



Chlorophyll and carotenoid content of wheat (*Triticum aestivum* L.) seedlings under heat stress as affected by trehalose application

Aparjot Kaur* and S. K. Thind

Department of Botany, Punjab Agricultural University, Ludhiana-141004 (Punjab), INDIA *Corresponding author. E-mail: aparjotranu@gmail.com

Received: June 12, 2016; Revised received: October 24, 2016; Accepted: August 5, 2017

Abstract: Presently, chlorophyll and carotenoid contents were evaluated under control $(25\pm2^{\circ}C)$, heat stress $(35\pm2^{\circ}C \text{ and } 40\pm2^{\circ}C)$ and interactive effect of heat stress and trehalose in six wheat (*Triticum aestivum* L.) genotypes (HD2967, PBW175, C306, PBW343, PBW621 and PBW590). Trehalose an osmoprotectant, at concentration of 1mM and 1.5mM was applied at 7 days after sowing (DAS) followed by heat stress of $35\pm2^{\circ}C$ (moderate) and $40\pm2^{\circ}C$ (severe) on 8DAS for 4 and8 hours. As chloroplast thylakoid membranes, are highly vulnerable to heat stress, the chlorophyll content decreased with increased temperature stress in all selected genotypes. Heat stress significantly reduced (P< 0.05) the carotenoid content in all genotypes. Severe heat stress (8 hours) more adversely affected these mentioned parameters. The application of Trehalose @ 1.5mM as compared with 1mM concentration was found more effective to ameliorate the adverse effect of heat stress on chlorophyll and carotenoid contents to sustain photosynthetic process.

Keywords: Heat stress, Photosynthetic pigments, Trehalose, Wheat

INTRODUCTION

All over the world wheat i.e. *Triticum aestivum* L. is an necessary cereal crop. In the form of various products wheat is used as staple food. Global wheat supplies are predicted remain commodious in 2016/17 marketing season. Although below the 2015 record, world wheat production in 2016 is to outstrip utilization for the fourth consecutive season, boosting world stocks to a 15 year high. World wheat utilization is seen to decline slightly mostly because of reduced feed use (FAO 2016). It has been found that wheat contain the higher value of vegetable protein in the food of human beings, as it contains higher content of proteins and amino acids than the other major cereals like maize rice (NPCS board, 2012).

As heat stress or temperature stress is described as increase in temperature up to a threshold level for a particular interval of time that cause the unrecoverable harm to the plant or crop growth and development. A sudden rise in temperature basically above $10-15^{\circ}$ C is consider heat shock or heat stress (Wahid *et al.*, 2007). Due to massive rise in the population the requirement or desire for wheat has been also increased than the previous times, but its yield or harvest decreased due to the environmental stress. Extreme or late temperature stress during the grain filling period of normal as well as delayed planted wheat is the large abiotic stress that severely reducing wheat yield (Khan *et al.*, 2007). Unstable or stable heat stress causes various changes in morpho-anatomical, biochemical and physiological

modifications in wheat, that severely affect the crop growth and development. Heat stress violently decreased both quantity and quality of wheat (Wardlaw *et al.*, 2002, Altenbach *et al.*, 2003, Dupont *et al.*, 2006).

Trehalose is a resolveable, non-reducing disaccharide of glucose. Three isomers exist: α , α -trehalose, α , β trehalose and β , β - trehalose. Of these, only α , α trehalose (1-O- (α -Dglucopyranosyl) α -glucopyranosi de) is found in biological material. It is present in a huge variety of organisms and can serve as reserve of carbohydrate and as a protectant in response to different environmental stress factors. Trehalose is known to protect biological membranes and macromolecules. Its stockpilling has been implicated in permit crops to indulge stress, including heat-stress. Trehalose does protect against desiccation in certain specialized resurrection plants. Gomez et al., 2011 described the discovery of trehalose metabolism in the recent years has pointed out the importance of trehalose biosynthesis in stress responses in plants. Therefore, a primary aim and the significance of this work was to determine the role of trehalose, whether exogenous application of trehalose helps to protect and maintain the chlorophyll and carotenoid content from the destructive effect of heat stress when wheat seedlings exposed to different $(35\pm2^{\circ}C \text{ and } 40\pm2^{\circ}C)$ levels of heat stress.

