Biochemical composition of promising leaves genotypes of buckwheat grown in Himachal Pradesh

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Abstract: Buckwheat originated from China and being cultivated all over the world, and has become a prominent pseudocereal. Among the pseudocereals (amaranthus, buckwheat and quinoa), buckwheat plant is economically important primarily due to their carbohydrate and protein rich grains, short growth span; besides foliage being used as a green vegetable and commercial source of the glycoside rutin used in medicine. In the present study, an attempt was made to evaluate the biochemical constituents of nutritional and nutraceutical significance of fourteen promising leaves genotypes of common buckwheat grown in Sangla region by following standard procedures. Wide variations in moisture content, crude protein, fat, ash, crude fiber, carbohydrates, ascorbic acid, oxalate and in vitro protein digestibility were observed to range from 87.4 to 92.2 %, 22.4 to 30.4 %, 1.8 to 3.7 %, 10.6 to 15.4 %, 12.0 to 13.9 %, 34.8 to 42.4 %, 25.0 to 29.2 mg/100g, 1375 to 1390 mg/100g and 53.4 to 65.1 % in that order. The content of minerals such as potassium, phosphorus, calcium, magnesium, zinc, manganese and copper varied from 1767.5 to 2035.0 mg/100g, 808 to 910 mg/100g, 394 to 409 mg/100g, 232.0 to 248.2 mg/100g, 3.1 to 6.1 mg/100g, 20.4 to 29.8 mg/100g and 0.2 to 1.4 mg/100g respectively. Based on cumulative grading done in respect of nutritionally desirable quality i.e., protein, ash, crude fiber, carbohydrates, ascorbic acid, in vitro protein digestibility, calcium, phosphorus, iron and oxalate content, the genotype IC-32731 followed by Kullugangetri and VL-27 emerged out to be overall superior versatile cultivars for cultivation under dry temperate climate.

Keywords: Ascorbic acid, Buckwheat, Carbohydrates, Oxalates, Protein

INTRODUCTION

Buckwheat is fast-growing annual, dicotyledonous broad plant belonging to the family Polygonaceae, the “smartweed” family, also called “knotweed” or “buckwheat” family. That is why, the seed of buckwheat is rather a fruit than a grain (the botanical term is “achene”) and referred as a pseudocereal (Goncalves et al., 2016). Buckwheat (FagopyrumesculentumMoench) is a temperate zone plant which originated from Southwest China (Kreft and Germ, 2008). Currently China, Russian Federation, Kazakhstan and Ukraine are the leading producers of buckwheat. The leading continents producers are Europe and Asia (FAO STAT, 2013). In India, the crop is grown in Western Himalayan states from Jammu and Kashmir in west to Arunachal Pradesh in east. The hilly terrains of Himachal Pradesh represent several diverse eco-geographic pockets which are suitable for the cultivation of hardy crops like small millet type pseudo-cereals like amaranthus, buckwheat and chenopods. In Himachal Pradesh, common buckwheat is grown in Kinnaur, LahulSpiti and Sirmour districts. Since the crop is adapted to temperate climate; hence, Himachal Pradesh can play an important role in production of this crop. Buckwheat, also called rhubarb or sorrel, is not a cereal but is usually grouped with cereals due to its ways of cultivation and utilization. This herbaceous erect, annual, melferos plant is commonly known as kathu, fafra, ogle or bharesh. This pseudo-cereal crop is grown in higher hills and mountains of the state at an altitude of >1500 to 4500 msl and covers about 1500 ha area (Phogat and Sharma, 2000).

The vegetable greens are extremely beneficial to the human health and form wonderful source of energy and vitality that powers life on earth. Greens contain a fair amount of good quality protein and are an excellent source of dietary fibre. Besides, freshly picked raw greens are rich source of vitamin C and nutritionally important minerals, which perform important metabolic functions in the body (Dogra, 2010). In literature, there is scanty information on the nutritional composition of buckwheat leaves. The present study was aimed to identify nutritionally superior leaves genotype of common buckwheat specially with protein and ascorbic acid content which may be useful for further crop improvement programme.
MATERIALS AND METHODS

