120 fruits for each treatment. The five lots of fruits were sprayed with aqueous solutions of ethrel (100, 200, 400, 600 and 800 ppm) as a donor of ethylene gas with the help of hand sprayer, remaining one lot sprayed with water (the control). Following each treatment, the fruits were air-dried at room temperature, packed in corrugated fiberboard (CFB) boxes and stored under ambient conditions (RH: 70–80%, Temperature: 32° ± 2°C) for 8 days. After 2, 4, 6, and 8 days fruits (n = 30) from each treatment were sampled at random and used for ripening related observations. Post-treatment physiological loss in weight (PLW) in both ethrel treated and control fruits were recorded at each sampling interval up to 8 days. PLW was calculated as the difference between the initial weight of fruit (on day-0) and the weight of fruits at the time of sampling, expressed in terms of percentage. Firmness in treated and untreated fruits was measured at three points per fruit (without peel) using a ‘McCormick fruit tester FT 327’ penetrometer with head diameter of 11 mm. Fruit firmness was expressed in Kg/cm². Total Soluble Solids (TSS) were measured by using hand refractometer (Erma, Japan), while titratable acidity by titrimetric methods using 0.1N NaOH (Ranganna, 2000). Peel chlorophyll was measured by using hand refractometer (Erma, Japan), while titratable acidity by titrimetric methods using 0.1N NaOH (Ranganna, 2000). Peel chlorophyll was extracted in 80% acetone and estimated in spectrophotometer as method described by Pandey et al. (2015) with minor modifications. Total carotenoids of fruit pulp was extracted (by repeated extraction) with petroleum ether and acetone (3:2 v/v ratio, 60–80 °C) according to the method of Roy (1973). The ripening percentage of the fruits was estimated by counting the total number of ripened fruits on the basis of their appearance and desirable colour. The data obtained were subjected to statistical analysis by using ‘Statistical Software Package for Agricultural Research Workers’ (Sheron et al., 1998) software at 5% significance level.

RESULTS AND DISCUSSION

Effect of ethrel spray on peel chlorophyll degradation: Total chlorophyll content in peel of 600 and 800 ppm ethrel treated fruits decreased from 5.2 to 0.14 and to 0.058 mg/100 g, respectively after 8 days of storage. Whereas in control fruits, the total chlorophyll content decreased from 5.2 to 2.51 mg/100 g after 8 days of storage. Fruits sprayed with 600 and 800 ppm ethrel reached the full yellow stage 4 days earlier than the untreated fruits. The results indicated that the degradation of chlorophyll pigments in fruit peel in ethrel treated fruits was more rapid than in untreated fruits (Fig 1). This could be due to accelerated rate of diffusion of exogenous ethylene into peel of ethrel treated fruits which triggered the degradation of chlorophyll pigments (Terai et al. 1973). Ethrel was reported to accelerate chlorophyll degradation in mango fruits including mango (Mohamed and Goukh, 2003, Siddqui and Dhua, 2009, Gupta et al., 2015), Banana (Kulkarni et al., 2011) and papaya (Singh et al., 2012). Total carotenoids content: The total carotenoid content in ethrel treated fruits increased from 0.807 to 7.12 and 7.14 mg/100 g in 600 and 800 ppm of ethrel treated fruits, respectively after 4 days of treatment, whereas in untreated fruits pulp developed 7.05 mg/100 g even after 8 days of storage (Fig. 1). The improper development of total carotenoid pigments in pulp of control samples could be due to delayed ethylene biosynthesis. Higher carotenoid content in ethrel treated fruits could be due to enhancement in activity of carotenoid β-hydroxylase enzyme responsible for carotene synthesis (Yah et al., 1998). These results were in agreement with the findings of Siddqui and Dhua (2009) in ‘Himsagar’ mango dipped in 700 ppm ethrel and Singh et al. (2012) in papaya treated with 1000 ppm ethrel.

Physiological loss in weight (PLW): Data on the cumulative loss in weight due to transpiration and respiration processes indicated that fruits kept without ethrel spray lost weight up to 13.92% on 8th day of storage (Fig 2A). The highest PLW was observed with 800 ppm (15.78%) ethrel spray during ripening period of 8 days, resulted in shriveling, softening and over-ripening of fruits and hence found unsuitable. Ethrel spray of 600 ppm recorded 14.58% weight loss during ripening period of 8 days leading to uniform ripening and colour development and softening of fruits. Results showed that increase in PLW was directly proportional to increase in ethrel concentration. This increase in PLW of ethrel treated fruits during ripening could be due to upsurge in respiration rate leading to faster

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and uniform ripening compared to untreated fruits. Similar results were reported by Singh and Janes (2001) in mango and Mahajan et al. (2008) in winter guava fruits during ripening process caused by ethylene application.