MATERIALS AND METHODS

Plant material: Six genotypes of wheat (T. aestivum

ISSN : 0974-9411 (Print), 2231-5209 (Online) All Rights Reserved © Applied and Natural Science Foundation www.jans.ansfoundation.org

L.) viz. HD 2967, C306, PBW621, PBW590, PBW343 and PBW175 were obtained from Department of Plant Breeding and Genetics (PAU) and used for studies related to chlorophyll and carotenoid content under control and different heat stress levels. Statistical Analysis: Analysis of variance (ANOVA), critical difference at 5% level of significance (P< 0.05%) was used for the data analysis.

With a view of assess the effect of heat stress on above stated parameters, only healthy seeds of six genotypes of wheat were used in experiments. Seeds were surface sterilized with 0.1 per cent mercury chloride for 2-3 min. to avoid any kind of mycosis during seed germination. Petri plates were sterilized in oven at 100°C for 1 hour. Ordinary blotting papers were used in Petri dishes and were autoclaved before use. Twenty seeds were sown in each Petri-dish lined with circular blotting paper and incubated at $25\pm2°C$ temperature. On seventh DAS trehalose (1mM and 1.5mM) application was given followed by heat stress, incubated at 35°C and 40°C, for 8 hrs. Controlled Petri-dishes were placed in an BOD in which heat was maintained at 25°C.

T1- control at 25°C, T2-T1+(tre-1mM), T3-T1+(tre-1.5mM), T4- at 35°C, T5-T4+(tre-1mM), T6-T4+(tre-1.5mM),T7- at 40°C, T8-T7+(tre-1mM), T9-T7+(tre-1.5mM)

Chlorophyll content and carotenoid content: Chlorophyll content and carotenoid contentwas determined by method as followed by Hiscox and Isrealstam (1979).

The photosynthetic pigments from the wheat leaves by placing the 100g of fresh leaves used in the photosynthesis and reflectance measurements in 5ml of the dimethyl sulfoxide (DMSO) and extracting for 12 h in the dark. The concentration of the extracted pigments was calculated by recording the absorbance values at 665,645 and 480 nm.

Chl a = 12.19 (OD 665)-3.45(OD 645) x V/1000xW Chl b= 21.99(OD 645)-5.32(OD665) x V/1000xW Total Chl = 20.2(OD 480) + 8.02(OD665) xV/1000 x W Carotenoids=(OD480)+0.114(OD665)-0.638(OD645) where, $OD_{663} = OD$ at 663 nm; $OD_{645} = OD$ at 645 nm; $OD_{480} = OD$ at 480 nm; V = Total volume of solution made; W = Weight of sample (g) taken

The chlorophyll and carotenoid contents were expressed as mg chl g^{-1} fresh weight.

RESULTS AND DISCUSSION

Chlorophyll content: During the present study six genotypes of wheat (T.aestivum L.) were exposed to varying levels of heat stress $(35\pm2^{\circ}C \text{ and } 40\pm2^{\circ}C)$ under laboratory conditions (Table 1-6)) and contents of chlorophyll a, b and total were estimated. The chl a content was in range of 0.598 to 0.646 and chl b in range of 0.312 to 0.432 in control. The contents of total chlorophyll varied significantly (P< 0.05) all treatments in all the wheat genotypes studied presently and was recorded more in PBW621 and HD2967 genotypes and least in PBW590. Under severe heat stress i.e. 40±2°C for 8 hours duration chlorophyll a value was recorded more in PBW621 genotype and chl b was more in HD2967 and PBW621 genotype. These results were similar to that of Gautam et al. (2016) as heat stress decreased the chlorophyll a content in the durum wheat genotypes. Khan et al. (2015) also observed that heat stress significantly reduced chl a and chl b contents of wheat genotypes similar to that of Ahmed and Hassan (2011) and Kumar et al. (2012).Presently, it has been also recorded that in all the studied genotypes the chlorophyll content decreased with increased temperature stress. Shirdelmoghanloo (2016) also observed that there was positive correlation between heat stress and chlorophyll degradation or chlorophyll loss. Chloroplast thylakoid membranes, are highly vulnerable to heat stress. High temperatures led not only to disintegration of the lipid bilayer (Losa et al., 2004), but also to damage of the oxygen- evolving complex of photosystem 2 (Komayama et al., 2007). There is an inverse correlation between growth temperature and membrane saturation level. Changes in lipid-protein interactions are thought to play a major role in heat-induced increase in