The leaves samples (200 g) of 14 varieties/genotypes of common buckwheat namely S-B-201, S-B-212, S-B-214, VHC-27, VL-27, Kullugangetri, IC-323731, Hassoska, Emka, PRB-9001, USDA-1, VL-7, PRB-1 and OC-2 were collected from Mountain Agriculture Research and Extension Centre, Sangla (HP) and evaluated for various biochemical parameters. The finely ground leaves samples of these genotypes were analyzed in triplicate for moisture, crude protein, ash, crude fibre and crude fat by AOAC method (1970) and carbohydrate as per Gopalan et al. (2004). The samples were ground and digested in di-acid mixture (nitric acid: perchloric acid in 3:1 v/v) for the determination of mineral elements namely Mg, Ca, Fe and Zn on atomic absorption spectrophotometer (model Perkin Elmer, USA) as per Piper (1966). Potassium was determined by flame photometer [model-129; Systronics (India)] whereas phosphorus was determined colorimetrically in the digested samples as per Olsen (1954). The data was analyzed statistically by using analysis of variance (Panse and Sukhatme, 1984). In order to identify multipurpose versatile cultivars of buckwheat, rating (ranking) of various genotypes was done based on nutritionally desirable traits i.e., protein, carbohydrates, calcium, iron content as adopted by Dogra (2010).

RESULTS AND DISCUSSION

The biochemical analysis of buckwheat leaves of various genotypes revealed that moisture content in genotypes varied from 87.4 to 92.2 % with maximum in genotype OC-2 and lowest in Kullugangetri (Fig.1). The moisture content roughly reflects the degree of maturity and accumulation of fibre in the crop plants. The protein content varied significantly from 22.4 to 30.4 % with highest content in genotype S-B-212, followed by genotypes IC-323731 and Hassoska and lowest in genotype OC-2. The crude fat, an estimate of

Table 1. Varietal grading of buckwheat leaves varieties / genotypes in search of versatile /multipurpose genotypes.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>IC-323731</th>
<th>Hassoska</th>
<th>VHC-27</th>
<th>VL-27</th>
<th>Kullugangetri</th>
<th>Emka</th>
<th>USDA-1</th>
<th>VL-7</th>
<th>PRB-1</th>
<th>OC-2</th>
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<tbody>
<tr>
<td>Protein*</td>
<td>10</td>
<td>14</td>
<td>12</td>
<td>15</td>
<td>15</td>
<td>12</td>
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<tr>
<td>Crude Fibre*</td>
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<td>12</td>
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<td>12</td>
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<tr>
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<td>Oxalate*</td>
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<td>3</td>
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<td>Iron*</td>
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<td>Cumulative</td>
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Fig. 1. Variation in biochemical composition of promising genotypes of the buckwheat leaves (n=3)(Values on (%) dry wt.).
the total fat content, was highest in genotype USDA-1 and lowest in S-B-214. The genotype VHC-27 had highest ash content while Hassoska had lowest ash content with overall range 10.6 to 15.4 %. The crude fibre content in genotypes varied from 12 to 13.9 % with maximum in genotype OC-2 and minimum in S-B-212. The carbohydrates form a major part of food which helps in building body strength by generating energy. The carbohydrate content in genotypes varied from 34.8 to 42.4 % with maximum in S-B-214 and minimum in S-B-212. Our results are in agreement with Dietrych-Szostak and Ploszynski (1988) who reported 8.3 per cent ash, 10.1 per cent fiber, 3.5 per cent fat and 13.5 per cent protein on dry weight basis in the leaves of buckwheat plants. However, Lahanov et al. (2004) reported 24.3 per cent crude protein and 14.1 per cent ash content in leaves of Fagopyrum esculentum ssp. esculentum. Gopalan et al. (2004) reported that spinach leaves contained 92.1 per cent moisture content. Leiber et al. (2012) observed crude protein and ether extract in buckwheat leaves 242 and 32 g/kg dry matter basis. Vojtiskova et al. (2012) reported that moisture, crude protein and fat content in buckwheat leaves 7.5, 22.7 and 3.1 %, accordingly. Variation noticed in these parameters by and large in accordance with the values reported by with Dietrych-Szostak and Ploszynski (1986) and Farooq and Tahir (1989). The status of ascorbic acid content in buckwheat varieties / genotypes was evaluated and pertinent data are presented in Fig.2. The overall variation in this parameter was observed from 25.0 to 29.2 per cent among various varieties / genotypes. The genotype PRB 9001 showed the highest ascorbic acid content followed by OC-2, PRB-1, S-B-214, VL-27, IC-323731, Emka, Hassoska and S-B-212 (statistically at par). However, the lowest value was noticed in the genotype S-B-201. The perusal of literature revealed variation in ascorbic acid content from 10 to 117 mg/100g in fresh spinach leaves (Siener et al., 2006 and Gopalan et al., 2004).