**Fruit firmness:** Fruit firmness is one of the most widely used indicators of fruit quality. It influences appearance, texture and consumer acceptability of fresh fruits. Various industries use firmness tests as part of their quality control procedure (Jackson and Harker, 1997). The rapid decline in fruit firmness (shear force) values were recorded from 8.5 Kg/cm² at 0 day to 0.68 and 0.96 Kg/cm² in 800 and 600 ppm ethrel treated fruits, respectively at full ripe stage of 4 days (Fig 2B). Acceptable quality characteristics of Dashehari mango were considered as pulp firmness of 0.96 to 0.31 Kg/cm². Firmness value of less than 0.31 Kg/cm² were indicative of over ripe fruits (Fig. 1B). Firmness of fruits decreased at faster rate in treated fruits with increase in ethrel concentration could be due to enhance activity of polygalactouronase and pectin lyase enzymes result into breakdown of insoluble proteopecin into soluble pectin or by cellular disintegration leading to membrane permeability (Brinston et al. 1988; Ali et al., 1995; Yashoda et al., 2006). In control fruits, firmness of fruits decreased slowly from 8.5 Kg/cm² at 0 day to 4.56 Kg/cm² at 4th day of storage. Similar results were obtained by Mohamed and Goukh (2003) in mango, Kulkarni et al., (2011) in banana.

**TSS and total titrable acidity:** TSS increased with increase in the concentration of ethrel during ripening. The TSS of pulp during ripening increased from 8.5° B (Brix) to 22.73° B in 400 ppm, 23.23° B in 600 ppm and 23.56° B in 800 ppm ethrel treated fruits at the end of 8 days storage. In unsprayed fruits, the change in TSS content in the pulp was steady and reached to a maximum of 18.42° Brix after 8 days of storage (Fig...
The increase in TSS of fruit pulp could be due to the breakdown of starch into soluble sugars. The results indicated that the conversion of starch into sugars was rapid in ethrel treated fruits than in untreated fruits. This could be due to the rapid induction of pre-climacteric and climacteric phases and onset of climacteric peak in respiratory metabolic pathways in starch hydrolysis (Marriot, 1980). Total titratable acidity reduced in all fruits irrespective of treatment though reduction of acidity is more pronounced in ethrel treated fruits compared to control (Fig. 2D). The decline in acidity on ripening of fruits appears to result, at least in part, from the conversion of acids into sugars and their derivatives and their further utilization in metabolic process i.e. respiration in the fruit. These findings are in the line with the reports of Yashoda et al. (2006) in Alphonso mango, Mahajan et al., (2008) in winter guava fruits treated with 750 ppm aqueous solution of ethrel and Kulkarni et al., (2011) in banana treated with 500 ppm ethrel who reported the similar results.

Ripening of fruits: For initial 2 days the fruits remained hard and green in all the treatments except 600 and 800 ppm ethrel treatments where 20 and 40% fruits are ripened (Table 1). However, on fourth day there was dramatic increase in ripening of fruits and highest ripening percentage (100%) of mango fruits was observed after 4 days with ethrel 600 ppm and 800 ppm while lowest was in control fruits (20%). After 6 days, 100% ripening takes place in 400 ppm ethrel treated fruits while in control only 50% fruits were ripened. Similar results were obtained by Mahajan et al. (2010) in ethrel treated banana fruits. The improvement in ripening of ethrel treated fruits is due to multifunctional nature of ethylene which coordinates expression of genes that are responsible for a variety of processes, including a rise in respiration and activities of ACC synthase and ACC oxidase, autocatalytic ethylene production and changes in color, texture, aroma and flavor (Kader and Mitcham 1994).

Cost of ripening: Ripening cost was calculated as Rs. 0.13-0.40 per kilogram fruits for the best treatment i.e. 600 ppm ethrel spray which was lowest among all ripening methods in vogue.

Conclusion

Post harvest spray of mature Dashehari mango fruits with aqueous solution of ethrel (600 ppm) followed by air drying, packing in CFB boxes and storage at room temperature ensures faster and uniform ripening in 4 days with development of uniform attractive yellow colour, desirable firmness and consumer acceptability. This method may be adapted by the growers and small traders as replacement to calcium carbide treatment for ripening.

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