Table 1. Effect of trehalose on chlorophyll a (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (4hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Treatments	Genotypes							
1 reatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.646	0.628	0.643	0.632	0.646	0.598		
T2-T1+(tre-1mM)	0.647	0.638	0.644	0.636	0.646	0.598		
T3-T1+(tre-1.5mM)	0.649	0.639	0.649	0.639	0.649	0.599		
T4- at 35°C	0.632	0.622	0.628	0.628	0.633	0.596		
T5-T4+(tre-1mM)	0.634	0.626	0.629	0.629	0.638	0.596		
T6-T4+(tre-1.5mM)	0.638	0.631	0.631	0.630	0.642	0.599		
T7- at 40°C	0.618	0.598	0.611	0.599	0.617	0.566		
T8-T7+(tre-1mM)	0.619	0.598	0.613	0.614	0.619	0.572		
T9-T7+(tre-1.5mM)	0.626	0.600	0.614	0.615	0.621	0.578		
CD 5%			V=0.077, T=0	0.094, V×T=0.232	2			

Table 2. Effect of trehalose on chlorophyll a (mg gm ⁻¹ fresh v	weight) content of wheat genotypes under heat stress (8hrs) of
$35\pm2^{\circ}C$ and $40\pm2^{\circ}C$.	

Treatments	Genotypes							
I reatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.641	0.622	0.632	0.631	0.645	0.598		
T2-T1+(tre-1mM)	0.646	0.626	0.638	0.636	0.654	0.602		
T3-T1+(tre-1.5mM)	0.451	0.631	0.642	0.636	0.661	0.636		
T4- at 35°C	0.590	0.546	0.588	0.566	0.600	0.532		
T5-T4+(tre-1mM)	0.596	0.546	0.591	0.566	0.616	0.538		
T6-T4+(tre-1.5mM)	0.598	0.548	0.596	0.571	0.617	0.542		
T7- at 40°C	0.538	0.510	0.536	0.518	0.586	0.506		
T8-T7+(tre-1mM)	0.548	0.516	0.568	0.532	0.588	0.516		
T9-T7+(tre-1.5mM)	0.551	0.518	0.572	0.549	0.589	0.526		
CD 5%			V=0.042, T	=0.051, V×T=0.12	26			

Table 3. Effect of trehalose on chlorophyll b (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (4hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

T	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.415	0.404	0.413	0.405	0.432	0.312		
T2-T1+(tre-1mM)	0.416	0.406	0.414	0.406	0.433	0.318		
T3-T1+(tre-1.5mM)	0.417	0.406	0.415	0.406	0.439	0.332		
T4- at 35°C	0.400	0.359	0.398	0.398	0.406	0.312		
T5-T4+(tre-1mM)	0.400	0.386	0.399	0.398	0.406	0.313		
T6-T4+(tre-1.5mM)	0.401	0.400	0.399	0.398	0.408	0.313		
T7- at 40°C	0.358	0.315	0.354	0.341	0.370	0.306		
T8-T7+(tre-1mM)	0.358	0.318	0.359	0.342	0.381	0.309		
T9-T7+(tre-1.5mM)	0.359	0.335	0.360	0.346	0.385	0.311		
CD 5%			V=0.042, T	=0.052, V×T=0.12	27			

Table 4. Effect of trehalose on chlorophyll b (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (8hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Treatments	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.418	0.402	0.410	0.406	0.432	0.330		
T2-T1+(tre-1mM)	0.418	0.412	0.411	0.407	0.433	0.330		
T3-T1+(tre-1.5mM)	0.419	0.413	0.412	0.407	0.433	0.331		
T4- at 35°C	0.361	0.316	0.343	0.342	0.370	0.296		
T5-T4+(tre-1mM)	0.376	0.318	0.346	0.348	0.371	0.303		
T6-T4+(tre-1.5mM)	0.376	0.349	0.348	0.356	0.372	0.326		
T7- at 40°C	0.306	0.296	0.298	0.296	0.306	0.217		
T8-T7+(tre-1mM)	0.307	0.297	0.298	0.296	0.316	0.218		
T9-T7+(tre-1.5mM)	0.307	0.297	0.299	0.297	0.317	0.276		
CD 5%			V=0.0023, T=	=0.0029, V×T=0.0	071			

the fluidity of the thylakoid membranes (Larkindale *et al.*, 2004).