The data showing variability in the in vitro protein digestibility in the leaves of different buckwheat varieties / genotypes is presented in Fig. 2. Significant variation in respect of this parameter among various varieties / genotypes was observed to range from 53.4 to 65.1 per cent. The genotypes Hassoska showed the highest value, whereas the genotypes OC-2, Emka and PRB-9001 possessed the next higher value on this aspect. However, the lowest value was observed in the variety PRB-1. with Dietrych-Szostak and Ploszynski (1986) reported 47.5 per cent in vitro digestibility in leaves and stemsof buckwheat plant residues and hulls. Ly and Preston (2001) have reported 68.8 per cent in vitro digestibility in Ipomoea aquatica. Low protein digestibility due to high crude fibre content and tannin content (Eggum et al., 1981). The extent of variation observed for in vitro protein digestibility of the present study are in conformity with the findings of above investigators on this aspect.

The status of oxalate content in the leaves of different buckwheat varieties / genotypes was evaluated and pertinent data on this aspect are depicted in Fig. 2. Significant variation in oxalate content of different buckwheat genotypes was observed to range from 1375 to 1390 mg/100g. Among different varieties / genotypes the lowest oxalate content was found in VHC-27, IC-323731 and VL-27. However, the highest value for oxalate content was noticed in the genotypes S-B-212 followed by VL-7. The released varieties PRB-9001, USDA-1, PRB-1 and OC-2 exhibited 1379, 1379, 1380 and 1380 mg/100g oxalate content, respectively. Siener et al.(2006) reported the oxalate content as 1391 mg/100g in raw leaves of Sorrel Rumex acetosa L. var. hortensis belonging to Polygonaceae family, whereas soluble oxalate content ranged from 800 to 1257 mg/100g and total oxalate content ranged from 1634 to 2285 mg/100g in raw leaves of spinach (Spinaciaoleracea), respectively and the findings of the accompanying work fall within this range of varia-
tion of these vegetable greens. The mineral analysis of buckwheat genotypes revealed that potassium content ranged from 1767.5 to 2035 mg 100 g-1 with lowest and highest values in genotype PRB-1 and USDA-1, respectively (Fig.3). Phosphorus content varied from 808 (in OC-2) to 910 (USDA-1); magnesium content from 232 (VL-27) to 248.2 (VHC-1); manganese content from 20.4 (Hassoska) to 27); calcium content from 394 (VL-27) to 409 (ICm); magnesium content from 232 (VL-27) to 248.2 (VHC-1); and Copper content from 0.2 (VL-27) to 1.4 (S-B-201); mg 100 g-1 respectively. Ikeda et al. (1999) analyzed calcium, magnesium, phosphorus, potassium, calcium, phosphorus and iron contents is presented in Table 1. The genotypes IC-323731 proved to be prominent genotypes followed by Kullugangetri, VL-27, S-B-214 and USDA-1 in order of cumulative ranking. The genotypic grading made it possible to identify multipurpose genotypes suited for their direct inclusion in the human diet.

Conclusion

Buckwheat is a plant from the family of Polygonaceae. Buckwheat leaves contains protein (30.4 %), carbohydrates (42.4 %), ascorbic acid (29.2 mg/100g ), oxalate (1390 mg/100g), potassium (2035 mg/100g), phosphorus (910 mg/100g), calcium (409mg/100g), magnesium (248.2 mg/100g) and zinc (6.1 mg/100g). Based on cumulative grading done in respect of nutritionally desirable quality, the genotype IC-323731 followed by Kullugangetri and VL-27 emerged out to be overall superior versatile cultivars for cultivation under dry temperate climate.

REFERENCES