In the present study it was also recorded that the application of trehalose increased the chlorophyll content in all genotypes. The application of higher concentration (1.5mM) of trehalose showed more increase in chlorophyll content as compare to 1mM of trehalose. Similar finding of Benaroudj *et al.*, 2001 also pointed out that trehalose accumulation during heat stress protects the cells and cellular proteins of *Saccharomyces cerevisiae* from damage by oxygen radicals. During heat stress, trehalose pretreatment protects the ultrastructure of chloroplasts some polypeptides in thylakoid membranes, and also improves the photosynthetic capacity of thylakoids , which indicates a protective role of trehalose or its metabolite for the thylakoid membrane.

Carotenoid content: Heat stress decreased the carotenoid content in all the selected wheat genotypes (Table 7 and 8). The carotenoid content recorded between 0.0291 to0.0298. The maximum carotenoid content was recorded in HD2967 and minimum was observed in PBW175 genotype. In all the genotypes it was recorded that the carotenoid content showed maximum value at 25±2°C as compared to moderate and severe heat stress. It was found all the genotypes showed increase in carotenoid content with the application of Trehalose. The concentration of 1mM of Trehalose showed increase in PBW621 and PBW590 under control conditions, whereas the concentration of 1.5mM of trehalose showed increase in carotenoid content in all the selected genotypes both in control and heat stressed conditions. The heat stress of 40±2°C (severe heat

Table 5. Effect of trehalose on total chlorophyll (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (4hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Treatments	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	1.055	1.036	1.049	1.049	1.059	0.938		
T2-T1+(tre-1mM)	1.056	1.037	1.049	1.049	1.059	0.941		
T3-T1+(tre-1.5mM)	1.058	1.038	1.049	1.051	1.069	0.943		
T4- at 35°C	0.916	0.906	0.930	0.936	0.940	0.836		
T5-T4+(tre-1mM)	0.917	0.906	0.939	0.936	0.941	0.838		
T6-T4+(tre-1.5mM)	0.918	0.907	0.942	0.937	0.942	0.839		
T7- at 40°C	0.736	0.846	0.746	0.741	0.726	0.696		
T8-T7+(tre-1mM)	0.736	0.847	0.746	0.742	0.727	0.697		
T9-T7+(tre-1.5mM)	0.737	0.848	0.747	0.743	0.727	0.699		
CD 5%			V=0.0114, T=	0.0140, V×T=0.0	344			

Table 6. Effect of trehalose on total chlorophyll (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (8hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Treatments	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	1.046	1.038	1.042	1.038	1.056	0.941		
T2-T1+(tre-1mM)	1.049	1.038	1.041	1.038	1.057	0.941		
T3-T1+(tre-1.5mM)	1.049	1.039	1.047	1.038	1.058	0.942		
T4- at 35°C	0.841	0.846	0.846	0.900	0.896	0.806		
T5-T4+(tre-1mM)	0.842	0.847	0.847	0.918	0.897	0.807		
T6-T4+(tre-1.5mM)	0.843	0.848	0.848	0.939	0.899	0.808		
T7- at 40°C	0.696	0.636	0.626	0.641	0.730	0.616		
T8-T7+(tre-1mM)	0.697	0.637	0.627	0.642	0.734	0.616		
T9-T7+(tre-1.5mM)	0.698	0.638	0.628	0.643	0.736	0.617		
CD 5%		V	=0.0152, T=0.	0187, V×T=0.045	58			

Table 7. Effect of trehalose on carotenoid (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (4hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Treatments	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.0298	0.0291	0.0295	0.0296	0.0293	0.0294		
T2-T1+(tre-1mM)	0.0298	0.0291	0.0295	0.0296	0.0294	0.0295		
T3-T1+(tre-1.5mM)	0.0299	0.0293	0.0296	0.0297	0.0296	0.0297		
T4- at 35°C	0.0280	0.0282	0.0276	0.0278	0.0279	0.0276		
T5-T4+(tre-1mM)	0.0281	0.0281	0.0283	0.0278	0.0281	0.0283		
T6-T4+(tre-1.5mM)	0.0282	0.0282	0.0281	0.0279	0.0282	0.0284		
T7- at 40°C	0.0253	0.0255	0.0251	0.0248	0.0264	0.0265		
T8-T7+(tre-1mM)	0.0271	0.0269	0.0264	0.0262	0.0265	0.0266		
T9-T7+(tre-1.5mM)	0.0272	0.0270	0.0265	0.0262	0.0266	0.0268		
CD 5%		V	=0.00278, T=0	.00341, V×T=0.0	0835			

Table 8. Effect of trehalose on carotenoid (mg gm⁻¹ fresh weight) content of wheat genotypes under heat stress (8hrs) of $35\pm2^{\circ}$ C and $40\pm2^{\circ}$ C.

Tucctments	Genotypes							
Treatments	HD2967	PBW175	C306	PBW343	PBW621	PBW590		
T1-Control at 25°C	0.0296	0.0293	0.0294	0.0295	0.0294	0.0295		
T2-T1+(tre-1mM)	0.0296	0.0293	0.0294	0.0295	0.0294	0.0296		
T3-T1+(tre-1.5mM)	0.0297	0.0295	0.0295	0.0296	0.0295	0.0297		
T4- at 35°C	0.0261	0.0262	0.0263	0.0270	0.0271	0.0268		
T5-T4+(tre-1mM)	0.0261	0.0261	0.0263	0.0271	0.0272	0.0266		
T6-T4+(tre-1.5mM)	0.0262	0.0262	0.0264	0.0272	0.0273	0.0267		
T7- at 40°C	0.0223	0.0225	0.0221	0.0218	0.0221	0.0218		
T8-T7+(tre-1mM)	0.0223	0.0225	0.0222	0.0221	0.0221	0.0220		
T9-T7+(tre-1.5mM)	0.0224	0.0226	0.0223	0.0221	0.0222	0.0228		
CD 5%			V=0.00493, T=0	0.00604, V×T=0.0	01479			

stress) for longer duration i.e. 8 hrs showed maximum carotenoid content decrease in all the genotypes.

The carotenoid content in leaf blade, awn and lemma were much lower after heat stress treatment than the control (Xu Xiao-Ling *et al.*, 2001). In response to heat stress, chloroplasts in the mesophyll cells of grape plants became round in shape, the stroma lamellae became swollen and the content of vacuoles formed clumps, whilst the cristae were disrupted and mito-chondria became empty (Zhang *et al.*, 2005).

Conclusion

Presently, as the six wheat genotypes were subjected to moderate $(35\pm2^{\circ}C)$ and severe $(40\pm2^{\circ}C)$ heat stress conditions. It has been observed in all the genotypes that the heat stress resulted in loss of photosynthetic pigments. More chlorophyll and carotenoid loss occurred under the severe heat stress conditions as compared to control and moderate stress. The genotype HD2967 showed high chlorophyll content even under the severe heat stress for 8 hours of duration. On the other hand PBW 175 followed by HD2967 showed higher carotenoid content under severe heat stress for 8 hours duration. Now in the treatments with the exogenously applied trehalose, there were increase in chlorophyll (a, b and total) and carotenoid contents as the trehalose known to protect the biological membranes from degradation by stabilizing them. Thus, the present findings demonstrated that exogenous application of trehalose to wheat has considerable potential for maintenance of thyllakoid membrane stability (mechanism unknown) in order to maintain the chlorophyll and carotenoid content in all the six studied wheat seedlings growing under heat stress conditions.

REFERENCES

- Ahmad, J. U. and Hassan, M. A. (2011). Evaluation of seedling proline content of wheat genotypes in relation to heat tolerance. *Bangladesh Journal of Botany*. 40(1):17-22
- Altenbach, S. B., DuPont, F. M., Kothari, K. M., Chan, R., Johnson, E. L. and Lieu, D. (2003). Temperature, water and fertilizer influence the timing of key events during grain development in a US spring wheat. J Cereal Sci, 37: 9–20
- Benaroudj, N., Lee, D. H. and Goldberg, A. L. (2001). Trehalose accumulation during cellular stress protects cells and cellular proteins from damage by oxygen radicals. J BiolChem, 276: 24261–67
- Dupont, F. M., Hurkman, W. J., Vensel, W. H., Tanaka, C., Kothari, K. M., Chung, O. K. and Altenbach, S. B. (2006). Protein accumulation and compo- sition in wheat grains: Effects of mineral nutrients and high temperature. *Eur J Agron*, 25: 96–107
- FAO (2016). Food outlook biannual report on global food markets, Food and Agriculture Organization of United Nations
- Gautam, A., Prasand, S. V. S., Ambati, D., Agarwal, D, and

Jajoo, A. (2016). Performance of durum wheat genotypes under drought and terminal heat stress conditions in changing climatic conditions. *Res & Review: J Botanical Sciences*, 5:4-5

- Gomez, M. L. And Lluch, C. (2011). Trehalose and abiotic stress tolerance. *Abiotic stress responses in plants*, 24:253-65
- Hiscox, J. D. and Israelstam, G. F. (1979). A method for the extraction of chlorophyll from leaf tissue without maceration. *Can J Bot*, 57: 1332–34
- Khan, M. I., Mohammad, T., Subhan, F., Amin, M. and Shah, S. T. (2007). Agronomic evaluation of different bread wheat (*TriticumaestivumL.*) genotypes for terminal heat stress. *Pak J Bot*, 39(7):2415-25
- Khan, S. U., Din, J. U., Qayyum, A., Jan, N. E. And Jenks, M. A. (2015). Heat tolerance indicators in Pakistani wheat (*TriticumaestivumL.*) genotypes. *Acta Bot. Croat.*,74(1):109–21
- Komayama, K., Khatoon, M., Takenaka, D., Horie, J., Yamashita, A., Yoshioka, M., Nakayama, Y., Yoshida, M., Ohira, S., Morita, N., Velitchkova, M., Enami, I. and Yamamoto, Y. (2007). Quality control of photosystem II: cleavage and aggregation of heat-damaged D1 protein in spinach thylakoids. *Biochimbiophys Acta*, 1767: 838-46
- Kumar, R. R., Goswami, S., Sharma, S. K., Singh, K., Gadpayle, K. A. and Kumar, N. (2012). Protection against heat stress in wheat involves change in cell membrane stability, antioxidant enzyme, osmolyte, H2O2 and transcript of heat shock protein. *International Journal* of Plant Physiology. Biochemical.4(4):83-91
- Larkindale, J. and Huang, B. (2004). Changes of lipid composition and saturation level in leaves and roots for heat -stressed and heat acclimated creeping bentgrass (*Agrostis stolonifera*). Environ Exp Bot, 51: 57-67
- Losa, D. A. and Murata, N. (2004). Membrane fluidity and its roles in the perception of environmental signals. *Biochimbiophys Acta*, 1666:142-57
- NPCS (2012): Niir Project Consultancy Service A1 Books.co.in. Select and start your own industry. Pp 448
- Shirdelmoghanloo, H., Cozzolino, D., Lohraseb, I, and Collins, N. C. (2016). Truncation of grain filling in wheat (*Triticum aestivum*) triggered by brief heat stress during early filling: association with senescence responces and reduction in stem reserves. *Func Plant Bio* 43: 919-30
- Wahid, A., Gelani, S., Ashraf, M. and Foolad, M. R. (2007). Heat tolerance in plants: A overview. *Envron and Expt Bot*, 61: 199-223
- Wardlaw, I. F., Blumenthal, C., Larroque, O. and Wrigley, C. (2002). Contrasting effects of heat stress and heat shock on kernel weight and flour quality in wheat. *Funct Plant Biol*, 29:25–34
- Xu, X. L., Wang, Z. M. and Zhang, J. P. (2001). Effect of heat stress on photosynthetic characteristics of different green organs of winter wheat during grain filling stage. *Acta Botanica Sinica*, 43(6): 571-77
- Zhang, J. H., Huang, W. D., Liu, Y. P. and Pan, Q. H. (2005). Effects of temperature acclimation pretreatment on the ultrastructure of mesophyll cells in young grape plants (Vitis vinifera L. Cv. Jingxiu) under cross- temperature stresses. *J Integr. Plant Biol*, 47: 959-70